

Neonatal Detection of Congenital Hypothyroidism of Central Origin

David A. van Tijn, Jan J. M. de Vijlder, Bernard Verbeeten, Jr., Paul H. Verkerk, and Thomas Vulmsma

Department of Pediatric Endocrinology (D.A.v.T., J.J.M.d.V., T.V.), Emma Children's Hospital AMC, Academic Medical Center, University of Amsterdam, NL-1100 DE Amsterdam, The Netherlands; Department of Radiology (B.V.), Academic Medical Center, University of Amsterdam, NL-1100 DE Amsterdam, The Netherlands; and Department of Social Pediatrics and Child and Youth Health Care (P.H.V.), The Netherlands Organization for Applied Scientific Research (TNO) Prevention and Health, NL-2301 CE Leiden, The Netherlands

Due to the high frequency of concurrent pituitary hormone deficiencies, congenital hypothyroidism (CH) of central origin (CH-C) is a life-threatening disorder. Yet only a minority of these patients are detected by neonatal CH screening programs worldwide. We conducted a prospective multicenter study involving a 2-yr cohort of neonatally diagnosed CH-C patients to determine whether a T_4 -TSH-based neonatal CH screening protocol extended with T_4 binding globulin determinations improves early detection of CH-C and to assess the extent of pituitary hormone deficiency among the identified CH-C patients. In all infants with screening results indicative of CH-C, the functional integrity of the hypothalamo-hypophyseal system was investigated by dynamic tests; the anatomical integrity was investi-

gated by magnetic resonance imaging. Initial test results were evaluated after 5 yr of follow-up. Among 385,000 infants screened over the 2-yr period, 19 cases of permanent CH-C were detected (prevalence, 1:20,263; 95% confidence interval, 1:12,976 to 1:33,654), representing 13.5% of all detected cases of permanent CH. The majority (78%) had multiple pituitary hormone deficiency, whereas 53% had pituitary malformations on magnetic resonance imaging. We conclude that infants with CH-C can very well be detected by neonatal screening. The estimated prevalence and the severity of pituitary dysfunction of this treatable disorder call for explicit attention for this entity of CH in neonatal screening programs worldwide. (*J Clin Endocrinol Metab* 90: 3350–3359, 2005)

CONGENITAL DISORDERS OF the hypothalamo-hypophyseal system have been known since the mid-1950s (1, 2), but comprehension of the pathogenesis of this gamut of disorders was long obscured by misinterpretation of birth characteristics and imaging studies in the pre-magnetic resonance imaging (MRI) era. Until the early 1990s, these disorders were generally considered the result of birth traumata (3). Only after the revelation of the well-defined magnetic resonance image of posterior pituitary ectopia was the plausibility of a developmental disorder recognized (4, 5). During the past decade, various transcription factors and the corresponding genes involved in pituitary development and function have been discovered. Defects in these genes give rise to multiple pituitary hormone deficiency and pituitary hypoplasia (6–11). Two of these genes are associated with posterior pituitary ectopia in a minority of cases (10, 12).

Despite the severity of hormonal dysfunction, congenital disorders of the hypothalamo-hypophyseal system are seldom diagnosed in early infancy. In contrast to a TSH-based screening program, a T_4 -based neonatal screening for congenital hypothyroidism (CH) is suited in principle for early detection of cases of CH of central origin (CH-C) (13), thus disorders of the hypothalamo-hypophyseal system. Unawareness of the vital risks and ignorance of the possibilities

to identify these patients by neonatal screening (13) have hampered the development of diagnostic protocols apt to detect this life-threatening entity of CH.

In 1995, after a 1-yr trial period, the Dutch neonatal CH-screening program, based on initial T_4 and consecutive TSH determination, was extended with a determination of T_4 binding globulin (TBG) (14). A pilot study had indicated that this addition, besides reduction of the number of false-positive screening results, could improve detection of CH-C (15). Therefore, for a period of 2 yr, all children with screening results indicative of CH-C were enrolled in a nationwide prospective study to determine whether the neonatal CH screening protocol extended with TBG determinations improves early detection of CH-C and to assess the extent of pituitary hormone deficiency among the identified CH-C patients. Of all patients detected by neonatal screening, the integrity of the hypothalamo-hypophyseal system was investigated following a standardized protocol of endocrine function testing and neuroradiologic imaging. We report the major results after 5 yr of follow-up of the cohort of 19 patients with permanent CH-C, detected by neonatal screening.

Patients and Methods

Neonatal CH screening

The neonatal CH screening procedure was based on determination of T_4 in dried blood spots obtained by heel puncture 5–7 d after birth and consecutive TSH determination of those samples in the lowest 20% of T_4 concentrations and TBG of those samples in the lowest 5% of T_4 concentrations (Fig. 1). The ratio between the T_4 and TBG concentrations was used as a measure of the free circulating T_4 concentration. In a pilot study, the cut-off value for the T_4 to TBG ratio was estimated at 8.5 (15). To prevent delayed diagnosis of the most severe cases, children with

First Published Online March 22, 2005

Abbreviations: CH, Congenital hypothyroidism; CH-C, CH of central origin; ICMA, immunochemiluminometric assay; IGFBP-3, IGF binding protein 3; MRI, magnetic resonance imaging; TBG, T_4 binding globulin; T_4 -SDS, T_4 SD score.

JCEM is published monthly by The Endocrine Society (<http://www.endo-society.org>), the foremost professional society serving the endocrine community.

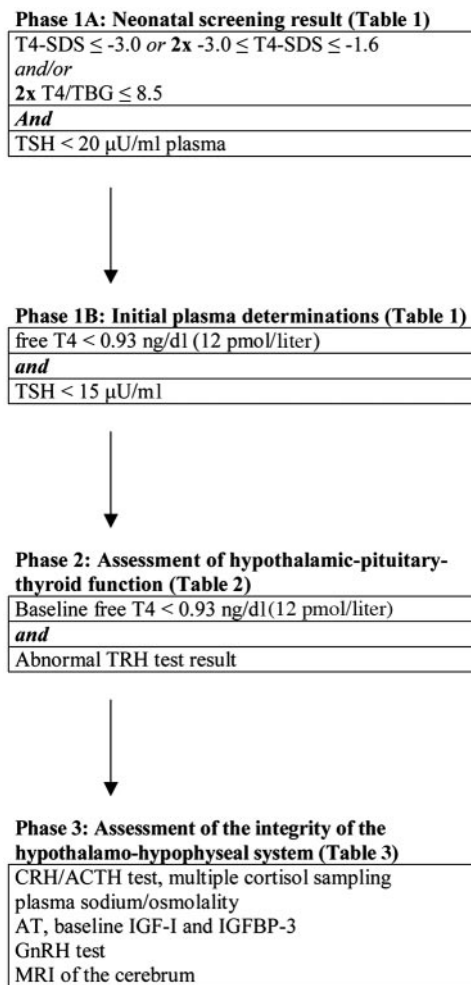


FIG. 1. Study design (flow chart). Neonatal CH screening procedure and inclusion criteria. T₄ was measured in all heel puncture samples. Of those samples in the lowest 20% of T₄ concentrations (*i.e.* ≤0.8 SD below the mean), the TSH concentration was measured in the same sample. Of those samples in the lowest 5% of T₄ concentrations (*i.e.* ≤1.6 SD below the mean), the TBG concentration was measured additionally. T₄/TBG was calculated by the following division: T₄-SDS + 5.1/TBG (nanomoles per liter). In a pilot study, the cut-off value for the T₄ to TBG ratio was estimated at 8.5 (15). To prevent delayed diagnosis of the most severe cases, children with TSH concentrations of at least 50 μU/ml plasma and/or T₄-SDS no greater than -3.0 were directly referred to a pediatrician. Children with borderline TSH (20–50 μU/ml plasma), and/or borderline T₄-SDS (-1.6 to -2.9), and/or T₄ to TBG ratio below 8.5 underwent a second heel puncture. When the result was abnormal or again borderline, the child was referred to a pediatrician. The screening result was considered indicative of CH-C if T₄-SDS and/or T₄ to TBG ratios were abnormal, with an accompanying heel stick TSH below 20 μU/ml plasma. Children with screening results indicative of CH-C were included in the study if the consecutive venous TSH plasma concentration was below 15 μU/ml (15 mU/liter) and free T₄ was below 0.93 ng/dl (12 pmol/liter). Conversion factors: TBG, 1 μg/dl = 18.52 nmol/liter; TSH, 1 μU/ml = 1 mU/liter; free T₄, 1 ng/dl = 12.87 pmol/liter. T₄-SDS, T₄-SDS (of the series of that day); TBG, TBG concentration (in nanomoles per liter); T₄/TBG, T₄ to TBG ratio; AT, arginine test.

TSH concentrations more than or equal to 50 μU/ml (50 mU/liter) plasma and/or T₄ SD scores (T₄-SDS) less than or equal to -3.0 were directly referred to a pediatrician. Children with borderline TSH [20–50 μU/ml plasma (20–50 mU/liter)], and/or borderline T₄-SDS (-1.6 to -2.9), and/or T₄ to TBG ratio less than 8.5 underwent a second heel

puncture. When the result was abnormal or again borderline, the child was referred to a pediatrician. The screening result was considered indicative of CH-C if T₄-SDS and/or T₄ to TBG ratio was abnormal, with accompanying heel stick TSH less than 20 μU/ml (20 mU/liter) plasma.

Study patients

The study design is depicted in Fig. 1. The subjects were participants in a Dutch nationwide prospective study, running from April 1, 1994, to April 1, 1996. Infants with neonatal CH-screening results indicative of CH-C and subsequent plasma free T₄ concentrations below the beforehand estimated cut-off of 0.93 ng/dl (12 pmol/liter) (16) and plasma TSH concentrations less than 15 μU/ml were enrolled. A diagnosis of CH-C was assessed by a TRH test. Anterior pituitary function was assessed primarily by stimulation tests. TRH and CRH tests took place on consecutive days as soon as the alleged CH-C patient was referred; T₄ supplementation was installed immediately after TRH test results had proven CH-C. Arginine and GnRH tests were performed at the age of 3 months, when euthyroid status had been accomplished by T₄ supplementation, using the opportunity provided by the postnatal surge of gonadotropins and sex steroids to evaluate gonadotropin function (17, 18). Also, in most patients, MRI of the cerebrum was performed around the age of 3 months, to assess the anatomical integrity of the hypothalamo-hypophyseal system. In 2001, all cases were reevaluated for revised diagnoses, additional morbidity, growth, and treatment data. The study protocol was approved by the Dutch Pediatric Endocrine Society and by the Medical Ethics Committees of the participating centers. Parental informed consent was obtained in all cases.

Investigation of the thyrotropic hormone axis

Plasma TSH was measured before and 15, 30, 45, 60, 120, and 180 min after iv administration of TRH (10 μg·kg⁻¹). An adequate TSH response to TRH (type 0) was defined by a peak concentration exceeding 15 μU/ml (19–21) and return to baseline within 3 h (19, 22). In response to TRH, CH-C patients either show diminished (type 2), or slightly delayed but excessive increase and delayed decrease of the TSH plasma concentration (type 3) (19, 20).

Investigation of the ACTH axis

Plasma ACTH and cortisol were measured before and 5, 10, 15, 30, 45, 60, 120, and 150 min after iv administration of CRH (1 μg·kg⁻¹). An adequate response was defined by an ACTH peak concentration exceeding 80 pg/ml or four times baseline level and a cortisol peak concentration exceeding 18 μg/dl (500 nmol/liter) or 7 μg/dl (200 nmol/liter) over baseline level (23). In selected cases, a short ACTH test was performed (see Table 2). An adequate response to ACTH was defined by a cortisol peak concentration exceeding 18 μg/dl (500 nmol/liter) or 7 μg/dl (200 nmol/liter) over baseline level (24, 25). Of all patients, multiple random plasma cortisol samples were taken. Concentrations exceeding 18 μg/dl (500 nmol/liter) were considered adequate (23).

Investigation of the somatotropic hormone axis

Plasma GH was measured before and 30, 45, 60, 75, 90, 120, and 180 min after iv administration of arginine (500 mg·kg⁻¹). A GH concentration exceeding 10 ng/ml (20 mU/liter) was considered an adequate response (26). Function test results were complemented by baseline plasma IGF-I and IGF binding protein 3 (IGFBP-3) determinations (27).

Investigation of the gonadotropic hormone axis

Plasma LH and FSH were measured before and 15, 30, 45, 60, and 120 min after iv administration of GnRH (10 μg·kg⁻¹). An adequate response was defined by peak concentrations of LH exceeding 3 mU/ml and FSH exceeding 6 mU/ml in girls, 3 mU/ml in boys (28).

Investigation of posterior pituitary function

In all patients, plasma sodium and potassium concentrations and diuresis rate were determined.

TABLE 1. Phase 1—Perinatal data and neonatal screening results

Subject	Perinatal data					First neonatal screening result					Second neonatal screening result					Inclusion		
	Sex	GA (wk)	BBM [g (percentile)]	Birth mode	Perinatal morbidity	First sign	Age (d)	T ₄ [μ g/dl (SDS)]	TBG (μ g/dl)	T ₄ /TBG (μ U/ml)	TSH (μ U/ml)	Age (d)	T ₄ [μ g/dl (SDS)]	TBG (μ g/dl)	T ₄ /TBG (μ U/ml)	TSH (μ U/ml)	Age (d)	F ₁₄ (mg/dl)
1	M	36.9	2310 (3–10)	H	1,2	S	5	0.9 (-4.7)	11.8	1.8	<3	16	6.4 (-2.7)	21.5	6.0	<3	7	0.33
2	F	40.9	2795 (3)	B, SC	1	S	5	6.0 (-2.1)	20.6	7.9	<3	16	6.4 (-2.7)	21.5	6.0	<3	25	0.58
3	F	34.1	2045 (25–50)	B	3,4	C(3)	5	7.2 (-2.3)	25.4	6.0	<3	16	6.4 (-2.7)	21.5	6.0	<3	17	0.58
4	M	41	3830 (50–75)	H, VE	3	S	5	7.8 (-1.9)	24.1	7.2	3.8	13	5.9 (-3.3)	17.1	5.7	5	49	0.61
5	M	40.4	4680 (>97)	H, VE	5	S	9	7.5 (-2.4)	23.1	6.3	5	20	6.2 (-2.3)	18.3	8.3	<2	39	0.64
6	M	40.3	4100 (75–90)	B	3,4,6	S	6	6.5 (-2.4)	19.0	7.7	<2	20	6.2 (-2.3)	18.3	8.3	<2	29	0.65
7	M	41	4950 (>97)	H	3	S	5	6.2 (-3.6)	9.9	8.2	5	28	6.5 (-2.1)	13.5	12.0	<3	36	0.65
8	F	39.7	3390 (50)	H	0	S	15	7.3 (-2.7)	14.1	9.2	<3	28	6.5 (-2.1)	13.5	12.0	<3	49	0.65
9	F	38.1	3230 (50–75)	H	3,4,6	S	7	4.4 (-3.9)	16.3	4.0	5	12	6.1 (-2.6)	22.5	6.0	5	15	0.66
10	M	41.7	4650 (97)	H	3,4	S	5	6.2 (-2.7)	16.4	7.9	5	12	6.1 (-2.6)	22.5	6.0	5	19	0.71
11	M	40	3000 (10)	H	4	S	5	0.6 (>-4.3)	31.8	1.3	<2	29	10.0 (-1.8)	32.3	5.9	5	137	0.78
12	M	32	1375 (10)	H, SC	4,7,8	C(7)	8	2.7 (-4.0)	17.1	3.5	<2	29	10.0 (-1.8)	32.3	5.9	5	50	0.82
13	M	40	3080 (10–25)	B, FE	1,4,7,9	S	6	8.8 (-1.9)	21.9	7.9	5	19	10.5 (-1.6)	32.3	5.9	5	39	0.84
14	M	42	3600 (25–50)	H	0	S	7	4.6 (-3.2)	21.9	7.9	1.2	14	7.6 (-2.9)	16.1	7.4	4	47	0.85
15	M	42	3720 (50)	H	0	S	7	10.4 (-2.1)	22.5	7.2	7	14	7.6 (-2.9)	16.1	7.4	4	23	0.86
16	M	37.4	1785 (<3)	B, FE	3,4,8,9	S	6	7.2 (-2.4)	18.8	7.8	5	13	7.7 (-2.2)	22.6	6.9	5	777	0.87
17	M	42	3400 (10–25)	H	0	S	6	11.3 (-1.6)	23.1	8.2	5	13	7.7 (-2.2)	22.6	6.9	5	56	0.88
18	M	38.3	1845 (<3)	H, FE	3,4,6,7,10	S	8	6.8 (-2.7)	15.5	8.4	<3	15	6.0 (-2.4)	16.4	8.9	<3	26	0.88
19	M	40.7	3530 (25–50)	H	0	S	7	4.4 (-3.3)	9.9	9.8	<3	17	6.9 (-3.1)	18.3	5.9	5	17	0.91
20	F	42	3220 (10–25)	H	0	S	5	7.0 (-2.7)	16.0	8.1	5	22	6.9 (-3.1)	18.3	5.9	5	36	0.92
21	F	39.7	3000 (10–25)	H	3,11	S	13	4.2 (-3.1)	21.1	5.2	<2	28	9.3 (-2.3)	22.5	6.3	4	17	0.96
22	M	41.4	4160 (75–90)	H	0	S	7	7.8 (-2.4)	22.4	6.6	8	28	9.3 (-2.3)	22.5	6.3	4	55	0.99
23	M	39.1	3250 (25–50)	H	0	S	8	9.4 (-2.0)	19.8	8.5	3	28	9.6 (-1.8)	22.7	7.8	1.5	40	1.00
24	M	41	3000 (3–10)	H	0	S	6	8.3 (-2.0)	19.8	8.5	5	18	7.9 (-1.9)	21.2	8.1	5	93	1.06
25	M	38.6	2740 (10)	H	12	S	5	1.7 (-3.7)	14.8	5.1	<3	17	9.5 (-1.1)	24.4	8.8	3	47	1.07
26	F	42	3920 (75)	H	0	S	7	3.6 (-3.2)	5.0	20.7	7	17	9.5 (-1.1)	24.4	8.8	3	14	1.54

Patients were ranked according to plasma free T₄ concentration at inclusion. GA, Gestational age; BBM, birth body mass. Percentiles: Niklasson *et al.* (48). Birth mode: H, head position; B, breech position; SC, cesarean section; VE, vacuum extraction; FE, forcipal extraction. Perinatal morbidity/first sign: 0, no manifest morbidity; 1, asphyxia; 2, cardiorespiratory insufficiency; 3, hypoglycemia; 4, hyperbilirubinemia and/or elevated transaminases; 5, umbilical hernia; 6, persistent vomiting; 7, hypogonadism; 8, maldescensus testis; 9, lethargy, poor feeding; 10, hyponatremia (transient); 11, neonatal seizures; 12, neonatal sepsis/meningitis; S, congenital hypothyroidism screening result: first sign recognized; C, recognition on clinical grounds. Neonatal screening: hormone determinations in eluates of filter paper blood spots, obtained by heel puncture. SDS, SD score. Conversion factors: TSH, 1 μ U/ml = 1 mU/liter; T₄, 1 μ g/dl = 12.87 nmol/liter; free T₄, 1 ng/dl = 12.87 pmol/liter; TBG, 1 μ g/dl = 18.52 nmol/liter.

MRI

To evaluate pituitary morphology, MRI studies were performed under general anesthesia, in most patients using a 1.5 Tesla Siemens Magnetom (Siemens, Munich, Germany). Transversal (5 mm), sagittal (3 mm), and coronal (3 mm) T1-weighted spin-echo images were obtained. In the transversal plane, proton density as well as T2-weighted turbo-spin echo sequences were used also. In addition, a T1-weighted 3D series (MPRage) was taken.

Assays

TSH was measured by immunochemiluminometric assay (ICMA; Behring, Amsterdam, The Netherlands). T_4 , T_3 , and rT_3 were measured by in-house RIA methods (AMC, Amsterdam, The Netherlands). Free T_4 was measured by a two step RIA assay (SPAC FT4 Fraktion; Byk-Sangtek Diagnostica, Dietzenbach, Germany). TBG was measured by RIA (Eiken Chemical Co., Tokyo, Japan). ACTH was measured by ICMA (Nichols Institute, Wychen, The Netherlands). Cortisol was measured by fluorescence polarization immunoassay (TDx; Abbott, Amstelveen, The Netherlands). LH and FSH were measured by ICMA (Amerlite; Amersham Biosciences, Little Chalfont, UK). 17- β -Estradiol was measured by RIA (estradiol-2; Sorin Biomedica, Saluggia, Italy). Testosterone was measured by RIA (Coat-a-Count, Diagnostic Products Corp., Los Angeles, CA). GH was measured by RIA (Spectria/Orion, Turku, Finland). IGF-I and IGFBP-3 were measured by RIA (in-house assays Wilhelmina Children's Hospital, University Medical Center Utrecht, Utrecht, The Netherlands).

Statistical analysis

SPSS 10.1 (SPSS, Inc., Chicago, IL) was used for statistical computations. All reported *P* values are two-sided.

Results

Study patients

Of 385,000 infants screened during the study period, 26 met with the inclusion criteria. Perinatal characteristics and endocrine test results of these subjects are summarized in Tables 1, 2, and 3.

Nineteen infants were diagnosed with permanent CH-C. In 17 of these, the CH-screening provided the first clue toward a diagnosis of CH-C. The age at inclusion (the day on which the TRH test was performed) ranged from 7–56 d (median 36 d), except for two patients, in whom investigations took place after the neonatal period; patient no. 16 was not referred and tested before the age of 2.1 yr, despite screening results indicative of CH-C; patient no. 11, born in India, was screened and tested at the age of 4 months, after the family had moved to The Netherlands. Patient no. 7 could not be completely tested in the neonatal period because T_4 supplementation had already been started at referral. TSH deficiency was confirmed by a TRH test at the age of 1 yr, after temporary discontinuation of T_4 supplementation. One patient (no. 1) with neonatal screening results and plasma TSH and free T_4 concentrations indicative of CH-C plus inadequately low plasma cortisol concentrations had multiple congenital defects associated with a chromosomal deletion del6pter→p22. He died of cardiorespiratory insufficiency before further endocrine tests could be performed.

One infant (patient no. 19), born to a mother with previ-

TABLE 2. Phase 2—Dynamic tests of thyrotropic function

Subject	Age (d)	TRH test				Overall result
		Free T_4 at t = 0 (ng/dl)	TSH at t = 0 (μ U/ml)	TSH peak concentration (μ U/ml) at time (min)	TSH at t = 180 (μ U/ml)	
1						NA
2	25	0.58	3.0	49.0 (120)	32.0	A
3	17	0.58	10.0	72.0 (30)	58.0	A
4	49	0.61	6.6	30.4 (120)	18.1	A
5	39	0.64	2.6	10.4 (30)	2.6	A
6	29	0.65	1.5	16.8 (120)	14.0	A
7	358	0.52	1.7	10.6 (30)	2.0	A
8	49	0.65	4.0	42.4 (45)	18.4	A
9	15	0.66	4.2	30.2 (20)	18.8	A
10	19	0.71	2.3	7.8 (30)	3.9	A
11	137	0.78	<0.05	<0.05 (all)	<0.05	A
12	50	0.82	4.8	4.8 (0)	4.0	A
13	39	0.84	1.9	54.0 (120)	28.4	A
14	47	0.85	3.8	10.6 (30)	4.1	A
15	23	0.86	4.0	11.2 (15)	4.4	A
16	777	0.87	0.8	8.2 (60)	4.4	A
17	56	0.88	3.2	9.2 (30)	4.4	A
18	26	0.88	3.3	31.2 (45)	14.8	A
19	17	0.91	2.1	4.4 (30)	2.2	A
20	36	0.92	4.0	36.8 (60)	17.6	A
21	17	0.96	6.5	34.0 (30)	7.0	N
22	55	0.99	11.2	37.6 (30)	2.5	N
23	40	1.00	2.4	14.0 (30)	3.5	N
24	93	1.06	2.0	28.2 (30)	4.8	N
25	47	1.07	4.2	14.8 (20)	3.4	N
26	14	1.54	5.0	21.6 (30)	6.0	N

A, Abnormal; N, normal; NA, not assessed. Patient 1, no TRH test; baseline values: TSH, 5.9 μ U/ml; T_4 , 1.3 μ g/dl (17 nmol/liter); free T_4 , 0.33 ng/dl (4.3 pmol/liter); triiodothyronine, 20 ng/dl (0.3 nmol/liter) at age 7 d. Patient 9, No TSH determination at 180 min; 120 min value used instead. Conversion factors: TSH, 1 μ U/ml = 1 mU/liter; T_4 , 1 μ g/dl = 12.87 nmol/liter; free T_4 , 1 ng/dl = 12.87 pmol/liter; T_3 , 1 ng/dl = 0.01536 nmol/liter.

TABLE 3. Phase 3—Assessment of the integrity of the hypothalamo-hypophyseal system; neuroradiologic imaging; clinical features

Patient	Adrenocorticotrophic hormone axis					Somatotrophic hormone axis					Gonadotrophic hormone axis					MRI	Clinical features	
	CRH test		ACTH test		Baseline	AT		Baseline		GnRH-test		Baseline		Overall result				
	ACTH peak (pg/ml)	Cortisol peak (μg/dl)	Cortisol peak (μg/dl)	ACTH test	Random cortisol (μg/dl)	Overall result	GH peak (ng/ml)	IGF-I (ng/ml)	IGFBP-3 (μg/ml)	Retarded growth	Overall result	LH peak (mU/ml)	FSH peak (mU/ml)		Testosterone (ng/ml)			Estradiol (pg/ml)
1					10	NA										NA	NA	1
2	120	4	4	21	1	A	6.2	27	0.54	+	A	<0.3	2.7		<5	A	0	
3					6*	A	8.5	30	0.30	+	A	5.4	20.0		11	N	NA	2
4	55	3			<2	A	1.6	23	0.69	+	A	<0.3	<0.5			A	1	
5	90	38			17	N	15.9	49	0.79	-	N	12.0	14.0			N	2,3,4	3
6	78	6			1	A	4.0			+	A					A	1	
7	140	32			26	N	11.3	45	0.59	-	N	5.4	6.6			N	0	
8	120	16			8	N	30.8	27	1.36	+	N†	6.4	22.0		13	N	1,2,4,5,6	4
9	24	2			2	A	<0.3	8		+	N	<0.3	1.3		<5	A	1,2	
10	75	18			4	B	20.7	48	0.99	-	N	10.0	4.4			N	5	5
11	165	25			33	N	<0.5	5	0.12	+	A					NA	0	6
12					<2	A	9.0	25	0.25	+	A	5.2	3.1			N	1,5	7
13	130	16			10	B	11.7	25	0.70	+	N†	0.9	2.0			A	1	
14	100	19			21	N	17.6	37	0.66	-	N	15.0	19.0			N	0	3
15	30	10			3	A	10.6	19	0.53	-	N	11.0	11.0			N	3	
16	260	48			24	N	2.7	15	0.52	+	A					NA	1	8
17	290	29			16	N	18.2	84	1.06	-	N	3.6	12.0			N	0	3
18	115	6			<2	A	4.6	13	0.61	+	A	0.6	1.5			A	1	9
19	110	17			18	N				-	NA					NA	NA	
20	165	7			8	A	5.2	28	0.72	+	A	0.7	4.8		<5	A	1	

AT, Arginine test; estradiol, 17β-estradiol. Random cortisol: *, during hypoglycemia. Overall result: NA, Not assessed; N, normal; A, abnormal; B, borderline result; patients showed normal adrenocorticotrophic function after 2-yr and 1-yr cortisol supplementation, respectively; (sec), patients 8 and 13 showed secondary growth retardation. MRI: 0, no manifest malformation; 1, posterior pituitary ectopia; 2, hydrocephalus externus; 3, hydrocephalus internus; 4, arachnoidal cyst(s); 5, corpus callosum agenesis; 6, bilateral periventricular nodular heterotopia. Clinical features: 1, deltopter->p22; 2, low-set ears, ventricular septal defect; 3, familial occurrence; patient no. 5, nephew and grandfather with same disorder, patients 14 and 17, younger brother with same disorder; 4, clubbed feet, father with cleft lip and palate; 5, brachycephaly; mother with hypothyroidism of unknown origin; 6, POU1F1 (Pit-1) deletion (49); 7, mental retardation, microcephaly, large ears, sparse hair, resembling Oliver-McFarlane syndrome; 8, multiple dysmorphias resembling fetal alcohol syndrome, small mandible, blepharophimosis; 9, (congenital) cataract, impaired hearing, clubbed feet, hypoplastic thumbs, atrial septal defect II. Conversion factors: ACTH, 1 pg/ml = 1 ng/liter; cortisol, 1 μg/dl = 27.59 nmol/liter; GH, 1 ng/ml = 2 mU/liter; IGF-I, 1 ng/ml = 1 μg/liter; IGFBP-3, 1 μg/ml = 1 mg/liter; LH, 1 mU/ml = 1 U/liter; FSH, 1 mU/ml = 1 U/liter; testosterone, 1 ng/ml = 3.467 nmol/liter; 17-β-estradiol, 1 pg/ml = 3.671 pmol/liter.

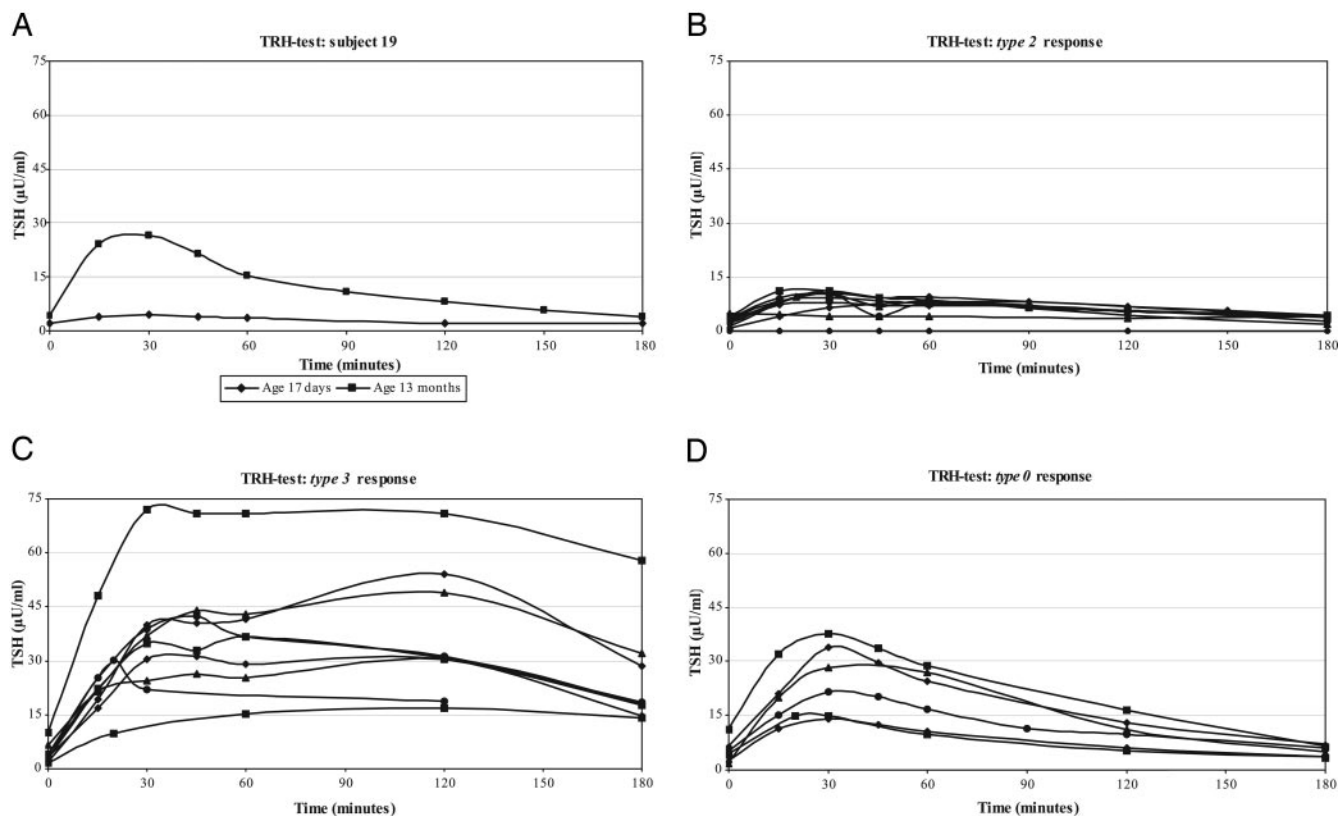


FIG. 2. Dynamic tests of thyrotropic function. A, Subject 19: type 2 response at the age of 17 d, type 0 response at the age of 13 months. B, Subjects 5, 7, 10–12, and 14–17: type 2 response. C, Subjects 2–4, 6, 8, 9, 13, 18, and 20: type 3 response. D, Subjects 21–26: type 0 response.

ously unnoticed Graves’ disease, had abnormal TRH test results at the age of 17 d [TSH peak concentration of 4.4 $\mu\text{U}/\text{ml}$ (4.4 mU/liter)]. By then, his free T_4 concentration had already increased to a near normal concentration of 0.91 ng/dl (11.7 pmol/liter) without medical intervention. Spontaneous restoration of thyroid function was awaited. From the age of 5 wk until the age of 13 months, TSH and free T_4 concentrations in plasma were within the normal range. A TRH test at the age of 13 months demonstrated normal TSH secretion (Fig. 2A) [TSH peak concentration, 26.4 $\mu\text{U}/\text{ml}$ (26.4 mU/liter)]. Consequently, he was diagnosed with transient CH-C (29).

Six infants (patient nos. 21–26) with neonatal screening results indicative of CH-C appeared to have baseline plasma free T_4 concentrations exceeding 0.93 ng/dl (12 pmol/liter) at referral. Therefore, a diagnosis of CH-C was rejected, and we refrained from further testing after the initial tests of

thyrotropic and adrenocorticotrophic function. The putative causes for the false-positive screening results are summarized in Table 4. At follow-up, none of the surviving false-positive subjects have developed signs or symptoms of hypopituitarism. Among these subjects with false-positive screening results, the age of inclusion ranged from 14–93 d (median 44 d).

Neonatal CH screening results

Neonatal CH screening took place 5–15 d after birth (median 6 d), except for patient no. 11, who was screened at the age of 130 d, after he had moved from India to The Netherlands. The patients’ TSH concentrations in dried heel puncture blood spots ranged from below detection level to 7 $\mu\text{U}/\text{ml}$ plasma, T_4 concentrations ranged from 0.6–11.3 $\mu\text{g}/\text{dl}$ (8–145 nmol/liter), and TBG concentrations ranged

TABLE 4. Putative explanations for false-positive screening results (subjects 21–26)

Subject	Putative explanation for false-positive screening results
21	Parenchymal hemorrhage in glandula pinealis region and intraventricular hemorrhage, Ohtahara epilepsy; transient central hypothyroidism, possibly due to elevated intracranial pressure or arterial insufficiency. Alternatively, the abnormal screening results could be due to a “sick euthyroid state.”
22	Transient mild central hypothyroidism of unknown origin; normal TSH and free T_4 concentrations at long-term follow-up.
23	Low normal (free) T_4 values, but adequate TSH response to TRH; no clinical signs of hypothyroidism, normal growth and development at follow-up: normal variant?
24	Transient mild central hypothyroidism of unknown origin; normal TSH and free T_4 concentrations at long-term follow-up.
25	Perinatal streptococcal meningoenephalitis; sinus thrombosis, extensive infarction a. cerebri posterior area, slightly dilated ventricles; transient central hypothyroidism, possibly due to elevated intracranial pressure or arterial insufficiency.
26	TBG deficiency and erroneous initial free T_4 determination.

from 9.9–31.8 $\mu\text{g}/\text{dl}$ (183–589 nmol/liter). All patients except nos. 8 and 19 had T_4 to TBG ratios below the cut-off level of 8.5, previously indicated for discrimination of CH-C patients from false-positives in the screening population (15). Patients 8 and 19 were referred on the basis of low screening T_4 -SDS and consecutive plasma free T_4 concentrations less than 0.93 ng/dl (12 pmol/liter) and plasma TSH concentrations less than 15 $\mu\text{U}/\text{ml}$ (15 mU/liter ; Table 1).

Investigation of the thyrotropic hormone axis

All infants with baseline plasma free T_4 concentrations less than 0.93 ng/dl (12 pmol/liter) exhibited type 2 ($n = 9$) or type 3 ($n = 9$) responses to TRH (Fig. 2, B and C). The cut-off level for diminished (type 2) TSH response could be estimated at 14 $\mu\text{U}/\text{ml}$ (14 mU/liter ; Table 2). The patients' type 2 and type 3 responses, expressed as area under the curve, were significantly different ($P = 0.001$ for type 2; $P = 0.013$ for type 3 by the Mann-Whitney U test) from the responses of subjects 21–26, who exhibited type 0 responses (Fig. 2D).

Investigation of the ACTH axis

Nine of 19 patients tested (47%) were found to be deficient, whereas eight had clearly adequate results and two (patients 10 and 13) had borderline results and were given cortisol-supplementation for 1 and 2 yr, respectively. After discontinuation of supplementation, they had normal adrenocorticotrophic function, as evidenced by a normal circadian cortisol rhythm.

Of the nine patients found deficient, four showed impaired response of both ACTH and cortisol to CRH. Another three patients (nos. 2, 18, and 20) showed adequate ACTH response, but impaired cortisol response. ACTH tests in two of the latter infants resulted in impaired cortisol response accordingly. Two patients were considered too small (no. 12) or too ill (no. 3) to undergo a CRH test, for which a relatively large amount of blood is required. Instead, ACTH tests were performed. The results of random plasma cortisol samples taken in all subjects were in accordance with the function test results (Table 3).

Investigation of the somatotropic hormone axis

After 5 yr of follow-up, 12 of 18 (67%) patients with permanent CH-C alive [from the initial 20 patients, one (no. 1) had died, and one (no. 19) had transient CH-C] have shown retarded growth. Arginine tests, performed at the age of 3 months when euthyroid status had been accomplished by T_4 supplementation, were abnormal in 10 of those infants and normal in all infants with normal growth (Table 3). Two infants (nos. 8 and 13) with normal arginine test results showed growth retardation around the age of 2.5 yr. Both had repeated GH stimulation tests that yielded severely impaired GH release after stimulation with arginine, GHRH (no. 8) and clonidine (no. 13), respectively. Baseline growth factors were measured at the age of 3 months. Significant differences between children with normal *vs.* retarded growth (exclusive of subjects 8 and 13) were observed in arginine test results ($P = 0.001$ by the Mann-Whitney U test) as well as baseline plasma concentrations of IGF-I and

IGFBP-3 at age 3 months ($P = 0.001$ and $P = 0.011$, respectively).

Investigation of the gonadotropic hormone axis

Six of 15 patients tested (40%) did not reach plasma peak concentrations of LH exceeding 3 mU/ml (3 U/liter) and FSH exceeding 6 mU/ml (6 U/liter ; girls) or 3 mU/ml (3 U/liter ; boys) after infusion of GnRH. Two of those had hypogonadism. All six hypogonadotropic patients had baseline morning plasma concentrations of testosterone (boys) less than 0.6 ng/ml (2 nmol/liter) or 17- β -estradiol (girls) less than 11 pg/ml (40 pmol/liter) at age 3 months. One patient (no. 6) who was not subjected to a GnRH test did have an abnormally low baseline plasma testosterone concentration (Table 3).

Investigation of posterior pituitary function

None of the subjects studied had symptoms indicating disturbance of posterior pituitary function.

MRI of the brain

Nine of 17 patients (53%) who underwent MRI had a small anterior pituitary lobe, located in the sella turcica, invisible or very thin pituitary stalk, and ectopic posterior pituitary lobe, located at the median eminence in the floor of the third ventricle (Fig. 3). Six patients had cerebral malformations, such as agenesis of the corpus callosum and bilateral periventricular nodular heterotopia and/or hydrocephalus. All patients with posterior pituitary ectopia had multiple pituitary hormone deficiency (Table 3).

Discussion

In the first 2 yr after introduction of the novel neonatal CH screening protocol in The Netherlands, we identified 19 cases of permanent CH-C among 385,000 infants screened, representing 13.5% of the total number of patients with permanent CH ($n = 141$) (30–32) as detected by neonatal screening.

Initial estimates of the prevalence of CH-C among live born children in the United States and Canada ranged from 1:110,000 to 1:29,000 (33–35). In a similar retrospective study of the 1981–1989 screening population in The Netherlands, we estimated a prevalence of 1:26,000 (36). Three fourths of these children turned out to have multiple pituitary hormone deficiency. Delayed detection and incomplete diagnosis of the hormonal deficiencies in question resulted in significant morbidity such as severe hypoglycemia and neonatal hepatitis and a mortality rate as high as 14% (36). Because all deficient hormones can readily be supplemented, timely diagnosis significantly improves the outcome (37). The prevalence of 1:20,263 live born neonates (95% confidence interval, 1:12,976 to 1:33,654), as estimated in the present study, is higher than previously reported (33–36), presumably due to the fact that this is the first representative and unbiased cohort, efficiently detected. In a recent evaluation of the 1995–2000 period, we estimated a prevalence of 1:16,404 (95% confidence interval, 1:13,174 to 1:21,173) at an estimated detection rate of 91.6% (38). Because it is unlikely that the prevalence of CH-C in The Netherlands is significantly dif-

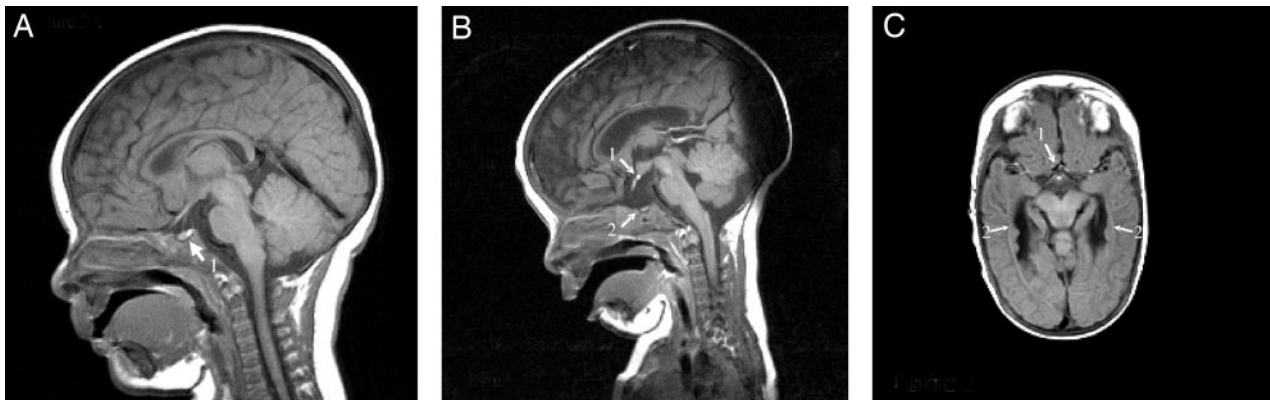


FIG. 3. MRI of the brain. A, T1-weighted spin echo midsagittal image of a 3-month-old boy, screened for neonatal hemangiomas. Normal configuration of pituitary and pituitary stalk. Arrow points at sella turcica with pituitary anterior lobe (gray) and posterior lobe (bright white). B and C, T1-weighted spin echo images of patient no. 8 at age 3 months. B, Midsagittal image showing ectopic posterior pituitary lobe (arrow 1) at the median eminence in the floor of the third ventricle and small anterior pituitary lobe in undersized sella turcica (arrow 2). No manifest pituitary stalk. C, Transversal image showing posterior pituitary ectopia (arrow 1) and bilateral periventricular nodular heterotopia (arrows 2).

ferent from that in North America, we presume that in prior studies (33–36) both the mildest and the most severe cases have been missed, similar to the situation in 21α -hydroxylase deficiency in the prescreening era (39, 40).

An anticipated Achilles' heel of our study is the fact that normal values for endocrine function tests for this specific age group are largely unavailable. However, it turned out that patients with CH-C could be discriminated very well from the infants with false-positive screening results on the basis of their TRH test results. Further evidence was acquired from the fact that the majority of CH-C patients (78%) had multiple pituitary hormone deficiency and 53% had a pituitary malformation as visualized by MRI. All patients had baseline plasma free T_4 concentrations less than 0.93 ng/dl (12 pmol/liter) at referral.

TRH tests of CH-C patients show either diminished (type 2) (19–21), or slightly delayed but excessive increase and delayed decrease of the TSH plasma concentration (type 3) (19, 20). Generally, type 2 responses are considered to reflect disturbance at the pituitary level, whereas type 3 responses reflect disturbance at the hypothalamic level. Accordingly, the clinical pictures associated with these responses are termed secondary/pituitary and tertiary/hypothalamic hypothyroidism. Remarkably, seven of the nine patients with the pituitary malformation posterior pituitary ectopia (Table 3) exhibit type 3 responses. Apparently, not the disturbance of the pituitary itself, but instead the disturbance of the interrelation between hypothalamus and (anterior) pituitary gland causes the pituitary deficiency.

Consequently, our study confirms other reports (41–43) that the distinction of secondary and tertiary hypothyroidism is improper. Therefore, we recommend the use of the term "congenital hypothyroidism of central origin" (CH-C) for neonates with apparent TSH deficiency (44) and congenital disorders of the hypothalamo-hypophyseal system to cover multiple pituitary hormone deficiency.

Forty-seven percent of the CH-C patients had evidence of glucocorticoid deficiency. Remarkable was our observation that three patients exhibited substantial rise of plasma ACTH but not cortisol concentrations in response to CRH. This phenomenon, indicative of adrenocortical hypoplasia as a

result of long-time insufficient stimulation by ACTH, has not been described before.

Sixty-seven percent of the CH-C patients had evidence of GH deficiency, as shown by growth retardation and catch-up growth after installment of GH treatment. Two of these patients had normal test results at the age of 3 months. They showed retarded growth in the third year of life and at repeated testing proved severely GH deficient.

Gonadal function cannot be definitively assessed at this age. However, we speculated that the temporary activity of the gonadotropic axis in infancy (17, 18) would enable function testing at the age of 3 months. It will take another decade to validate the results, but GnRH test results and baseline testosterone and 17β -estradiol concentrations clearly divided the study population into two groups. Nine of 15 infants tested had results comparable to those in Tanner stages 2–4. Six infants (40%) had baseline (morning) plasma concentrations of testosterone less than 0.6 ng/ml (2 nmol/liter) or 17β -estradiol less than 11 pg/ml (40 pmol/liter) and did not reach LH concentrations above 3 mU/ml (3 U/liter) and FSH concentrations above 6 mU/ml (6 U/liter; girls) or 3 mU/ml (3 U/liter; boys) after infusion of GnRH.

None of the patients had symptoms indicating disturbance of posterior pituitary function neonatally or at follow-up. However, partial diabetes insipidus cannot be ruled out, because it has been shown that, even in the absence of polyuria, polydipsia, or nocturnal enuresis, the vasopressin response to osmolar stimuli might very well be subnormal in patients with (congenital) hypopituitarism and posterior pituitary ectopia, suggesting damage in the hypothalamic vasopressin secreting centers (45).

The MRI studies performed in 17 of the patients revealed nine cases (53%) of posterior pituitary ectopia, a pituitary malformation often associated with multiple pituitary hormone deficiency (5, 46, 47). All of these infants had deficiencies of one or more pituitary hormones besides TSH. In fact, the emergence of a second deficiency of anterior pituitary function was predicted by the finding of posterior pituitary ectopia in patient no. 8, who initially had TSH deficiency only.

In conclusion, our data indicate that infants with permanent CH-C, making up 13.5% of all permanent CH patients, can be detected very well in a T₄-based neonatal CH screening program, extended with TSH and TBG determinations. The majority of patients detected have multiple pituitary hormone deficiency, a life-threatening disorder. Therefore, neonatal screening should be followed by assessment of the integrity of the hypothalamo-hypophyseal system without delay. In this respect, the protocol here presented proved feasible and effective. CH-C warrants early detection and timely treatment, and thus putative reduction of mortality and severe morbidity, such as neuroglycopenia and consequent irreversible damage to the central nervous system. The estimated prevalence and the severity of pituitary dysfunction of this treatable disorder call for explicit attention for this entity of CH in neonatal screening programs worldwide.

Acknowledgments

We are indebted to the patients and their parents; to the referring pediatricians and pediatric endocrinologists; Johan Waelkens, for the National Advisory Board on Congenital Hypothyroidism; Gerard Loeber, for the National Institute of Public Health and the Environment [Rijksinstituut voor Volksgezondheid en milieu (RIVM)]; Erik Endert, Academic Medical Center, University of Amsterdam; Jeany Huijser-Geenen, Free University Medical Center, Amsterdam; Jaap van Doorn, Wilhelmina Children's Hospital, University of Utrecht Medical Center, for the collaborating endocrinologic reference laboratories; Brenda Wiedijk, for assistance in collection of the data; and the Ludgardine Bouwman Foundation, for continuous support of the scientific work of Prof. De Vijlder.

Received December 13, 2004. Accepted March 10, 2005.

Address all correspondence and requests for reprints to: David A. van Tijn, M.D., Department of Pediatric Endocrinology, Emma Children's Hospital AMC, Academic Medical Center, G2-133, University of Amsterdam, P.O. Box 22700, 1100 DE Amsterdam, The Netherlands. E-mail: tijn1@planet.nl.

This work was supported by Grant 28-1060-2 (to J.J.M.d.V. and T.V.) from The Netherlands Organization for Health Research and Development (ZON-MW, The Hague, The Netherlands).

References

- Mosier HD 1956 Hypoplasia of the pituitary and adrenal cortex. *J Pediatr* 48:633–639
- Blizzard RM, Alberts M 1956 Hypopituitarism, hypoadrenalism and hypogonadism in the newborn infant. *J Pediatr* 48:782–792
- Fujita K, Matsuo N, Mori O, Koda N, Mukai E, Okabe Y, Shirakawa N, Tamai S, Itagane Y, Hibi I 1992 The association of hypopituitarism with small pituitary, invisible pituitary stalk, type 1 Arnold-Chiari malformation, and syringomyelia in seven patients born in breech position: a further proof of birth injury theory on the pathogenesis of "idiopathic hypopituitarism". *Eur J Pediatr* 151:266–270
- Kelly WM, Kucharczyk W, Kucharczyk J, Kjos B, Peck WW, Norman D, Newton TH 1988 Posterior pituitary ectopia: an MR feature of pituitary dwarfism. *Am J Neuroradiol* 9:453–460
- Triulzi F, Scotti G, Di Natale B, Pellini C, Lukezic M, Scognamiglio M, Chiumello G 1994 Evidence of a congenital midline brain anomaly in pituitary dwarfs: a magnetic resonance imaging study in 101 patients. *Pediatrics* 93:409–416
- Ingraham HA, Albert VR, Chen R, Crenshaw 3rd EB, Elsholtz HP, He X, Kapiloff MS, Mangalem HJ, Swanson LW, Treacy MN, Rosenfeld MG 1990 A family of POU-domain and Pit-1 tissue-specific transcription factors in pituitary and neuroendocrine development. *Annu Rev Physiol* 52:773–791
- Wu W, Cogan JD, Pfäffle RW, Dasen JS, Frisch H, O'Connell SM, Flynn SE, Brown MR, Mullis PE, Parks JS, Phillips 3rd JA, Rosenfeld MG 1998 Mutations in PROP1 cause familial combined pituitary hormone deficiency. *Nat Genet* 18:147–149
- Dattani MT, Martinez-Barbera J-P, Thomas PQ, Brickman JM, Gupta R, Martensson IL, Toresson H, Fox M, Wales JK, Hindmarsh PC, Krauss S, Beddington RS, Robinson IC 1998 Mutations in the homeobox gene HESX1/Hesx1 associated with septo-optic dysplasia in human and mouse. *Nat Genet* 19:125–133
- Netchine I, Sobrier ML, Krude H, Schnabel D, Maghnie M, Marcos E, Duriez B, Cacheux V, Moers A, Goossens M, Gruters A, Amselem S 2000 Mutations in LHX3 result in a new syndrome revealed by combined pituitary hormone deficiency. *Nat Genet* 25:182–186
- Machinis K, Pantel J, Netchine I, Leger J, Camand OJ, Sobrier ML, Dastot-Le Moal F, Duquesnoy P, Abitbol M, Czernichow P, Amselem S 2001 Syndromic short stature in patients with a germline mutation in the LIM homeobox LHX4. *Am J Hum Genet* 69:961–968
- Burrows HL, Douglas KR, Seasholtz AF, Camper SA 1999 Genealogy of the anterior pituitary gland: tracing a family tree. *Trends Endocrinol Metab* 10:343–352
- Brickman JM, Clements M, Tyrell R, McNay D, Woods K, Warner J, Stewart A, Beddington RS, Dattani M 2001 Molecular effects of novel mutations in Hesx1/HESX1 associated with human pituitary disorders. *Development* 128:5189–5199
- Dussault JH 1999 The anecdotal history of screening for congenital hypothyroidism. *J Clin Endocrinol Metab* 84:4332–4334
- Robertson EF, Wilkins AC, Oldfield RK, Pollard AC 1980 Blood-spot thyroxine-binding globulin: a means to reduce recall rate in a screening strategy for neonatal hypothyroidism. *J Pediatr* 97:604–607
- Verkerk PH, Loeber JG, Vulsma T, De Vijlder JJM 1995 [The benefit of extension of the CH-screening procedure by determination of thyroxine-binding globulin; Dutch]. Leiden: Netherlands Organisation for Applied Scientific Research TNO (TNO-publication PG 95.062. ISBN 90-6743-391-8)
- Vulsma T 1991 Etiology and pathogenesis of congenital hypothyroidism. Dissertation, University of Amsterdam
- Forest MG, Sizonenko PC, Cathiard AM, Bertrand J 1974 Hypophyso-gonadal function in humans during the first years of life. *J Clin Invest* 53:819–828
- Main KM, Schmidt IM, Skakkebaek NE 2000 A possible role for reproductive hormones in newborn boys: progressive hypogonadism without the postnatal testosterone peak. *J Clin Endocrinol Metab* 85:4905–4907
- Okuno A, Taguchi T, Nakayama K, Takimoto M 1979 Kinetic analysis of plasma TSH after TRH stimulation. *Horm Metab Res* 11:293–295
- Gruñeiro de Papendieck L, Iorcansky S, Rivarola MA, Heinrich JJ, Bergadá C 1982 Patterns of TSH response to TRH in children with hypopituitarism. *J Pediatr* 100:387–392
- Rapaport R, Sills I, Patel U, Oppenheimer E, Skuza K, Horlick M, Goldstein S, Dimartino J, Saenger P 1993 Thyrotropin-releasing hormone stimulation tests in infants. *J Clin Endocrinol Metab* 77:889–894
- Jacobsen BB, Andersen H, Dige-Petersen H, Hummer L 1976 Thyrotropin response to thyrotropin-releasing hormone in full-term, euthyroid and hypothyroid newborns. *Acta Paediatr Scand* 65:433–438
- Forest MG 1992 Adrenal function tests. In: Ranke MB, ed. *Functional endocrinologic diagnostics in children and adolescents*. 1st ed. Mannheim, Germany: J&J Verlag; 248–274
- Lytras N, Grossman A, Perry L, Tomlin S, Wass JA, Coy DH, Schally AV, Rees LH, Besser GM 1984 Corticotrophin releasing hormone: responses in normal subjects and patients with disorders of the hypothalamus and pituitary. *Clin Endocrinol (Oxf)* 20:71–84
- Schürmeyer TH, Avgerinos PC, Gold PW, Gallucci WT, Tomai TP, Cutler Jr GB, Loriaux DL, Chrousos GP 1984 Human corticotropin-releasing factor in man: pharmacokinetic properties and dose-response of plasma adrenocorticotropin and cortisol secretion. *J Clin Endocrinol Metab* 59:1103–1108
- Guyda HJ 1999 Four decades of growth hormone therapy for short children: what have we achieved? *J Clin Endocrinol Metab* 84:4307–4316
- Blum WF, Albertsson-Wikland K, Rosberg S, Ranke MB 1993 Serum levels of insulin-like growth factor I (IGF-I) and IGF binding protein 3 reflect spontaneous growth hormone secretion. *J Clin Endocrinol Metab* 76:1610–1616
- De Muinck Keizer-Schrama SMPF, Hazebroek FWJ 1986 The treatment of cryptorchidism: why, how, when? Dissertation, Erasmus University; 110–136
- Kempers MJ, van Tijn DA, van Trotsenburg AS, de Vijlder JJ, Wiedijk BM, Vulsma T 2003 Central congenital hypothyroidism due to gestational hyperthyroidism: detection where prevention failed. *J Clin Endocrinol Metab* 88:5851–5857
- Verkerk PH, Lanting CI 2000 [Report on CH screening results of children born in 1999 and follow-up on children born in 1994]. Leiden: Netherlands Organisation for Applied Scientific Research TNO (TNO-publication PG/JGD 00.059) (Dutch)
- Lanting CI, Verkerk PH 2001 [Report on CH screening results of children born in 2000 and follow-up on children born in 1995]. Leiden: Netherlands Organisation for Applied Scientific Research TNO (TNO-publication 2001.158) (Dutch)
- Blankespoor MN, Lanting CI, Verkerk PH 2002 [Report on CH screening results of children born in 2001 and follow-up on children born in 1996]. Leiden: Netherlands Organisation for Applied Scientific Research TNO (TNO-publication 2002.260) (Dutch)
- Fisher DA, Dussault JH, Foley Jr TP, Klein AH, LaFranchi S, Larsen PR, Mitchell ML, Murphey WH, Walfish PG 1979 Screening for congenital hypothyroidism: results of screening one million North American infants. *J Pediatr* 94:700–705

34. Fisher DA, Klein AH 1981 Thyroid development and disorders of thyroid function in the newborn. *N Engl J Med* 304:702–712
35. Hanna CE, Krainz PL, Skeels MR, Miyahira RS, Sesser DE, LaFranchi SH 1986 Detection of congenital hypopituitary hypothyroidism: ten-year experience in the Northwest Regional Screening Program. *J Pediatr* 109:959–964
36. Vulsma T, Delemarre HA, de Muinck Keizer SMPF, Wiedijk BM, Gons MH, Verkerk PH, De Vijlder JJM 1989 Detection and classification of congenital thyrotropin deficiency in the Netherlands. In: *The thyroid gland, environment and autoimmunity*. Amsterdam: Excerpta Medica, International Congress Series; 896:343–346
37. Hintz RL 1996 Eternal vigilance—mortality in children with growth hormone deficiency. *J Clin Endocrinol Metab* 81:1691–1692
38. Lanting CI, Van Tijn DA, Loeber JG, Vulsma T, De Vijlder JJM, Verkerk PH, Clinical and cost-effectiveness of the use of T4/TBG ratio to detect congenital hypothyroidism of thyroidal and central origin in a neonatal screening program. *Pediatrics*, in press
39. Balsamo A, Cacciari E, Piazzini S, Cassio A, Bozza D, Pirazzoli P, Zappulla F 1996 Congenital adrenal hyperplasia: neonatal mass screening compared with clinical diagnosis only in the Emilia-Romagna region of Italy, 1980–1995. *Pediatrics* 98:362–367
40. Jääskeläinen J, Voutilainen R 2000 Long-term outcome of classical 21-hydroxylase deficiency: diagnosis, complications and quality of life. *Acta Paediatr* 89:183–187
41. Mehta A, Hindmarsh PC, Stanhope RG, Brain CE, Preece MA, Dattani MT 2003 Is the thyrotropin-releasing hormone test necessary in the diagnosis of central hypothyroidism in children. *J Clin Endocrinol Metab* 88:5696–5703
42. Cohen LE, Wondisford FE, Salvatoni A, Maghnie M, Brucker-Davis F, Weintraub BD, Radovick S 1995 A “hot spot” in the Pit-1 gene responsible for combined pituitary hormone deficiency: clinical and molecular correlates. *J Clin Endocrinol Metab* 80:679–684
43. Vieira TC, Dias da Silva MR, Cerutti JM, Brunner E, Borges M, Arnaldi LT, Kopp P, Abucham J 2003 Familial combined pituitary hormone deficiency due to a novel mutation R99Q in the hot spot region of Prophet of Pit-1 presenting as constitutional growth delay. *J Clin Endocrinol Metab* 88:38–44
44. Vulsma T, De Vijlder JJM 2005 Genetic defects causing hypothyroidism. In: Braverman LE, Utiger RD, eds. *Werner, Ingbar’s the thyroid: a fundamental and clinical text*. 9th ed. Chap 48. Philadelphia: Lippincott Williams & Wilkins
45. Lukezic M, Righini V, Di Natale B, De Angelis R, Norbiato G, Bevilacqua M, Chiumello G 2000 Vasopressin and thirst in patients with posterior pituitary ectopia and hypopituitarism. *Clin Endocrinol (Oxf)* 53:77–83
46. Chen S, Leger J, Garel C, Hassan M, Czernichow P 1999 Growth hormone deficiency with ectopic neurohypophysis: anatomical variations and relationship between the visibility of the pituitary stalk asserted by magnetic resonance imaging and anterior pituitary function. *J Clin Endocrinol Metab* 84:2408–2413
47. Osorio MG, Marui S, Jorge AA, Latronico AC, Lo LS, Leite CC, Estefan V, Mendonca BB, Arnhold IJ 2002 Pituitary magnetic resonance imaging and function in patients with growth hormone deficiency with and without mutations in GHRH-R, GH-1, or PROP-1 genes. *J Clin Endocrinol Metab* 87:5076–5084
48. Niklasson A, Ericson A, Fryer JG, Karlberg J, Lawrence C, Karlberg P 1991 An update of the Swedish reference standards for weight, length and head circumference at birth for given gestational age (1977–1981). *Acta Paediatr Scand* 80:756–762
49. Hendriks-Stegeman BI, Augustijn KD, Bakker B, Holthuisen P, van der Vliet PC, Jansen M 2001 Combined pituitary hormone deficiency caused by compound heterozygosity for two novel mutations in the POU domain of the Pit1/POU1F1 gene. *J Clin Endocrinol Metab* 86:1545–1550

JCEM is published monthly by The Endocrine Society (<http://www.endo-society.org>), the foremost professional society serving the endocrine community.