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Assessment of dermal absorption of aluminium from a representative antiperspirant formulation using a (²⁶Al)Al microtracer approach: a follow-up study in humans

Rianne de Ligt 🔟^{1,*}, Joost Westerhout², Dimitri Grossouw¹, Thomas P. Buters³, Robert Rissmann³, Jacobus Burggraaf³,

Albert D. Windhorst⁴, Sarah Tozer⁵, Gerlinde Pappa⁶, Brian Wall⁷, Dagmar Bury⁸, David R. Mason⁹, Wouter H.J. Vaes¹

- ¹TNO, P.O. Box 2215, 2301CE Leiden, The Netherlands,
- ²TNO, P.O. Box 80015, 3508TA Utrecht, The Netherlands,
- ³Center for Human Drug Research, 2333CL Leiden, The Netherlands,
- ⁴Department of Radiology and Nuclear Medicine, Free University Medical Center, P.O. Box 7057, 1007MB, Amsterdam, The Netherlands,

- ⁶Beiersdorf AG, Unnastrasse 48, 20245 Hamburg, Germany,
- ⁷Colgate Palmolive Company, 909 River Road, Piscataway, NJ 08855, USA,
- ⁸L'Oréal Research & Innovation, 9 rue Pierre Dreyfus, 92110 Clichy, France,
- ⁹Safety and Environmental Assurance Centre, Unilever, Colworth Science Park, Sharnbrook MK44 1LQ, UK

*Corresponding author: Rianne de Ligt, TNO, P.O. Box 2215, 2301CE Leiden, The Netherlands. Email: Rianne.deligt@tno.nl

A follow-up study was performed in 12 healthy women to evaluate systemic exposure to aluminium following topical application of a representative antiperspirant formulation under real-life use conditions (part A) and to assess the local fate of topically applied aluminium by taking additional tape strips and skin biopsies (Part B). A simple roll-on formulation, containing the maximal possible radioactive dose, was prepared with [²⁶Al] aluminium-labeled chlorohydrate (ACH). The microtracer of [²⁶Al] was used to distinguish aluminium from the natural background, using accelerator mass spectrometry. [²⁶Al] aluminiumcitrate was administered intravenously to estimate the dermal fraction absorbed. Despite the 25-fold increase of the topical dose compared with the previous study, only 12 blood samples gave results above the lower limit of quantitation (0.118 fg/mL). The most reliable estimates of the dermal fraction absorbed are derived from noncompartmental analysis with the urine data. By using the intravenous dose to normalize the urinary excretion to 100% bioavailability, the best estimate of the fraction absorbed of [²⁶Al] from a topical application of [²⁶Al] aluminium-labeled chlorohydrate in an antiperspirant formulation was 0.00052%. Part B of the study demonstrated that the majority of the aluminium in the formulation remained associated with the external layers of the skin without penetration through the skin.

Key words: microtracer research; accelerator mass spectrometry; dermal bioavailability of aluminium.

Introduction

Aluminium (Al) is a commonly occurring metal in the earth's crust and is naturally present in water and agricultural products. Humans are exposed to Al through food, drinking water, pharmaceuticals, and cosmetic products. Al salts, such as Al chlorohydrate (ACH), are widely used as antiperspirants and as treatment for hyperhidrosis.¹ In 2012, Joint FAO/WHO Expert Committee on Food Additives² established a provisional tolerable weekly intake of 2 mg/kg bw/week, based on a pivotal 12-month oral rat study that included a multigenerational and a developmental toxicity study with aluminium citrate.³ Regulatory review in Europe^{4,5} revealed no conclusive evidence of Al playing a role in cancer^{6,7} and neurodegenerative disorders. Previous studies have shown that systemic exposure following

dermal Al exposure is so low that sensitive analytical techniques such as accelerator mass spectrometry (AMS) are required.^{8,9} To enable robust quantitative risk assessment, EU authorities requested an accurate measurement of the skin penetration of Al from antiperspirant use.⁴

Therefore, an absolute bioavailability study was performed with 12 healthy women evaluating systemic exposure to Al following topical application of a representative antiperspirant formulation under real-life use conditions, including single and repeated dosing and shaving of the axillae.⁹ A [²⁶Al] microtracer was used to distinguish Al dosed from natural background. [²⁶Al]-Al-citrate was administered intravenously (IV) to estimate fraction absorbed (F_{abs}).⁹ Following topical application, only 2 blood samples were just above

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⁵Procter & Gamble Technical Centres Ltd, Reading RG2 0QE, UK,

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the lower limit of quantitation (LLOQ; 0.12 fg/mL). From urinary excretion data, a conservative mean $F_{\rm abs}$ (0.0094%) was estimated, using the half LLOQ-based method.¹⁰ No apparent difference between the various use conditions (single and repeated dosing as well as shaving) was observed.⁹ Having reviewed this study, the SCCS requested further experimental work to address residual data gaps, particularly referring to the local fate of Al and the ability to determine an $F_{\rm abs}$ value.

This follow-up study consisted of 2 parts (Part A and Part B), each with 6 females (12 females in total). Part A included additional features compared with the previous study:⁹ (i) increased proportion of radiolabel (~25fold) incorporated into the dermal dose to improve the chance to quantify absorption, (ii) collection of total urine throughout the first 24 h up to Day 11 to improve the estimates of Al excreted in urine, (iii) collection of feces until Day 11 to enrich data on recovery and excretion, (iv) analysis of [²⁶Al] on protective gauzes, T-shirts, and washes to recover as much of the applied dose as possible, and (v) tape stripping and skin biopsies.⁹ The subjects also received an IV dose containing [²⁶Al] at Day 36 to determine the absolute dermal bioavailability analogous to our earlier study.9 Part B, conducted in a separate cohort of 6 females, included tape stripping and skin biopsies at different time-points to obtain valuable information on the fate of the topically applied Al. This investigation was performed separately in order not to compromise the real-life consumer exposure scenario in Part A. Although the aluminium concentration in the test formulation and amount applied was identical for the cohort in Part B, the proportion of radiolabel within the test formulation could be reduced to 1 Bq [compared with part A (2500 Bq)] without losing analytical sensitivity. All 12 subjects received a single dermal dose after 2 weeks of daily shaving and use of a marketed Al-containing antiperspirant.

Materials and methods Production and analysis of [²⁶Al]-labeled dose formulations

The full worldwide stock of purified [²⁶Al]Al isotope was purchased from Los Alamos National Laboratory, USA. For Part A, the [²⁶Al]-labeled ACH was prepared and incorporated into an antiperspirant formulation as previously described,⁹ including further downsizing of the batch and increasing dose (2,500 Bq versus ~100 Bq⁹). In short, aluminium powder, aluminium chloride solution, [²⁶Al]Al (concentrated by evaporation), and water were mixed and heated to initiate the reaction. The [²⁶Al]-labeled ACH used in this study was comparable to specifications for commercially available antiperspirant actives used in marketed products. For part B of the study, this material was diluted 2,500× with commercial ACH (Elementis, USA) to obtain the required dose of ~1 Bq.

The solution for intravenous (IV) administration was prepared according to the Principles of Good

Manufacturing Practice (GMP) at the GMP hotlab of the department of Radiology & Nuclear Medicine of the Free University Medical Center (Amsterdam, The Netherlands).⁹ The procedures were fully described in the Investigational Medicinal Product Dossier, which was approved by the ethical committee prior to administration. For logistic reasons, 2 batches of IV solution were manufactured.

Details of the dose formulations are given in Table S1 (see online supplementary material for a color version of this table).

Sample analysis

All samples were analyzed for [²⁶Al] content using AMS. The AMS methods⁹ were (re)qualified for the analysis of blood, urine, fecal homogenates, and extracts in 0.1 M HCl. The methods were accepted as they fulfilled the requirements for linearity, accuracy, and precision. Each analytical batch consisted of study samples together with 6 calibration samples in duplicate and 3 QC samples, analyzed in triplicate.

Urine samples were analyzed for ²⁷Al content by Inductively Coupled Plasma Mass Spectrometry (ICP-MS) using Rhodium (m/z 103) as the internal standard.⁹ The analytical method was qualified for selectivity, linearity, accuracy, precision, and LLOQ.

Data analysis

 26 Al/ 27 Al isotope ratios of the samples were converted to mBq/mL by plotting these on the linear calibration line, using a weighing factor or $1/x^2$. Concentrations in mBq/mL were then converted to fg Aluminium/mL based on the specific activity for [26 Al]Al of 722 Bq/µg. Blood concentration-time area under the curves (AUCs) were determined using the trapezoid method (linear up, loglinear down method).⁹ AUC_{0:672} for each subject was used to calculate the dermal bioavailability (F_{abs}) as the ratio of AUC dermal to IV, then averaged over subjects.

Study design and subject characteristics

The study was designed as a single-centre, openlabel, 2-period, fixed sequence study, conducted in the Netherlands (Center for Human Drug Research, Leiden). Twelve healthy women (18–43 years of age, body mass index within 18.1–27.3 kg/m², 6 in each cohort) were included. Subjects were familiar with frequent wet shaving using an appropriate female safety razor. Exclusion criteria included clinically significant abnormality of the axilla (e.g. scars, tattoos, and/or dermal abnormalities); use of Al-containing medications; and axillary hyperhidrosis. Both parts of the study were approved by the Medical Ethics Review Board Brabant (The Netherlands); informed consent was obtained from all subjects. A schematic overview of the study design can be found in Fig. 1.

Two treatment periods were included (Part A): 1 topical application (\sim 2,500 Bq) and 1 IV administration (\sim 0.1 Bq).

Part A	Part B
SJ001 – SJ006	SJ007 – SJ012
Adaptation (2 weeks):	Adaptation (2 weeks):
daily use of antiperspirant and daily shaving	daily use of antiperspirant and daily shaving
Treatment 1: Topical application of ²⁶ Al in a representative antiperspirant formulation (2500 Bq) After 24h: skin wash After 48h: skin wash and discharge from clinic	Treatment 3: Topical application of ²⁶ Al in a representative antiperspirant formulation (1 Bq)
Sample collection (4 weeks):	Tape stripping after:
10 day 24h urine and feces (up to 240h)	20 min (drying of formulation only)
24h urine collection 336h, 504h and 672h	1h (semi-occlusion)
Tape stripping 168h	6h (semi-occlusion)
daily use of antiperspirant after skin wash at 48h	24h (semi-occlusion), followed by skin biopsy
Tape stripping and skin biopsy 840h Treatment 2: IV administration of 26Al as a bolus injection 5 mL (0.1 Bq)	
Sample collection (4 weeks): 10 day 24h urine (up to 240h) 24h urine collection 336h, 504h and 672h	

Fig. 1. Study design.

Part B consisted of a single topical application (\sim 1 Bq). Each topical application was preceded by an adaptation period of 2 weeks [daily shaving of the armpits and application of antiperspirant product with Al (as ACH)]. The [²⁶Al]-labeled formulation was applied to both axillae within a delineated area of $\sim 100 \text{ cm}^2$ per armpit. After the formulation had dried (~15-20 min), an air permeable gauze was applied over the axillae and a cotton Tshirt was worn to avoid loss of radiolabel to the environment, and 24 and 48 h after application, the axillae were washed using cotton gauze and a mild soap solution. The axillae were covered in-between with a fresh gauze and T-shirt. Subjects were released from the unit after the 48-h washing procedure. All T-shirts, gauzes, and washings were collected for AMS. Blood samples were collected at various time points up to 28 days after application. All urine and feces were collected up to 240 h postdose. Additionally, selected urine was collected up to 28 days postdose. On Day 8 (168 h postdose), the skin in the axillae was stripped (D-squame [®] discs, CuDerm, USA) until the shiny interface of the viable epidermis became visible. On Day 36 (840 h postdose), the procedure was conducted in the opposite armpit. After stripping, a skin biopsy (3 mm) was taken within the stripped area. After the skin biopsy was taken, the IV solution was administered as a bolus injection of 5 mL into a suitable antecubital vein. Blood and urine samples were collected up to 28 days after IV administration.

In part B, the armpits were divided into 4 designated areas for tape stripping (just left and right of the middle part). The formulation (~1 Bq) was applied and allowed to dry for 20 min. Thereafter, one designated area was stripped until the shiny interface of the viable epidermis became visible. After stripping, an air permeable gauze was applied over both axillae and the subjects wore T-shirts. After 1, 6, and 24 h following dermal application, tape strips were taken from the remaining spots. At 24 h, a skin biopsy (3 mm) was taken within the stripped area.

Results

No clinically relevant changes in vital signs, laboratory, or ECG measures were observed throughout the study. All subjects completed the study without any relevant complaint and topical treatments were well tolerated. Mild discomfort was commonly observed upon IV administration of the [²⁶Al]-labeled solution; 1 subject experienced moderate discomfort, most likely due to subcutaneous (mis)dosing. A total number of 32 adverse events (AEs) were reported during the study period (Table S2, see online supplementary material for a color version of this table). All AEs considered to be possibly or probably related to study treatment (including the semi-occlusive conditions) were of moderate (n = 2) or mild severity (n = 10, Table S2, see online supplementary material for



Fig. 2. Part A: $[^{26}$ Al] concentrations in whole blood after IV injection (zoom 0–12h).

a color version of this table). No serious AEs occurred in this study.

In Part A, 186 blood samples were analyzed for ^{[26}Al] content. Albeit the topical dose of ^{[26}Al]-ACH was increased 25-fold compared with the previous study,⁹ only 12 whole blood samples (from 2 subjects) contained a concentration above the LLOQ of 0.118 fg/mL. These concentrations were well below 1 fg/mL and thus very low compared with the nominal dose given (~3.73 μ g [²⁶Al]Al). After IV administration of 0.1 Bq (~120 pg [²⁶Al]Al), [²⁶Al] could be detected in blood up to 72 h. The analysis for the IV administration is based on only 5 subjects due to the inadvertent mis-dosing of one subject, who showed a clearly different PK profile, resembling intramuscular or subcutaneous injection (Fig. 2). All values below LLOQ were replaced with the LLOQ value to provide conservative estimates of exposure. This approach resulted in a calculated averaged percentage absorbed of less than 0.0021% (n = 5), principally driven by samples that were below LLOQ.

The entire urine output was collected up to 240 h postdose (24-h intervals); the first 24 h were divided into 0-6, 6-12, and 12-24 h collections. In urine samples (192 in total), [²⁶Al] was detected up to 144 h (topical application) or up to 672 h (IV administration). The cumulative excretion of [²⁶Al] in urine from both routes is presented in Fig. 3; the urinary excretion represents 0.00026% of the topical dose applied and 68.8% of the IV dose. The dermal fraction absorbed was calculated by the ratio of the total fraction excreted in urine following the topical dose to the total fraction excreted following the IV dose. Using the same conservative strategy of replacing values <LLOQ with LLOQ.⁹ the average fraction absorbed was found to be at least smaller than 0.00052% (n = 5). This means that of 200,000 Al atoms applied on the skin, only maximally 1 atom was actually absorbed.

All urine samples were also analyzed for the total amount of Al present ($[^{27}Al]$ and $[^{26}Al]$) using ICP-MS and found vary day-to-day within normal ranges.¹¹⁻¹³ With these results and taking into account the urinary



Fig. 3. Part A: Cumulative urinary excretion of [²⁶Al] after topical application (open triangles, 2500 Bq) or IV injection (black circles, 0.1 Bq).

output, the total urinary excretion of Al [from all possible routes (oral, inhalation, dermal)] was calculated and is presented in Fig. 4 (open symbols). To assess the contribution of antiperspirant use to this total excretion of Al in urine, the following approach was taken. First, the ratio between [²⁶Al] and Al was calculated. The amount of Al in the dose was calculated to be 0.9166 g Al; the amount of [²⁶Al] in the dose was calculated to be 25.55 μ g, resulting in a [²⁶Al]: Al ratio of 1: 35,868. Since absorption of both isotopes in the antiperspirant formulation would be the same, this ratio was used to convert the radiolabel in the urine, measured as [²⁶Al]/mL (i.e. pg/L) to the equivalent μg total Al/L (Fig. 4, closed symbols). Importantly, since antiperspirants may be used on a daily basis, one might expect that exposure on preceding days would also contribute to aluminium excretion; therefore, to account for the fact that [²⁶Al] was administered only once, the exposure from [²⁶Al] was extrapolated to a multiple dose by adding up the amount of each preceding day (values below LLOQ were set to ½ LLOQ, i.e. representing 0.0020 μ g/L) and this is presented by the solid line in Fig. 4. When comparing the solid line with the open symbols in Fig. 4, it can be concluded that, even in the daily use scenario, only a minor part of the Al excreted in urine originally derived from the topically administered antiperspirant formulation.

In this study, every practical effort was taken to recover as much of the [²⁶Al] dosed as possible by collecting: skin wash samples (containing gauzes, cloths, and razor blades) and the T-shirts worn by the subjects. The average recovery from these samples was 70% (59–77%, Table 1). Recovery in remaining samples (urine, fecal homogenates, skin biopsies, and tape strips) represented 0.021% (Table 1). In fecal samples between 24 and 240 h after dermal application, Al was detected at low levels (mean: 0.0017%, Table 1). The amount of [²⁶Al] recovered from the tape strips after 168 and 840 h was rather similar.

The fate of [²⁶Al] on the skin and within the stratum corneum was studied in more detail by collecting tape strips from the axilla at 4 different time points (Fig. 5,



Fig. 4. Al excretion in microgram for all subjects (black circles Al derived from antiperspirant (AP), open circles total Al excretion, solid line cumulative Al excretion from AP).

Table 1. Overview of average % of dose recovered in all study samples in part A (n = 6).

Sample	Recovery in % of dose (range)
Skin wash 24 h	62.0 (54.1–73.6)
T-shirt 24 h	6.0 (1.1–14.6)
Skin wash 48 h	1.6 (0.8–3.0)
T-shirt 48 h	0.09 (0.07–0.15)
Subtotal (non-absorbed dose)	69.7 (58.7–76.7)
Urine	0.0003 (0.0001–0.0007)
Feces	0.0017 (0.0008–0.0057)
Skin biopsy (840 h, $n = 2$, remaining <lloq (<math="">n = 4))</lloq>	0.00003-0.00004
Tape strips (168 h)	0.0097 (0.0019–0.0417)
Tape strips (840 h)	0.0090 (0.00004–0.0525)
Subtotal (potential absorbed dose)	0.021 (0.004–0.095)

Table S3, see online supplementary material for a color version of this table). At 20 min, most of the recovered dose was found in the outer tape strip. The percentage

of the applied dose decreased substantially with each sequential tape strip. This does not necessarily represent formulation that is within the stratum corneum, since



Fig. 5. Part B: Representation of the average amount $[^{26}\mbox{Al}]$ (in fg) recovered from tape strips.

these tape strips may still contain formulation sticking in the troughs/furrows of the heavily crenulated axilla skin. By 24 h, the amount recovered decreased to less than 2% of the normalized dose applied. Importantly, the profile within the stratum corneum did not change over time; the majority of the recovered Al was in the initial tape strip and diminished rapidly with each strip. In skin biopsies (taken at 24 h), 0.08% of the dose applied was recovered (n = 4, 2 samples were found to be <LLOQ). All results were normalized for the total amount of [²⁶Al] recovered after 20 min of application, since this first tape strip contained more than 100% of the applied dose in all subjects. This was likely due to the uneven surface of the axillae which prevented homogeneous distribution within the designated area. The first time-point, where the axilla was undisturbed, should best reflect the maximum dose applied.

Discussion and conclusion

This follow-up study (Part A) on the assessment of dermal absorption of Al from a representative antiperspirant formulation under real-life conditions represents a refinement of our previous study⁹ and supports a more precise quantitative assessment of consumer exposure. Part B, performed separately, not to compromise the reallife consumer exposure scenario of Part A, was conducted to generate more detailed data on the fate of [²⁶Al] after dermal application to strengthen the limited "mass balance" data generated in part A.

Previously, no apparent difference was observed between the various treatments (shaving, single, or daily use).⁹ Thus, this new investigation was simplified to a single treatment (daily shaving and product use). Several improvements were included in the study design to enhance data interpretation, such as increased proportion of radiolabel in the topical dose, collection of total urinary output, feces, and "mass balance"-like samples. A detailed list of the differences in the design of both studies is given in Table 2. It should be noted that both studies have been conducted in the Caucasian population only. While no people of color (PoC) have been included in the study, at a mechanistic level, this would not be expected to impact on the results. Antiperspirants function by interacting with bicarbonate buffered sweat and ductal mucins to temporarily occlude sweat ducts. This mechanism of action relies on skin physiology that is not significantly different between Caucasian and PoC, including pH on the skin surface and within the skin. The major difference between Caucasian and PoC skin resides in the pigmentation; however, it would not be expected to alter the action of aluminium in sweat ducts.

In theory, blood concentrations would provide an absolute bioavailability value. However, despite a maximal increase of the dermal radioactive dose (25-fold using the full worldwide stock) and the use of the most sensitive AMS techniques, the majority of measures in blood remained below LLOQ and the dermal AUC could not be derived using noncompartmental analysis. Instead, a very conservative scenario was applied by replacing all values below LLOQ with the LLOQ value, which resulted in an estimated average dermal bioavailability of ~0.0021%, principally driven by samples below LLOQ. This estimate was markedly lower when compared with our initial study (0.0116%, upper estimate, average of all treatments), representing a substantial refinement to the conservative approach taken previously.⁹ To strengthen this estimate of \sim 0.0021%, we focused on the urinary excretion following IV and dermal dosing.

As the topical dose had 25-fold more radioactivity compared with our first study,⁹ more urine samples were above LLOQ. Also, the measurement of urinary Al excretion was considerably refined by collecting 3 samples within the first 24 h and the complete 24 h urine output for the first 10 days. Again, values below LLOQ were replaced with LLOQ, resulting in a conservative mean estimated fraction absorbed of 0.00052% (0.00026–0.00108%). These results confirm that our previous approach using the half LLOQ-based method¹⁰ provided a conservative estimate (average fraction absorbed was 0.0094%⁹). Since estimates of the fraction absorbed are driven by LLOQ, the 25-fold increase in the topical dose (sensitivity) resulted in a comparable 20-fold reduction in estimated dose.

The low Al levels observed in fecal samples between 24 and 240 h were unexpected since the elimination via feces is very unlikely.¹⁴ Thus, these low levels were considered secondary to contamination, possibly by small quantities dropping from the T-shirt or being ingested following hand to mouth contact. They seem an artifact that should be interpreted with caution and no comparable samples from the IV dose are available. However, these would not account for environmental cross contamination anyway. Albeit fecal excretion is unlikely, this assumption represents an uncertainty in the exposure calculation.

Achieving a good mass balance of Al is challenging,⁸ particularly without disturbing the desired real-life consumer scenario by strict measures such as occlusion. This study was not designed to be a classical mass

	De Ligt, 2018 ⁹	Current study Part A
Number of subjects	12	6
Dose	100 Bq [²⁶ Al]-ACH in a representative topical formulation	2500 Bq [²⁶ Al]-ACH in a representative topical formulation
Application site	Both axillae (50 Bq [²⁶ Al] each)	Both axillae (1250 Bq [²⁶ Al] each)
Antiperspirant use	Single and repeated*	Repeated
Application details	Nonocclusion: subjects were wearing non-occlusive T-shirts during the first 24 h and to avoid loss to the environment	Semi-occlusion: the application site was covered with gauzes loosely attached under the arms and subjects were wearing non-occlusive T-shirts during the first 48 h and to minimize loss to the environment
Shaving regimen	Adaptation period of 4 weeks with either daily wet shaving ^a or no shaving at all	Adaptation period of 2 weeks with daily wet shaving ^a
Urine collection	Morning spot urine samples (24, 48, 72, 168, 336, 504, and 672 h)	2- h interval urine samples (0–6, 6–12, 12–24, and 24 h intervals up to 216–240, 312–336, 480–504, and 648–672 h)
Feces collection Samples to assess the local fate of [²⁶ Al]	Not done Not done	24-h intervals for 10 days Skin wash (including gauzes) and T-shirt samples (24 and 48 h), tape strips (at 168 and 840 h), skin biopsy (at 840 h)

Table 2. Differences in study design between de Ligt (2018,⁹) and Part A of the current study.

^aLast shaving was performed on the morning prior to [²⁶Al]-ACH application at the clinical site. ^bDosing after adaptation period without antiperspirants considered to represent a single dose of ACH and dosing after adaptation period with daily use of antiperspirants considered to represent repeated dosing.

balance study, since the necessary occlusion would have created an entirely artificial exposure scenario, not resembling real-life consumer use conditions. Besides, absolute bioavailability studies are the first choice study design for the estimation of internal exposure, while mass balance studies are assessing excretion (routes). As demonstrated by Flarend *et al.*,⁸ attempts to occlude/tape-strip the axillae of subjects results in rapid damage of the axilla skin, which impacts skin absorption and the harm caused to the subjects is considered to be unethical. Here, the application sites were semioccluded, which is why a significant amount of [²⁶Al] was found in all T-shirts collected after 24 h.

Overall recovery of the topically applied [²⁶Al] in this study was ~70% and significantly greater compared with the previously published study, where recovery was below 50%.⁸ Recovery was predominantly in samples considered as nonabsorbed: skin washes and T-shirts. Based on the significant amount of [²⁶Al] on the T-shirt, loss to the environment is the most plausible explanation for the proportion not accounted for. This is further supported by the Al found in the 48 h samples (skin wash and T-shirt), which, after 2 days, still contained measurable amounts of Al (more than 1% of the dose applied). Also, all control tape strip samples taken from the upper back at 168 h (part A) or 24 h (part B) after application contained measurable levels of [²⁶Al], demonstrating that transfer occurred to other areas of the body via contamination of the environment. It is clear that the mean recovery from the biological samplesskin biopsy, tape strips, urine, and feces together-is only 0.021% and this shows that an extremely small amount crosses the skin barrier (Table 1). Moreover, the amount of [²⁶Al] recovered from the tape strips after 168 and 840 h was rather similar, which indicates that the [²⁶Al] remained on the skin surface and did not penetrate to lower skin layers. This is further supported by the results of part B, which demonstrate that most of the formulation remained external on the skin surface (Fig. 5). Virtually, all the radioactivity was removed in the first few tape strips, indicating that the applied labeled substance was predominantly associated with external layers of the skin without absorption. Also, the similarity of the tape strip profiles at the various time points shows no evidence of inward distribution *within* the stratum corneum and lower skin layers over the time-course of part B, as one might expect for substances that penetrate into the skin.¹⁵

Our data are consistent with the hypothesis that the soluble Al salts form insoluble gels of Al hydroxide at physiological pH on the skin surface; then, the insoluble Al precipitate forms superficial plugs in the openings of sweat ducts^{16–18} and strongly associates with proteins on the surface of the stratum corneum. These temporary plugs are lost from the skin surface through natural sloughing of the stratum corneum. In reality, loss of the antiperspirant formulation to the environment is expected to be even greater. To have a controlled, yet conservative, exposure scenario, we used semi-occlusive gauze and a standard T-shirt 20 min after application. However, in normal life, consumers would likely dress soon after applying antiperspirant, which would remove some of the freshly applied formulation from the surface of the axilla. Furthermore, the presence of the gauze would have protected the test formulation from the gentle abrasion of the fabric on the skin which might further dislodge adherent formulation.

A question raised regarding the interpretation of the first study⁹ was the potential difference in excretion kinetics of the Al species immediately after IV dosing with Al (in citrate-buffered saline), which might be subtly different to the kinetics of the topically applied

formulation (with ACH). Normally, approximately 10-20% of Al in plasma is bound to citrate and is therefore potentially more accessible for renal filtration, whereas 80–90% of Al in the plasma is bound to transferrin and unfilterable.¹⁹ There is an equilibrium between the amount of Al bound to plasma citrate and the amount of Al bound to plasma transferrin; the kinetics of this equilibrium are known to be relatively rapid.²⁰ It is reasonable to assume that any dermally applied Al reaching the systemic circulation would have reached equilibrium and behave in a similar manner to endogenous Al ions (i.e. being carried predominantly by transferrin). However, following the IV administration, there is suddenly a relatively large amount of Al complexed with citrate. Based on a validated PBPK model for Al distribution,^{20,21} the equilibrium between Al citrate and Al bound to transferrin was estimated to stabilize within only 15 min after the IV administration (see also Fig. S1, see online supplementary material for a color version of this figure).

Although equilibrium is reached relatively quickly, Al complexed with citrate might be more rapidly excreted than transferrin bound Al. For the brief period (15 min²⁰ and Fig. S1, see online supplementary material for a color version of this figure) following IV dosing with Al (in citrate-buffered saline), more rapid excretion may overestimate the total fraction absorbed until equilibrium with transferrin is established. Consequently, the dermal fraction absorbed might be underestimated due to a more rapid excretion of Al during these first 15 min following IV administration. This subtle difference in excretion kinetics would only have an impact on the IV phase of the study for the time following dosing, until the equilibrium between citrate and transferrin has been reached.

To quantify the uncertainty associated with this period of citrate:transferrin equilibration, the fraction absorbed was also estimated by excluding the first urine sample (0–6 h). The assumption that the Al in the first urine sample was completely excreted within 15 min resulted in only a modest increase in the estimate of mean fraction absorbed to 0.00068% (vs. 0.00052% calculated including total urine from 0 h). Thus, the impact of subtle differences in excretion for the initial 15 min post IV dose can be considered negligible. This is also supported by the observation that the AUC of the blood profile from the "misdosed" subject was comparable to the other 5 subjects. On the other hand, as fecal excretion is unlikely, this represents a slight uncertainty in the exposure calculation.

Taken together, the most reliable estimates of the dermal fraction absorbed are derived from noncompartmental analysis with the urine data; these data are supported by the noncompartmental analysis using the limited whole blood data. Furthermore, by using the IV dose to normalize the urinary excretion to 100% bioavailability, this study provides sufficiently robust data to support a reliable estimation of the fraction of Al absorbed after topical application of a representative antiperspirant formulation. The SCCS acknowledged our interpretation of the current data in their updated risk assessment.²²

In conclusion, the best estimate of the fraction absorbed of $[^{26}Al]$ from a topical application of $[^{26}Al]$ -ACH in an antiperspirant formulation is considered to be 0.00052%. Moreover, the vast majority of the Al in the formulation remains associated with the outer layers of the stratum corneum and does not penetrate the skin, but appears to be lost from the skin surface to clothing and the environment.

Supplementary material

Supplementary material is available at TOXRES Journal online.

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Authors' contributions

R.d.L. and W.V. designed the research and wrote the manuscript.

D.G., J.W., T.B., R.R., K.B., and A.W. performed the research. R.d.L., S.T., G.P., B.W., D.B., D.M., and W.V. analyzed the data.

Abbreviations

ACH: Aluminium chlorohydrate AMS: Accelerator Mass Spectrometry; Al: Aluminium; AUC: Area Under the Curve; ECG: electrocardiogram; GMP: Good Manufacturing Practice; ICP-MS: Inductively Coupled Plasma Mass Spectrometry; JECFA: Joint FAO/WHO Expert Committee on Food Additives; LLOQ: Lower Limit of Quantitation; mBq: milli-Becquerel; TNO: Nederlandse Organisatie voor Toegepast Natuurwetenschappelijk Onderzoek (Netherlands Organization for applied scientificresearch; US DOE: United States Department of Energy.

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S.T., G.P., B.W., D.B., and D.M. are employed by their respective companies, each of which use Aluminium compounds in cosmetic products.

References

- 1. Draelos DZ. Antiperspirants and the hyperhidrosis patient. Dermatol Ther. 2002:**14**(3):220–224.
- Joint FAO/WHO Expert Committee on Food Additives. Evaluation of certain food additives and contaminants. WHO Technical Report Series; 2012. p. 940.
- 3. Poirier J, Semple H, Davies J, LaPointe R, Dziwenka M, Hiltz M and Mujibi D. Double-blind, vehicle-controlled randomized twelve-month neurodevelopmental toxicity study of common aluminum salts in the rat. *Neuroscience*. 2011:338–362.
- Opinion on the safety of aluminum in cosmetic products. SCCS/1525/14 Revision of 18 June. Scientific Committee on Consumer Safety (SCCS); 2014.
- European Food Standards Agency. Safety of aluminium from dietary intake. EFSA J. 2008:754:1–34.
- 6. Willhite CC, Karyakina NA, Yokel RA, Yenugadhati N, Wisniewski TM, Arnold IMF, Momoli F and Krewski D. Systematic review of potential health risks posed by pharmaceutical, occupational and consumer exposures to metallic and nanoscale aluminum, aluminumoxides, aluminum hydroxide and its soluble salts. Crit Rev Tocicol. 2014:44 (Suppl 4):1–80.
- Allam MF. Breast cancer and deodorants/antiperspirants: a systematic review. Centr Eur J Public Health. 2016:24(3):245–247.
- Flarend R, Bin T, Elmore D and Hem SL. A preliminary study of the dermal absorption of aluminium from antiperspirants using aluminium-26. Food Chem Toxicol. 2001:39(2):163–168.
- de Ligt RAF, van Duijn E, Grossouw D, Bosgra S, Burggraaf J, Windhorst A, Peeters PAM, van der Luijt GA, Alexander-White C, Vaes WHJ. Assessment of dermal absorption of aluminum from a representative antiperspirant formulation using a ²⁶Al microtracer approach. Clin Transl Sci. 2018:**11**(6):573–581.
- 10. Risk assessment of exposure to aluminum through food and the use of cosmetic products in the Norwegian population. VKM Norwegian scientific committee for Food Safety; 2013.

- 11. Bundesgesundhbl, BD. 1998:**41**(6):271.
- Valkonen and Aitio. Analysis of aluminium in serum and urine for the biomonitoring of occupational exposure. Sci Total Environ. 1997:199(1–2):103–110.
- Ljunggren KG, Lidums V, Sjögren B. Blood and urine concentrations of aluminium among workers exposed to aluminium flake powders. Br J of Indus Med. 1991:48(2):106–109.
- Priest ND. The biological behaviour and bioavailability of aluminium in man, with special reference to studies employing aluminium-26 as a tracer: review and study update. J Environ Monit. 2004:6(5):375–403.
- Moss T, Howes D, Williams FM. Percutaneous penetration and dermal metabolism of triclosan (2,4,4'-Trichloro-2'-hydroxydiphenyl Ether). Food Chem Toxicol. 2000:38(4): 361–370.
- Bretagne A, Cotot F, Arnaud-Roux M, Sztucki M, Cabane B, Galey J-B. The mechanism of eccrine sweat pore plugging by aluminium salts using microfluidics combined with small angle X-ray scattering. Soft Matter. 2017:13(20): 3812–3821.
- Strassburger J, Coble DW. Infrared characterization of human sweat glands inhibited with aluminum chlorohydrate. J Soc Cosmet Chem. 1987:38(2):109–124.
- Quatrale RP, Coble DW, Stoner KL, Felger CB. The mechanism of antiperspirant action by aluminum salts. II. Histological observations of human eccrine sweat glands inhibited by aluminum chlorohydrate. J Soc Cosmet Chem. 1981:32:107–136.
- Shirley DG, Lote CJ. Renal handling of aluminium. Nephron Physiol. 2005:101(14):99–103.
- Steinhausen C, Kislinger G, Winklhofer C, Beck E, Hohl E, Nolte E, Ittel TH, Alvarez-Brückmann MJL. Investigation of the aluminium biokinetics in humans: a ²⁶Al tracer study. *Food Chem* Toxicol. 2004:**42**(3):363–371.
- Nolte E, Beck E, Winklhofer C, Steinhausen C. Compartmental model for aluminium biokinetics. *Hum Exp Toxicol.* 2001:20(2):111–117.
- Opinion on the safety of aluminum in cosmetic products. Submission II. SCCS/1613/19. Scientific Committee on Consumer Safety (SCCS); 2019.