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Catalytic organosolv fractionation of willow wood and wheat straw as pretreatment for enzymatic cellulose hydrolysis

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Abstract

BACKGROUND: Ethanol-based organosolv fractionation of lignocellulosic biomass is an effective pretreatment technology for enzymatic cellulose hydrolysis to produce sugars and lignin within a biorefinery. This study focuses on the catalytic effect of H₂SO₄, HCl, and MgCl₂ on organosolv pretreatment of willow wood and wheat straw.

RESULTS: The use of catalysts improved fractionation of both feedstocks. The maximum enzymatic cellulose digestibility obtained was 87% for willow wood (using 0.01 mol L⁻¹ H₂SO₄ as catalyst) and 99% for wheat straw (0.02 mol L⁻¹ HCl). Non-catalytic organosolv fractionation at identical conditions resulted in 74% (willow wood) and 44% (wheat straw) glucose yield by enzymatic hydrolysis. Application of catalysts in organosolv pretreatment was particularly effective for wheat straw. The influence of the acid catalysts was found to be primarily due to their effect on the pH of the organosolv liquor. Acid catalysts particularly promoted xylan hydrolysis. MgCl₂ was less effective than the acid catalysts, but it seemed to more selectively improve delignification of willow wood.

CONCLUSION: Application of catalysts in organosolv pretreatment of willow wood and wheat straw was found to substantially improve fractionation and enzymatic digestibility. The use of catalysts can contribute to achieving maximum utilization of lignocellulosic biomass in organosolv-based biorefineries.

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Keywords: organosolv; catalyst; pretreatment; fractionation; enzymatic hydrolysis; wheat straw; willow wood

INTRODUCTION

Organosolv fractionation of lignocellulosic biomass has been widely studied as a pretreatment technology for enzymatic cellulose hydrolysis within a biorefinery to produce fermentable sugars and high-purity lignin.^{1,2} 'Organosolv' was originally developed as pulping technology for paper production³ and comprises a class of processes in which lignocellulosic biomass is treated with an organic solvent at elevated temperatures. In the case of application of organosolv as pretreatment technology, the function of the organic solvent is to delignify the lignocellulosic biomass prior to enzymatic cellulose hydrolysis.¹ Thus, the enzymatic digestibility of the biomass is improved by reducing the recalcitrance of the biomass as well as minimizing the non-productive binding of hydrolytic enzymes to lignin.^{1,2,4} The extracted lignin fraction can be recovered from the organosolv liquor for further processing.^{1,5} Organosolv lignins tend to have a high purity and be essentially free of sulphur and ash. Moreover, the functionalities of native lignin are largely preserved in organosolv lignins.^{6,7} These characteristics are beneficial for the possible conversion of lignin into valuable products such as aromatic chemicals, resins, and antioxidants.^{8–11} The valorization potential of the lignin fraction is a major benefit of the organosolv process compared with other pretreatment technologies like steam explosion and mild acid pretreatment.^{1,4} In order to be cost-competitive with these other pretreatment technologies, valorization of the organosolv lignin has to compensate for the

higher costs of the organosolv process due to, for example, the required solvent recycling.^{1,4} Finally, the third major structural constituent of lignocellulosic biomass, hemicellulose, is converted during the organosolv process into sugars and sugar derivatives like furfural.^{1,4}

Various organic solvents have been used for organosolv including alcohols (such as methanol and ethanol), acetone, and organic acids (such as formic and acetic acid).¹ Ethanol has been studied extensively both at lab-scale¹ and at pilot-scale (Ligno^{12,13}). Ethanol-based organosolv pretreatment has been successfully applied to all major types of lignocellulosic biomass: hardwoods (e.g. poplar¹⁴ and olive tree¹⁵), softwoods (e.g. pine¹⁶ and a mixture of spruce, pine, and Douglas fir¹³), and herbaceous crops (e.g. miscanthus¹⁷ and sugar cane bagasser¹⁸). If ethanol-based organosolv is carried out without the use of an additional catalyst, the process is often termed autocatalytic. Acetic acid is formed during the organosolv process from acetyl side groups of the hemicellulose. Subsequently, this acid acts as a catalyst by lowering the pH of the organosolv liquor. However,

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the autocatalytic effect of acetic acid might not be effective enough depending on, among others, the type of biomass and limitations of other process conditions, such as reaction temperature. In those cases, the addition of a catalyst might be required. Examples of catalysts that have been studied for alcohol-based organosolv pulping and pretreatment are: mineral acids (e.g. H_2SO_4 , HCl , H_3PO_4), organic acids (e.g. formic and acetic acid), bases (e.g. NaOH) and salts (e.g. MgCl_2).^{1,3} Catalysts have been reported to promote hemicellulose hydrolysis,^{4,19} cleavage of ether bonds within the lignin fraction (delignification),^{10,20} and enzymatic cellulose digestibility.^{19,21} Possible drawbacks of (acid) catalysts include costs of catalysts, corrosion issues,^{3,21} and promotion of carbohydrate and lignin degradation and condensation reactions^{1,22} resulting in increased formation of fermentation inhibitors like furfural.^{4,21}

Although numerous studies have been published on catalytic organosolv, only a few compare the effect of different catalysts on a specific type of lignocellulosic biomass. Moreover, these studies are mainly limited to organosolv treatment of softwoods. For example, Yawalata and Paszner studied the effect of different chlorine salts,²³ magnesium salts,²² and acid catalysts²² on methanol-based organosolv pulping of spruce. They obtained optimum results with MgCl_2 . Besides, H_2SO_4 and HCl were found to be capable of liberating the fibres.²² Park *et al.* tested the use of sulphuric acid, magnesium chloride, and sodium hydroxide in ethanol-based organosolv pretreatment of pitch pine.²¹ The use of sulphuric acid was reported to give the highest theoretical ethanol yield based on the enzymatic cellulose digestibility obtained. Finally, Del Rio *et al.* reported on organosolv pretreatment of mountain pine beetle-killed lodgepole pine using ethanol and butanol as solvents and various catalysts including MgCl_2 and H_2SO_4 .¹⁹ They found that acid pretreatment led to pulps that were readily hydrolyzed by enzymes due to a lower degree of cellulose polymerization, shorter fibre lengths, and increased substrate porosity.

The objective of this work is to study the effect of different catalysts on organosolv pretreatment of both a hardwood (willow wood) and a herbaceous crop (wheat straw). Their effect on both the fractionation degree and enzymatic cellulose digestibility was studied. Both acid and non-acid catalysts, which are common for organosolv pulping and pre-treatment, were used (i.e. H_2SO_4 , HCl , and MgCl_2). Finally, a comparison of the effect of the different catalysts was made for both feedstocks.

MATERIALS AND METHODS

Feedstocks

Two types of lignocellulosic biomass were used for this study: willow wood and wheat straw. Willow wood was selected as hardwood species because it is considered a potential energy crop within The Netherlands. Willow wood was received as <3 cm ambient-dry chips from Oostvaardershoeve, Slootdorp, The Netherlands, and was milled with a cutting mill (Retsch SM 2000) to <0.5 mm. Wheat straw was selected because it is an abundant agricultural residue in Europe. In addition, it currently has limited commercial value.²⁴ Winter wheat straw was kindly provided by Wageningen UR - Food & Biobased Research, Wageningen, The Netherlands and received ambient-dry and cut to <2 cm. Both feedstocks were stored in closed vessels at room temperature until use.

For organosolv experiments, a representative sample was taken and used without further treatment. The moisture content of the feedstocks was determined regularly either by drying at

Table 1. Composition lignocellulose feedstocks

Composition (% w/w dry biomass)			Biomass	
			Willow wood	Wheat straw
Biochemical	Extractives	H_2O	4.4	8.4
		EtOH	1.1	2.0
		Polysaccharides	Arabinan	1.1
	Xylan		11.7	19.9
	Mannan		1.4	0.2
	Galactan		1.1	0.7
	Glucan		32.9	36.9
	Rhamnan		0.4	0.1
	Lignin	AIL	28.5	16.7
		ASL	2.1	1.1
		Ash	4.1	6.1
	Sum	88.8	93.8	
	Elemental ^a	C	48.0	44.3
H		5.5	5.4	
N		0.6	0.3	
O		42.6	42.7	
Al		0.067	0.004	
Ca		0.614	0.236	
Cl		0.027	0.022	
Fe		0.056	0.005	
K		0.350	0.289	
Mg		0.072	0.039	
Na		0.045	0.017	
P		0.112	0.030	
S		0.064	0.052	
Si	0.233	2.238		
Sum	98.3	95.6		

^a Other elements measured <0.01% w/w dry biomass.

105 °C according to protocol NREL/TP-510-42621²⁵ (willow wood) or using a halogen moisture analyzer (Mettler Toledo HR83, Columbus, OH) (wheat straw). The average moisture content of the willow wood and wheat straw raw materials was ~10% and ~9%, respectively (w/w dry biomass, variations in moisture content for both feedstocks were <1%). The composition of both feedstocks is given in Table 1. The biochemical composition of the feedstocks was determined as described in the section 'composition analysis of solids'. The elemental composition was measured with an elemental analyzer (Carlo Erba Instruments FLASH EA 1112, Wigan, UK) (C,H,N, and O), ion chromatography after bomb combustion (Cl), and inductively coupled plasma atomic emission spectroscopy (ICP-AES) (other elements).

Finally, the acid neutralizing capacity (ANC) of both feedstocks was determined according to CEN TS 14429²⁶ using a final pH of 2 and measurement time of 48 h. The ANC was 0.36 mol H^+ kg^{-1} dry biomass for willow wood and 0.26 mol H^+ kg^{-1} dry biomass for wheat straw. The difference in ANC is probably caused by a difference in composition of the mineral fraction of both feedstocks. The mineral fraction of the wheat straw mainly consists of Si (Table 1). The major element in the mineral fraction of the willow wood is Ca, which provides alkalinity. Thus the ANC of willow wood is higher than that of wheat straw in spite of the lower ash content of willow (Table 1).

Selection of catalysts

Three different catalysts were selected for this study: H₂SO₄, HCl, and MgCl₂. Sulphuric acid is the acid catalyst most often used for organosolv fractionation of various types of lignocellulose, both for pulping³ and for pretreatment prior to enzymatic cellulose hydrolysis.¹ HCl was used as alternative acid, since it has been reported to be effective in methanol, ethanol, and acetic acid-based (Acetosolv) organosolv pulping.³ Finally, MgCl₂ was selected as a non-acid catalyst based on work by Yawalata and Paszner.^{22,23} They reported MgCl₂ to be the optimum catalyst in an extensive study on the catalytic effect of various salts on methanol-based organosolv pulping of spruce wood for paper applications (neutral alkali earth metal (NAEM) process).^{22,23} Although limited information about the (potential) catalytic mechanism of MgCl₂ is available,²⁷ its catalytic effect on the organosolv process has been confirmed by other authors, particularly for softwood pulping.^{19,21}

Organosolv experiments

Organosolv experiments were performed in six parallel 125 mL batch reactors (acid digestion bomb type 4748, SS 316 with Teflon liner, Parr Instrument Company, Moline, IL). 6 g (as received) lignocellulose was mixed with 31 g ethanol (ethanol 96%, Merck, purity: 92.6–95.2% w/w, average 94% w/w used within calculations), 21 g demineralized water and a catalyst. The resulting ethanol–water ratio was 55–56% w/w including moisture content of the feedstock. The liquid–solid (L/S)-ratio was 9.5–9.7 kg kg⁻¹ dry biomass for all experiments. A catalyst was added in a concentration of 0–0.025 mol L⁻¹ (H₂SO₄), 0–0.05 mol L⁻¹ (HCl) or 0–0.1 mol L⁻¹ (MgCl₂). The acid catalysts were added in equal normalities (0–0.05 N) to enable direct comparison. The closed reactor vessels were placed in a heating block (adapted RS600, Thermo Fisher Scientific, Rochford, UK) and the block temperature was set at 190 °C. Stirring was provided by a magnetic stir bar at the bottom of the vessels (1000 rpm). Visual tests with model slurries showed that the mixing in the reactor was adequate, except at the beginning of an experiment. After 180 min, the heating block was switched off and the reactors were allowed to cool down. Experiments were performed overnight with fixed start and end time.

The experimental set-up used did not allow measurement of the internal reactor temperature during the organosolv experiments. As an estimate of the development of this temperature, Fig. 1 shows the average internal temperature profile of two reactors filled with 75 mL Shell Thermia oil B (recorded with a thermocouple, identical settings as for organosolv experiments). The average maximum reactor temperature measured was 187 °C. A pretreatment severity factor (log *R*₀) of 4.65 was calculated by integration over the entire temperature profile ($R_0 = \sum e^{\frac{T - T_{ref}}{14.75} \cdot \Delta t}$ with *T*_{ref} = 100 °C).²⁸ Temperature differences between the six individual reactors were estimated to be less than 2 °C for all experiments.

After organosolv treatment, the suspension was filtered quantitatively over a Whatman GF/D filter and the pH of the organosolv liquor was measured at room temperature as an approximation of its hydrogen ion concentration. The solid fraction remaining after organosolv was washed with 60 mL 56% w/w ethanol–water and dried overnight in a vacuum oven at 50 °C. Drying of the pulp was performed for practical reasons and was purposely performed at mild conditions to limit its possible influence on the enzymatic digestibility of the pulp.^{29,30} Finally,

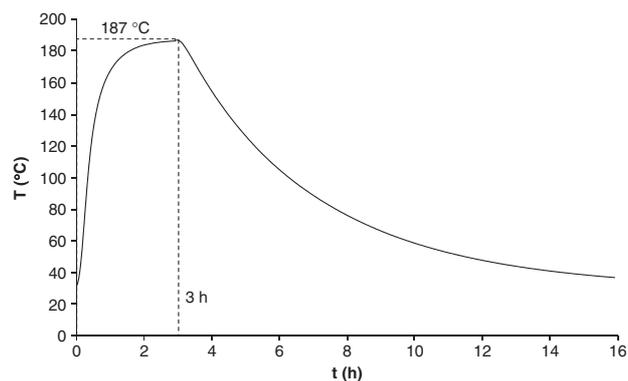


Figure 1. Average temperature profile recorded of two reactors filled with Thermia B oil (set-point: 190 °C, switch off heating after 3 h).

the pulp yield was calculated (% dw recovered solids/dw starting material).

During the wheat straw fractionation experiments, the organosolv liquor and the washing solution were combined and analyzed for their composition. Monomeric sugars including glucose and xylose were determined with high-performance anion exchange chromatography with pulsed amperometric detection (HPAEC-PAD) (Dionex ICS3000, Sunnyvale, CA). The sugar derivatives furfural, hydroxymethylfurfural (HMF), and levulinic acid were determined using high-performance liquid chromatography (HPLC-RI/UV, with BIO RAD Aminex HPX-87H column, 65 °C, 5 mmol L⁻¹ H₂SO₄).

Composition analysis of solids

The composition of raw materials and a selection of pretreated samples were analyzed using analytical procedures published by NREL²⁵ and TAPPI.³¹ The moisture content was determined using a halogen moisture analyzer (Mettler Toledo HR83). The ash content of the raw materials was measured by combustion at 550 °C according to protocol NREL/TP-510-42622.²⁵ The biochemical composition (i.e. content of extractives, lignin, hemicelluloses, and cellulose) of these solids was determined in duplicate using a modified hydrolysis protocol based on TAPPI methods T 222 and 249.³¹ First, in the case of raw materials, extractives were removed with two successive Soxhlet extractions using water and ethanol (NREL/TP-510-42619²⁵). Second, the sample was milled with a cutting mill and hydrolyzed in two steps: (1) 72% w/w H₂SO₄ (30 °C, 1 h) and (2) 4% w/w H₂SO₄ (100 °C, 3 h). The solid residue was determined gravimetrically and its ash content was measured according to NREL/TP-510-42618.²⁵ The acid-insoluble lignin (AIL) content was based on the amount of ash-free residue. Finally, the hydrolysate was analyzed for monomeric sugars (HPAEC-PAD) as well as acid-soluble lignin (ASL, UV-VIS absorption at 205 nm). Analyses of a reference sample (willow wood pretreated with non-catalytic ethanol-based organosolv), included in each series of biochemical composition analyses, yielded relative experimental errors of: AIL (1.5%), ASL (6.3%), glucan (2.8%), and xylan (3.1%) (σ , *n* = 8). For more details about the analysis procedure and the calculation principles applied, we refer the reader to Huijgen *et al.*³⁰

Enzymatic digestibility

The enzymatic digestibility of fresh and a selection of pretreated lignocellulose samples was measured in duplicate according to

NREL/TP-510-42629.²⁵ The moisture content of the substrate was determined using a halogen moisture analyzer (Mettler Toledo HR83). A suspension of 0.6 g biomass, 20 mL 0.05 mol L⁻¹ Na-acetate buffer (pH 4.8), and an antibiotic (12 μL PenStrep containing 5000 U mL⁻¹ penicillin and 5 mg mL⁻¹ streptomycin) to eliminate potential bacterial growth was shaken in an incubator. After heating to 50 °C, 0.32 mL of enzyme was added and incubation was continued for 72 h. The enzyme used was Accellerase 1500 (Genencor, Rochester, NY) and the cellulase activity of the enzyme batch used was 62 FPU mL⁻¹, determined according to the protocol of Ghose.³² The enzyme dose used corresponds to about 33 FPU g⁻¹ substrate. This dose has not been optimized and is assumed to be sufficiently high to ensure an excess of enzyme. At fixed intervals, samples were taken from the mixture for glucose determination. Glucose concentrations were determined with a colorimetric protocol derived from Yee and Goodwin.³³ The enzymatic glucose yield was calculated on the basis of the mean glucose concentration in the hydrolysate after 72 h and the glucan content of the substrate. For more details we refer the reader to our earlier work.³⁰

A reference sample was included in each series of enzymatic hydrolysis tests. The variation in maximum glucose concentrations resulting from the reference sample within 72 h enzymatic hydrolysis was 3.2% between the series (σ , $n = 4$). The variation in glucose concentration at each time interval was 1.3–4.5% (σ , $n = 4$, excluding $t = 0$ h).

Error analysis

In each of the three series of organosolv experiments per feedstock, an organosolv experiment in which no catalyst was added was included as a reference point (Table 2). Thus, the non-catalytic experiment was performed in triplicate for each feedstock. From its results, the experimental errors made were calculated both for fractionation degree and enzymatic digestibility. Subsequently, these errors were used as an estimate of the experimental error made in all (catalytic) organosolv experiments. All errors are based on a single standard deviation.

RESULTS AND DISCUSSION

Fractionation

Figure 2 shows the pulp yield and final pH of the organosolv liquor obtained from organosolv pretreatment of willow wood over the full range of catalyst concentrations applied. Based on these results, it has been decided to select MgCl₂ concentrations of 0–0.025 mol L⁻¹ for pulp composition analysis and enzymatic hydrolysis tests. First, a higher MgCl₂ dose has limited effect on the pulp yield. Second, a maximum dose of 0.025 mol L⁻¹ has also been applied for H₂SO₄, which simplifies comparison. In Table 2, the yield and composition of the pulp obtained during selected organosolv experiments are given. Figure 3 shows the corresponding fractionation degrees (i.e. xylan hydrolysis, delignification, and glucan recovery in pulp).

No catalyst

Non-catalytic organosolv, performed in triplicate for each feedstock (Table 2), has resulted in an average pulp yield of: 69.3 ± 0.7% dw (willow) and 76.4 ± 2.0% dw (straw). The willow pulp consists on average of 46.6 ± 1.2% glucan, 12.2 ± 0.3% xylan, and 26.6 ± 1.5% total lignin (i.e. AIL + ASL). The corresponding fractionation degrees are: xylan hydrolysis: 27.5 ± 2.5%, glucan recovery:

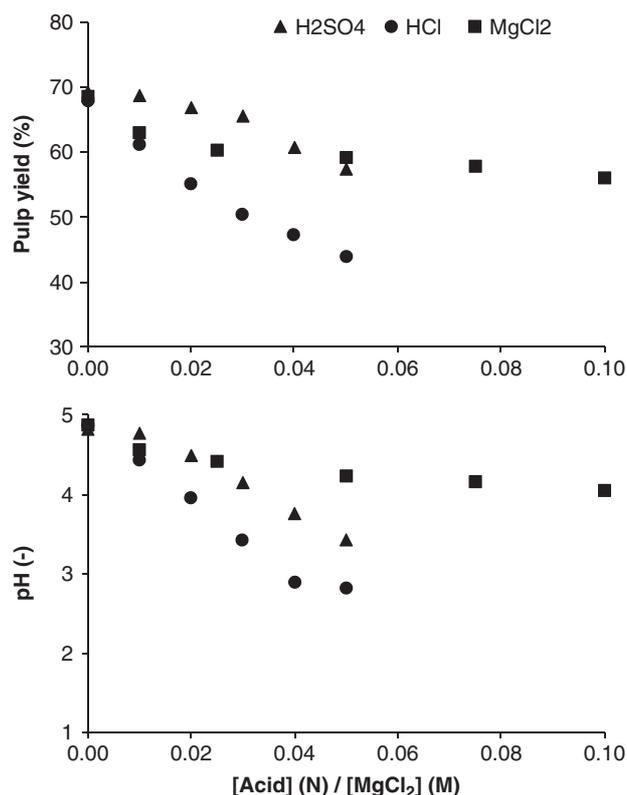


Figure 2. Pulp yield and final pH of liquor resulting from organosolv fractionation of willow wood as function of catalyst dose.

98.1 ± 1.9%, and delignification: 39.6 ± 4.0%. The straw pulp contains on average 49.1 ± 1.0% glucan, 20.3 ± 0.3% xylan, and 16.0 ± 0.8% total lignin, resulting in the following fractionation degrees; xylan hydrolysis: 21.8 ± 1.8%, glucan recovery: 101.8 ± 1.2%, and delignification: 31.2 ± 5.1%. Based on these numbers, the organosolv experiments performed seem reproducible. The delignification degree has the largest experimental uncertainty. These experimental uncertainties are used in Fig. 3 as an estimate of the experimental error made. Finally, during non-catalytic organosolv experiments, the pH of the organosolv liquor decreases to 4.8–4.9 by formation of acetic acid for both feedstocks (Table 2).

Acid catalysts

Addition of an acid catalyst (H₂SO₄ or HCl) reduces the pH of the organosolv liquor (Table 2) and thereby lowers the pulp yield of the organosolv pretreatment step for both feedstocks (Fig. 3). Acid catalysts particularly promote xylan hydrolysis up to 96% and 99% for willow wood and wheat straw, respectively (0.05 mol L⁻¹ HCl). In addition, acid catalysts increase delignification up to 59% (0.05 mol L⁻¹ HCl) and 74% (0.025 mol L⁻¹ H₂SO₄) for willow wood and wheat straw, respectively (Fig. 3). The splitting of α - and β -ether linkages within the lignin molecule are known to be acid-catalysed reactions.³⁴ At (too) high acid concentrations, the glucan fraction of the biomass is partly hydrolyzed (up to 38% in the case of 0.05 mol L⁻¹ HCl and wheat straw). Based on the pH values given in Table 2, it can be concluded that the final pH of the organosolv liquor should not be lower than about 3.5 in order to avoid cellulose hydrolysis. Yawalata and Paszner claimed a pH of about 4 to be optimal for organosolv pulping.²²

Table 2. Catalyst dose, pH, pulp yield, and biochemical composition organosolv pulps

Experimental conditions		pH liquor			Biochemical composition (%) ^c											
Biomass	Catalyst	Dose			Before ^a (–)	Final ^b (–)	Pulp yield (%) ^c	Polysaccharides				Lignin		Ash ^e Total		
		(M)	(N)	(%) ^c				Ara ^d	Xyl	Man	Gal	Glu	AIL		ASL	
Willow wood	H ₂ SO ₄	0	0	0.0	5.9	4.8	69.3	^f	12.4	1.7	0.4	47.5	24.6	0.9	1.6	89.0
		0.010	0.020	1.1	3.0	4.5	66.9		10.3	1.6	0.2	49.0	24.4	0.7	2.1	88.4
		0.025	0.050	2.7	2.3	3.4	57.5		5.5	1.5	0.1	53.3	26.4	0.4	2.6	89.7
	HCl	0	0	0.0	5.8	4.9	70.0		12.4	1.6	0.4	45.2	27.3	1.0	2.8	90.7
		0.020	0.020	0.8	2.4	4.0	55.1		5.8	1.3	0.1	52.9	26.8	0.5	2.5	90.0
		0.050	0.050	2.0	1.8	2.8	44.0		1.1	1.0	0.1	60.1	28.3	0.3	2.8	93.8
	MgCl ₂	0		0.0	5.8	4.9	68.7		11.8	1.7	0.4	47.2	25.1	0.9	2.1	89.1
		0.010		1.1	4.8	4.6	63.0		8.7	1.7	0.2	50.9	25.2	0.7	2.2	89.6
		0.025		2.7	4.8	4.4	60.2		7.4	1.6	0.1	52.6	25.0	0.6	2.8	90.1
Wheat straw	H ₂ SO ₄	0	0	0.0	6.5	4.9	74.4	0.5	20.6		0.3	50.2	14.1	1.1	5.9	92.8
		0.010	0.020	1.1	2.8	4.0	62.3	0.2	14.6		0.2	59.5	10.5	0.7	6.7	92.5
		0.025	0.050	2.7	2.2	2.6	43.8	0.1	1.6		0.2	69.5	9.9	0.5	10.9	92.6
	HCl	0	0	0.0	6.3	4.9	78.5	0.7	20.3		0.5	48.4	15.6	1.1	5.6	92.3
		0.020	0.020	0.8	2.5	3.4	52.2	0.1	6.6		0.3	67.5	10.3	0.5	8.3	93.7
		0.050	0.050	2.0	2.0	2.2	35.0		0.6		0.2	65.6	14.0	0.3	12.9	93.7
	MgCl ₂	0		0.0	6.5	4.9	76.4	0.5	20.1		0.4	48.7	15.0	1.1	6.3	92.0
		0.010		1.1	5.9	4.4	63.7	0.1	15.0		0.3	55.7	13.1	0.8	7.0	91.9
		0.025		2.6	5.6	4.2	58.8	0.0	11.6		0.2	61.5	11.5	0.6	7.0	92.4

^a pH of slurry at room temperature at start of organosolv experiment.

^b pH of organosolv liquor at room temperature after process.

^c w/w dry biomass.

^d Abbreviations used: arabinan (Ara), xylan (Xyl), mannan (Man), galactan (Gal), glucan (Glu), acid insoluble lignin (AIL), and acid soluble lignin (ASL).

^e Ash part of hydrolysis residue (acid soluble part of ash not included).

^f Empty cell: concentration below detection limit in hydrolysate. The concentration of rhamnose was below detection limit in all hydrolysates.

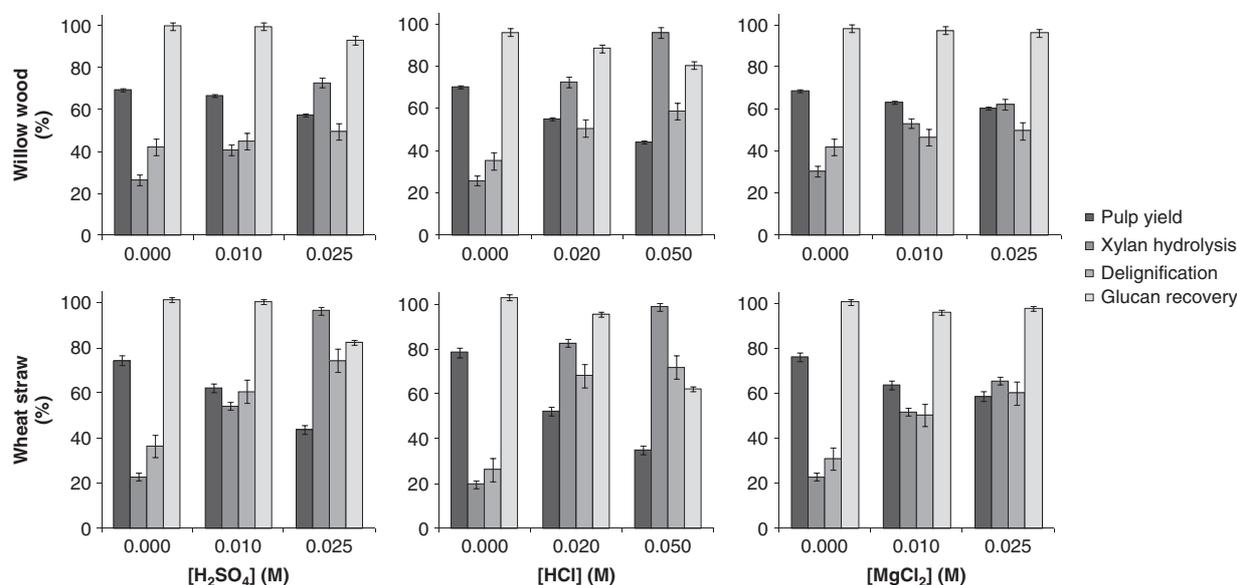


Figure 3. Effect of catalyst dose on pulp yield and fractionation of willow wood (top) and wheat straw (bottom).

Figure 4 compares the effect of H₂SO₄ and HCl on organosolv fractionation of both feedstocks based on the final pH of the organosolv liquor. Figure 4 shows that the pulp yield and xylan hydrolysis degree are well correlated with the pH of the organosolv liquor. The correlation between pH of the organosolv liquor and delignification degree is worse (especially for wheat straw).

Figure 4 shows no clear differences between both acid catalysts. Therefore, the effect of the acid catalysts seems to be caused solely by their acidity rather than by the type of anion. Table 2 shows that the addition of HCl reduces the pH of the organosolv liquor to a substantially larger extent than H₂SO₄ at equal normality. Therefore, it can be concluded that acid strength (pKa) of both

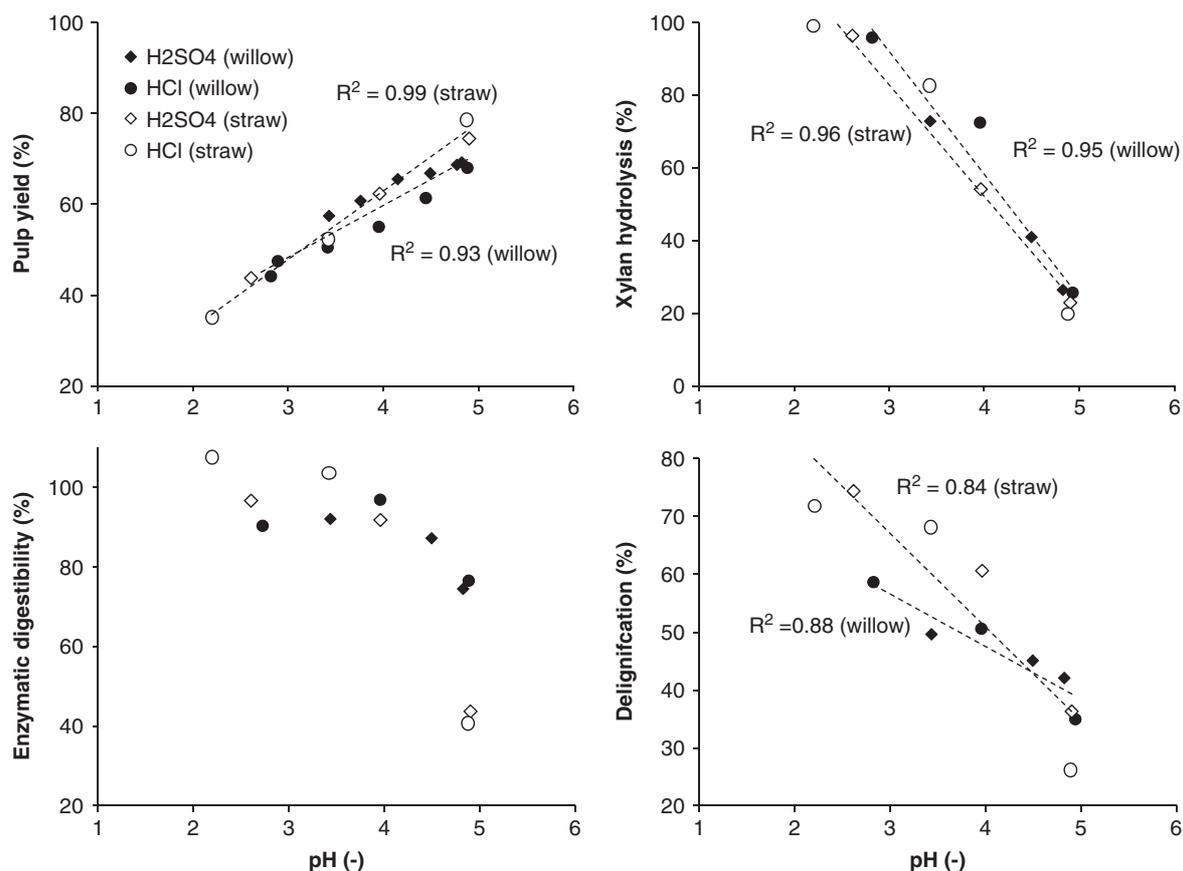


Figure 4. Effect of acid catalysts on fractionation and enzymatic digestibility as function of the final pH of the organosolv liquor.

acids differs. In an aqueous environment both H₂SO₄ and HCl are strong acids resulting in complete dissociation. However, in aqueous ethanol the second dissociation step of H₂SO₄ seems to be weak resulting in higher pH values of the organosolv liquor than for HCl at equal normality (Table 2). Overall, addition of HCl has a larger effect on the pulp yield (Fig. 2 and Table 2) and fractionation (Fig. 3) than the addition of an equal dose of H₂SO₄, expressed in terms of normality.

Finally, the influence of an acid catalyst on xylan hydrolysis and delignification is more pronounced for wheat straw than for willow (Fig. 3). Table 2 shows that the final pH of any organosolv liquor from catalytic willow wood pretreatment is higher than that from wheat straw at a given acid dose. This difference is probably caused by the somewhat higher acid neutralization capacity of willow wood compared with that of wheat straw (see section 'feedstocks'). It seems that the mineral fraction of willow neutralizes the added acid catalyst to a larger extent, thereby reducing its effect on fractionation.

MgCl₂

The use of the third catalyst, MgCl₂, also increases xylan hydrolysis and delignification (Fig. 3). Table 2 shows that addition of MgCl₂ decreases the pH of the organosolv slurry at the start of an organosolv experiment. For both feedstocks, mixtures containing 0.025 mol L⁻¹ MgCl₂ have a pH of about 1 unit lower than those without any catalyst added. It remains unclear how a neutral salt like MgCl₂ lowers the pH, although Paszner and Cho hypothesized that adsorption of Mg²⁺ on the carboxylic acid groups of the

biomass causes release of protons.²⁷ On the other hand, the type of anion in Mg-salts has also been reported to play a role, with Cl⁻ as most effective anion tested.²² Moreover, the addition of a salt influences the ionic strength of the organosolv liquor. During MgCl₂-catalysed organosolv fractionation, the pH of the organosolv liquor decreases because of the formation of organic acids. In contrast, the pH of the organosolv liquor increases during acid-catalyzed organosolv fractionation due to neutralization by the ash component. Overall, during the total process the pH of the MgCl₂-catalyzed experiments remains relatively constant and high compared with acid-catalyzed experiments (Table 2).

Maximum fractionation using MgCl₂ has been obtained at its highest dose (0.025 mol L⁻¹ or ~2.7% w/w dry biomass). For willow wood, 62% xylan hydrolysis and 49% delignification have been obtained. For wheat straw, 66% xylan hydrolysis and 60% delignification. The use of MgCl₂ fractionates wheat straw to a smaller extent than the use of the acid catalysts (maxima obtained with acid catalysts: 99% xylan hydrolysis, and 74% delignification). This results in a lower glucan content of the pulp obtained (up to 70% and 62% for acid catalysts and MgCl₂, respectively). In contrast to the acid catalysts, no significant glucan hydrolysis is observed at any dose of MgCl₂ added. The glucan recovery at the highest catalyst dose is still 96% (willow wood) and 98% (wheat straw).

MgCl₂ seems to have a less pronounced effect than acid catalysts on xylan hydrolysis of willow wood compared with delignification. For example, willow wood has been delignified for about 50% using either 0.025 mol L⁻¹ H₂SO₄, 0.02 mol L⁻¹ HCl, or 0.025 mol L⁻¹ MgCl₂. At these catalyst concentrations, the xylan

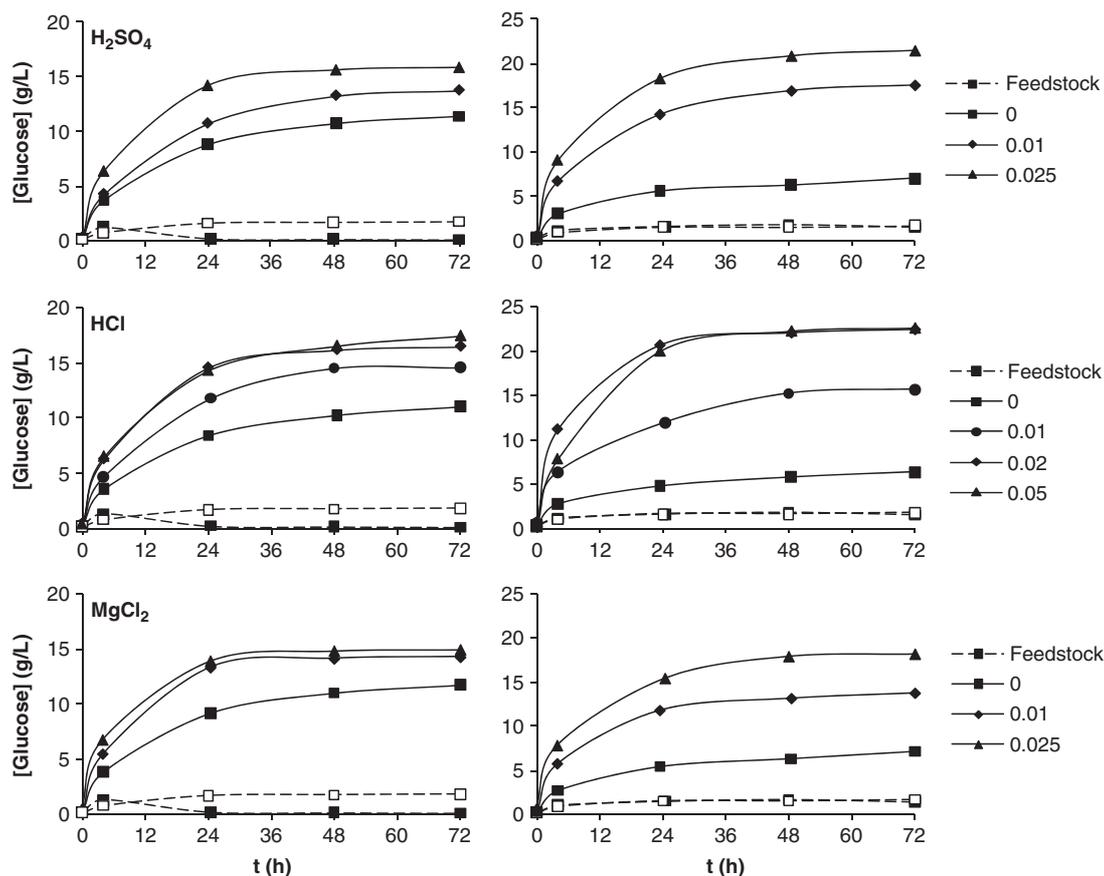


Figure 5. Average glucose concentration during enzymatic hydrolysis of raw materials and pretreated biomass substrates ($n = 2$); Biomass: willow wood (left) and wheat straw (right). Antibiotic applied: PenStrep (solid symbols) or sodium azide (open symbols).

hydrolysis degree is 73% for both H_2SO_4 and HCl and only 62% for $MgCl_2$. This difference in behaviour between $MgCl_2$ and acid catalysts has also been mentioned by others for softwoods.^{19,22} For wheat straw, this apparent difference in effects of both types of catalysts could not be observed in the data presented.

Enzymatic hydrolysis of (pretreated) biomass

The enzymatic hydrolysis profiles of the feedstocks and selected pretreated materials are given in Fig. 5. Glucose concentrations follow the typical patterns observed during enzymatic cellulose hydrolysis,^{15,16,29,30} except for the willow wood raw material. During enzymatic hydrolysis of this substrate, the glucose concentration drops unexpectedly after 4 h in spite of the PenStrep antibiotic applied to prevent the growth of sugar-consuming bacteria. Repetition of the experiment as well as doubling the amount of antibiotic gave similar results. Therefore, the enzymatic hydrolysis of both feedstocks was repeated using 0.02% w/v sodium azide instead of PenStrep according to NREL/TP-510-42629²⁵. No sugar consumption occurred when NaN_3 was used (Fig. 5). For both feedstocks, the maximum glucose concentrations (obtained using NaN_3) correspond to an enzymatic digestibility of 18% (willow wood) and 15% (wheat straw). No indications of sugar consumption were observed during enzymatic hydrolysis of any other substrate.

Non-catalytic organosolv

Organosolv pretreatment improves the enzymatic digestibility of both willow wood and wheat straw under all conditions studied.

Following non-catalytic organosolv pretreatment, maximum glucose concentrations during enzymatic hydrolysis increases from 2 g L^{-1} for non-treated willow wood to 11 g L^{-1} for pretreated willow wood (Fig. 5). This corresponds to an average enzymatic digestibility of the pulps of $76.2 \pm 1.6\%$ glucose based on the glucan content of the pulp ($74.0 \pm 2.5\%$ based on the raw material) (Fig. 6). Non-catalytic organosolv pretreatment of wheat straw increases enzymatic glucose concentrations from 2 g L^{-1} (raw material) to 7 g L^{-1} . The corresponding glucose yields are $43.4 \pm 2.5\%$ (pulp-based) and $44.1 \pm 2.0\%$ (raw material-based). The substantially higher enzymatic digestibility of pretreated willow wood compared with wheat straw resulting from non-catalytic organosolv is opposite to the relative lignin content of both pulps. The average lignin content of non-catalytically pretreated willow wood is 26% versus 16% for wheat straw (Table 2). Therefore, the higher enzymatic digestibility of pretreated willow wood seems to be caused by other properties of the pulp such as the lower xylan content of pretreated willow wood compared with wheat straw (12 and 20% w/w dry biomass, respectively). Finally, the spread in enzymatic digestibility results of non-catalytic organosolv pretreatment has been used as an estimate of the experimental error made in Fig. 6.

Catalytic organosolv

The use of catalysts further improves the enzymatic digestibility of both feedstocks (Figs 5 and 6). Maximum glucose concentrations during enzymatic hydrolysis increase to $14\text{--}17\text{ g L}^{-1}$ for willow wood and $16\text{--}23\text{ g L}^{-1}$ for wheat straw following catalytic organosolv pretreatment (Fig. 5). Maximum glucose concentrations were

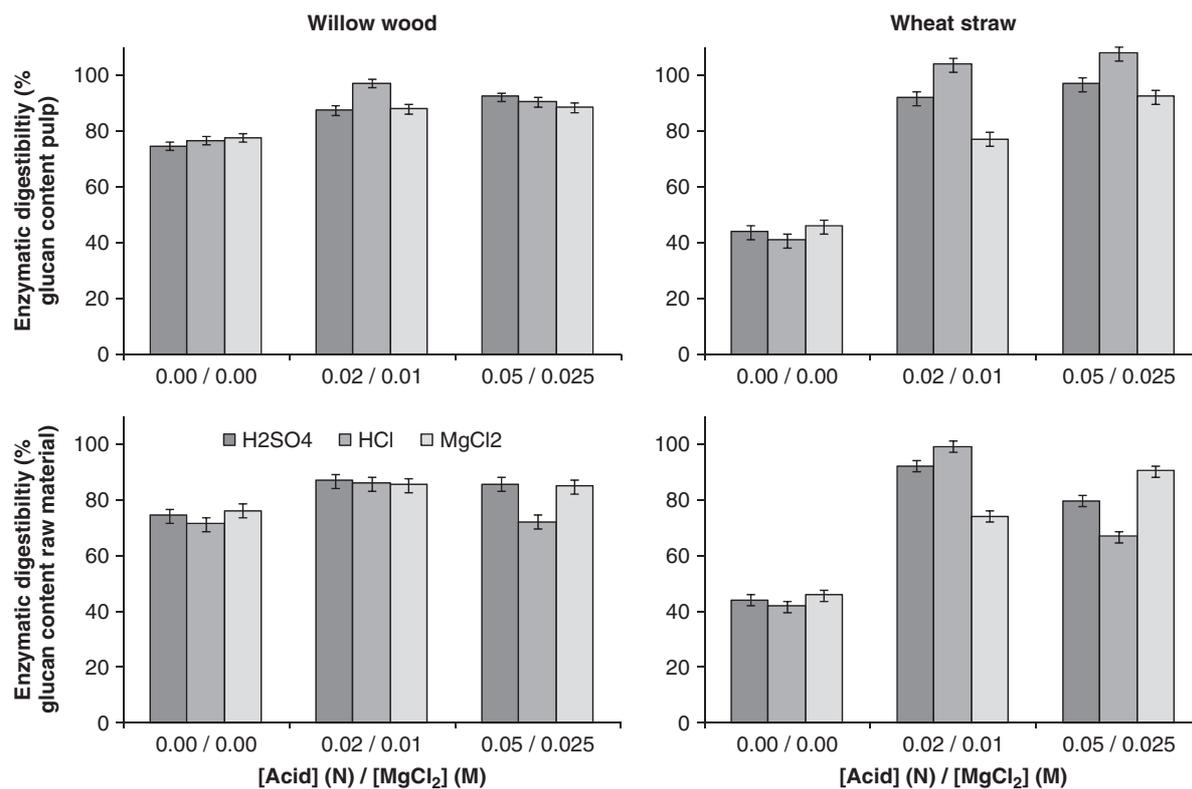


Figure 6. Maximum enzymatic digestibility based on glucan content of either the pretreated biomass (top) or the biomass feedstock (bottom).

obtained with 0.05 mol L⁻¹ HCl for both raw materials (17 g L⁻¹ for willow wood and 23 g L⁻¹ for wheat straw).

The corresponding glucose yields by enzymatic hydrolysis are 87–97% (willow wood) and 77–108% (wheat straw) based upon the glucan content of pretreated materials (Fig. 6, top). For both feedstocks, maximum enzymatic glucose yields were obtained with HCl followed by H₂SO₄ and MgCl₂. Remarkably, wheat straw shows a higher digestibility after catalytic organosolv than willow wood at all catalyst doses (except 0.01 mol L⁻¹ MgCl₂), while after non-catalytic organosolv wheat straw has a substantially lower enzymatic digestibility. For the acid catalysts, their larger influence on the fractionation of wheat straw compared with willow wood because of differences in acid neutralization capacity, as discussed above, is probably contributing to substantial enhancement of the enzymatic digestibility of wheat straw. Table 2 shows that the xylan content of pretreated wheat straw is more strongly reduced by acid catalysts than that of willow wood. In the case of raw materials and pulps resulting from non-catalytic organosolv, the xylan content of wheat straw is higher than that of willow wood. In the case of pulps resulting from catalytic organosolv in which a high dose of acid was applied, the xylan content of wheat straw is lower than that of willow wood. The enzymatic digestibility of pretreated willow wood seems to reach an optimum at a final pH of the organosolv liquor of 4.0 (Fig. 4). At lower pH values, the enzymatic digestibility decreases, in contrast to that of wheat straw.

The enzymatic digestibility of wheat straw after pretreatment with 0.02 or 0.05 mol L⁻¹ HCl slightly exceeds the theoretical maximum of 100% (104 and 108%, respectively). This might be due to an analytical overestimation of the glucose concentration in the hydrolysate. In the colorimetric method used, minor absorption of non-glucose components is known to occur, as was discussed in

earlier work.³⁰ For verification, the hydrolysates obtained after 72 h of enzymatic hydrolysis of the 0.02 and 0.05 mol L⁻¹ HCl pretreated wheat straw samples were also analyzed by HPAEC-PAD for their glucose concentration. It was found that the colorimetric method used overestimated the glucose concentrations in these samples by (only) 3 and 1%, respectively. Another effect that might play a role is the experimental error in the determination of the glucan content of the substrate.

Feedstock-based enzymatic glucose yields

The overall enzymatic glucose yield based on the composition of the feedstocks was determined taking into account glucan hydrolysis during organosolv fractionation (Fig. 6, bottom). Most significant deviations from the enzymatic glucose yield based on the composition of the substrate occur at high acid catalyst doses (i.e. doses at which significant glucan hydrolysis occurred during organosolv). Most noticeably, the enzymatic digestibility of wheat straw at 0.05 mol L⁻¹ HCl expressed on the glucan content of the raw material is only 67% compared to 108% expressed on the glucan content of the pulp due to the 38% glucan hydrolysis during organosolv fractionation. The maximum overall glucose yield obtained is 87% (willow wood, 0.01 mol L⁻¹ H₂SO₄) and 99% (wheat straw, 0.02 mol L⁻¹ HCl). For willow wood, other catalyst doses give similar yields, i.e. 0.02 mol L⁻¹ HCl (86%), 0.05 mol L⁻¹ H₂SO₄ (86%) and 0.01 and 0.025 mol L⁻¹ MgCl₂ (85%). Therefore, no optimum catalyst can be selected for willow wood based on the overall enzymatic glucose yield. For wheat straw, the optimum enzymatic glucose yield of this study is comparable with, for example, yields reported in the literature using dilute (organic) acid pretreatment,³⁵ dilute H₂SO₄ impregnation followed by steam explosion,²⁴ or mild oxidation with hydrogen peroxide.²⁴

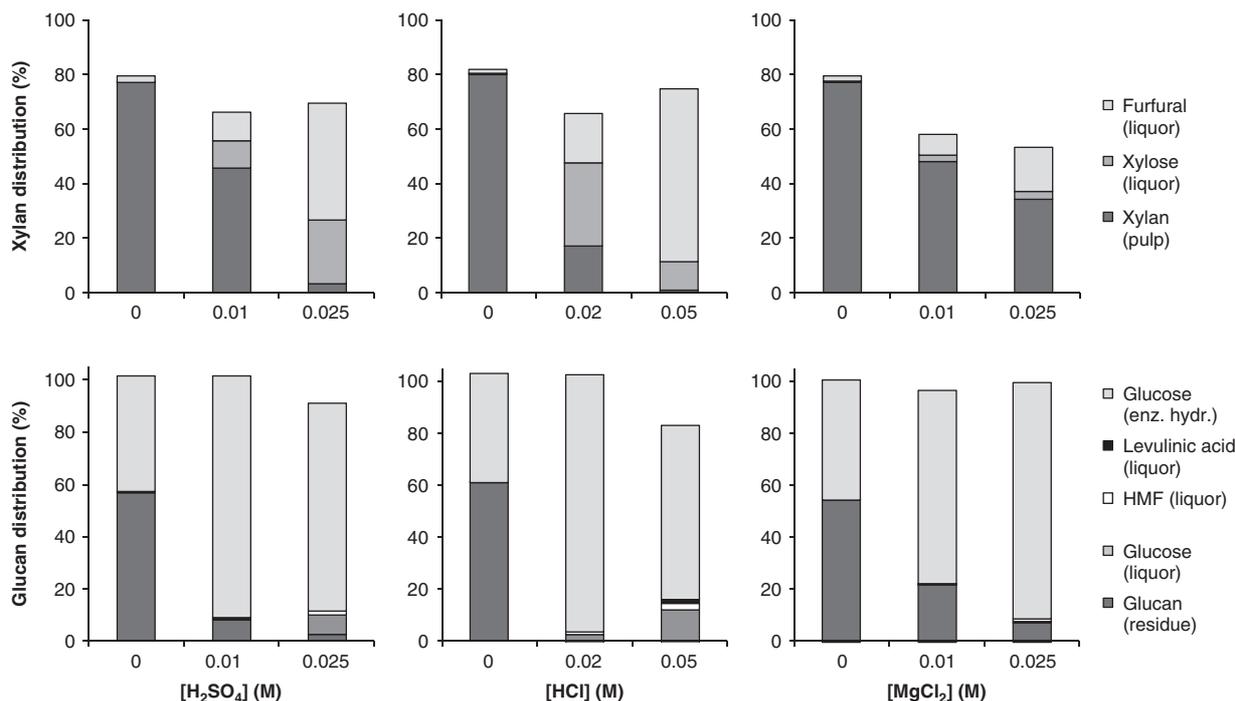


Figure 7. Product distribution xylan (top) and glucan (bottom) for wheat straw (mol%). Components included for xylan: xylose and furfural in organosolv liquor and xylan in organosolv pretreated wheat straw. For glucan: glucose, HMF and levulinic acid in organosolv liquor, glucose in enzymatic hydrolysate, and glucan in solid residue after enzymatic hydrolysis.

Xylanase activity

The Accellerase 1500 enzyme cocktail used also exhibits xylanase activity according to the supplier. In the literature a xylanase activity of 75 IU mL⁻¹ has been reported.³⁶ HPAEC-PAD analysis of the 72 h hydrolysate of the 0.02 mol L⁻¹ HCl pretreated wheat straw substrate resulted in 2.0 g L⁻¹ xylose and 91% enzymatic xylose yield based on the xylan content of the pulp. Since the xylan hydrolysis degree during organosolv fractionation at these conditions is 83%, the additional enzymatic xylose yield based on the composition of the feedstock is only 16%. In total, 99% of the xylan present in the wheat straw raw material is converted at these conditions.

Overall, it can be concluded that application of all three catalysts tested improves the susceptibility of both feedstocks for hydrolytic enzymes. Underlying mechanistic factors most likely include improved hemicellulose hydrolysis and enhanced delignification resulting in better cellulose accessibility for enzymes and less non-productive binding of enzymes to lignin.⁴

Yields sugar products wheat straw fractionation

Figure 7 shows the yields of glucan- and xylan-derived products after organosolv fractionation of wheat straw followed by enzymatic hydrolysis of the pulp (expressed in mol%). Optimum glucose yields were obtained at medium acid catalyst doses and the highest MgCl₂ dose. Maximum glucose yield in a single product stream occurs at 0.02 mol L⁻¹ HCl (99%, enzymatic hydrolysate). The highest yields of monomeric xylose in the organosolv liquor occur at 0.025 mol L⁻¹ H₂SO₄ (24%) and 0.02 mol L⁻¹ HCl (30%). Together with xylose formed during enzymatic hydrolysis of the 0.02 mol L⁻¹ HCl pretreated wheat straw the total xylose yield is 46%. In addition, 18% of the xylan is converted into furfural at this condition. Furfural is the major product from xylan in acid-catalyzed organosolv experiments (up to 63% at 0.05 mol L⁻¹ HCl).

The use of MgCl₂ leads to very low yields of monomeric xylose (max 3%).

For glucan, the balances are roughly closed except at severe hydrolysis conditions (0.05 N H₂SO₄ and HCl). At these conditions, substantial hydrolysis of cellulose occurs, probably with subsequent degradation of glucose in the organosolv liquor. Since the yields of the glucose derivatives HMF and levulinic acid are small (max 2.6 and 1.3% at 0.05 mol L⁻¹ HCl, respectively), HMF and glucose have probably reacted away and formed so-called humins or lignin condensation products.^{1,37} For xylan, balances are less complete than for glucose (losses between 18% and 47%). First of all, furfural formed has been reported to react further including formation of humins³⁷ and lignin-furfural condensation products,³⁰ especially at acidic conditions. For the non-acid catalysed experiments, oligomeric xylose fragments are probably present in significant amounts, as was discussed in earlier work.³⁰

Comparison of catalysts

As discussed above, the effect of the acid catalysts H₂SO₄ and HCl seems primarily due to their effect on the pH of the organosolv liquor rather than the type of acid. In addition, similar results in terms of fractionation degree and enzymatic hydrolysis yields have been obtained with both catalysts, especially for willow wood. Since H₂SO₄ has been found to be a weaker acid than HCl in aqueous ethanol, a higher dose of this catalyst is required than of HCl. On the other hand, H₂SO₄ is cheaper and less corrosive than HCl.

The use of MgCl₂ leads to lower delignification and xylan hydrolysis degrees than the acid catalysts. In addition, the enzymatic glucose yields obtained using MgCl₂ are slightly lower than those obtained using acid catalysts (within the range of process conditions tested). On the other hand, a benefit of MgCl₂ might be that it can yield similar enzymatic glucose yields to the

acid catalysts, while retaining more xylan in the pulp (i.e. lower xylan hydrolysis degree). For example, willow wood pretreated using $0.01 \text{ mol L}^{-1} \text{ MgCl}_2$ results in 85% overall enzymatic glucose yield at relatively low xylan hydrolysis (52%). A similar enzymatic glucose yield (86%) has been found for willow wood pretreatment using $0.02 \text{ mol L}^{-1} \text{ HCl}$, but at substantially higher xylan hydrolysis (83%). In a process scheme in which the pulp is enzymatically hydrolyzed and the resulting C5 and C6 sugars are fermented, this could potentially imply higher yields of bioethanol.

This study has focused on the effect of different catalysts on organosolv pretreatment of both a hardwood and a herbaceous crop at a single set of process conditions, such as temperature and solvent:water ratio. A next step would be to perform an optimization study taking into account all other major process variables in addition to catalyst dose. Final selection of the optimum catalyst should be performed based on an economic analysis of the process including cost of catalyst, additional costs caused by corrosive-resistant materials, and product values.

CONCLUSIONS

The effect of the catalysts H_2SO_4 , HCl , and MgCl_2 on ethanol-based organosolv pretreatment of willow wood and wheat straw was studied and compared. For willow wood, maximum enzymatic cellulose hydrolysis was obtained using $0.01 \text{ mol L}^{-1} \text{ H}_2\text{SO}_4$ as catalyst (87%, together with 73% xylan hydrolysis and 51% delignification). For wheat straw, the maximum enzymatic glucose yield obtained based on the glucan content of the feedstock was 99% (together with 83% xylan hydrolysis and 68% delignification). For comparison, non-catalytic organosolv at identical conditions yielded 74% and 44% enzymatic glucose yield based on the glucan content of the feedstock for willow wood and wheat straw, respectively (28% and 22% xylan hydrolysis, 40% and 31% delignification). The reverse in relative susceptibility of both feedstocks for enzymatic hydrolysis, comparing non-catalytic and acid catalyzed-organosolv pretreatment, seems to be due to a more pronounced reduction by catalysts of the xylan content of wheat straw than that of willow wood. The effect of the acid catalysts was found to be primarily due to their effect on the pH of the organosolv liquor rather than the type of anion. The use of MgCl_2 was found to enhance fractionation and enzymatic digestibility to a lesser extent than both acid catalysts. On the other hand, MgCl_2 seems to more selectively promote delignification and enzymatic digestibility of willow wood. In general, application of catalysts in organosolv pretreatment was found to improve fractionation and enzymatic digestibility, especially for wheat straw. Thus, the use of catalysts can contribute to achieving maximum utilization and valorization of lignocellulosic biomass in organosolv-based biorefineries.

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