

## Effect of ileo-rectal anastomosis and post-valve T-caecum cannulation on growing pigs

### 2. Blood variables and mineral balances

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In a long-term study nine ileo-rectally anastomosed (IRA) and seven post-valve T-caecum (PVTC)-cannulated pigs were compared with six intact pigs with regard to different blood variables, sodium and potassium retention and weights of selected organs. After surgery, apart from urea and K measured 13 weeks post-surgery, there were no differences in the blood variables between the PVTC-pigs and intact pigs. In IRA-pigs concentrations of creatinine ( $P < 0.01$ ), Na ( $P < 0.001$ ), base excess ( $P < 0.001$ ), pH ( $P < 0.01$ ) and bicarbonate ( $P < 0.001$ ) in blood were lower than those in intact pigs. At 13 weeks after surgery the blood K concentration in IRA-pigs was higher ( $P < 0.001$ ) than that in PVTC-pigs or intact pigs. At 6 weeks after surgery the blood urea concentration in IRA-pigs was higher ( $P < 0.001$ ) than that in intact and PVTC-pigs. At 13 weeks after surgery the urea concentration in PVTC-pigs was higher ( $P < 0.001$ ) than those in IRA-pigs or intact pigs. The Na ( $P < 0.01$  11 weeks after surgery) and ( $P < 0.05$  and  $P < 0.01$  5 and 11 weeks after surgery respectively) balances in IRA-pigs were lower than those in intact animals. Na retention was negative for IRA-animals 11 weeks after surgery. Na and K retentions were similar in PVTC-pigs and in intact pigs. The urinary: faecal excretion of Na differed slightly between PVTC-animals and intact animals. At 13 weeks after surgery there were no differences in organ weights between the PVTC-pigs and intact animals. In the IRA-pigs the weights of the liver ( $P > 0.05$ ), the kidneys ( $P > 0.05$ ) and the adrenal glands were higher ( $P < 0.001$ ) than those in the intact animals.

Ileo-rectal anastomosis: Post-valve T-caecum cannulation: Blood variables: Na and K balances: Pig

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The post-valve T-caecum (PVTC) cannula, as described by van Leeuwen *et al.* (1988), has been shown to provide reliable measurements of digestibilities at the distal ileum of pigs (Köhler *et al.* 1990; Van Leeuwen *et al.* 1991). Ileo-rectal anastomosis (IRA) has been proposed as an alternative technique for cannulation (Picard *et al.* 1984; Laplace *et al.*

1985; Souffrant *et al.* 1985; Green, 1988). Both methods may affect the physiological state of the animals. Digestibility as a physiological variable has to be determined in animals in a normal physiological state. Findings for growth performance, digesta flow and intestinal fermentation have been presented previously (Köhler *et al.* 1992*b*). The objective of the present study was to compare the recently developed PVTC-cannulation technique with the end-to-side IRA technique with regard to different blood variables, sodium and potassium balances and selected organ weights.

#### MATERIALS AND METHODS

Twenty-two crossbred castrates (Yorkshire × Dutch Landrace) with an average body-weight of 30 kg were used. Nine pigs were provided with an end-to-side IRA and seven pigs with a PVTC cannula. Details of surgical techniques and the experimental procedures have been reported previously (Köhler *et al.* 1992*b*). The experimental design is shown schematically in Fig. 1. The animals were fed at 2.4 times maintenance requirement for energy (Agricultural Research Council, 1981). The composition of the experimental diet is shown in Table 1.

##### *Blood samples*

At 6 and 13 weeks after surgery blood samples were obtained 3 h post-prandially by puncture of the external jugular vein. The following analyses in blood were carried out: Na, K, chloride, calcium, magnesium, urea, creatinine, pH, active bicarbonate and the base excess. In addition Na, urea and creatinine levels were determined 1 week before surgery.

##### *Analysis of blood samples*

Urea, creatinine, Na, K and Cl concentrations were measured in serum obtained after centrifugation of the blood samples at 3000 rev./min for 20 min.

Urea and creatinine concentrations were estimated using Biomérieux kits. Na and K were measured using a flame photometer (model 243, Instrumentation Laboratory, Cheltenham, PA, USA), and Cl using a Chlor-O-Counter (Laméris Instruments Bv, Utrecht, The Netherlands). Mg was measured in heparinized plasma using an atomic absorption spectrophotometer (model 305 B Perkin Elmer, Pomona, USA). Plasma Ca concentration was measured according to Willis (1960). pH, bicarbonate and base excess were measured in blood, sampled air-free in a syringe, kept at 0° in ice until analysed using a blood gas pH meter (ABL2 acid base laboratory radiometer, Analytical 175/Medical 175 Bagsvaerd, Copenhagen, Denmark). These analyses were carried out within 90 min of collection of the blood.

##### *Na and K balances*

At 5 and 11 weeks after surgery the Na and K balances were determined over a period of 5 d.

##### *Organ weights*

At 13 weeks after surgery the animals were slaughtered and the weights of the liver, kidneys and the adrenal glands as well as the circumference of the rectum were registered.

#### RESULTS

##### *Blood variables*

Values for blood variables are shown in Table 2. After surgery the blood creatinine contents of pigs fitted with an IRA were significantly lower than those in intact animals or in animals fitted with a PVTC cannula ( $P < 0.01$ ). Pre-operative urea concentrations were not different between the treatment groups. At 6 weeks after surgery urea levels in the IRA-animals were increased and higher ( $P < 0.001$ ) than in the other treatments. At 13 weeks

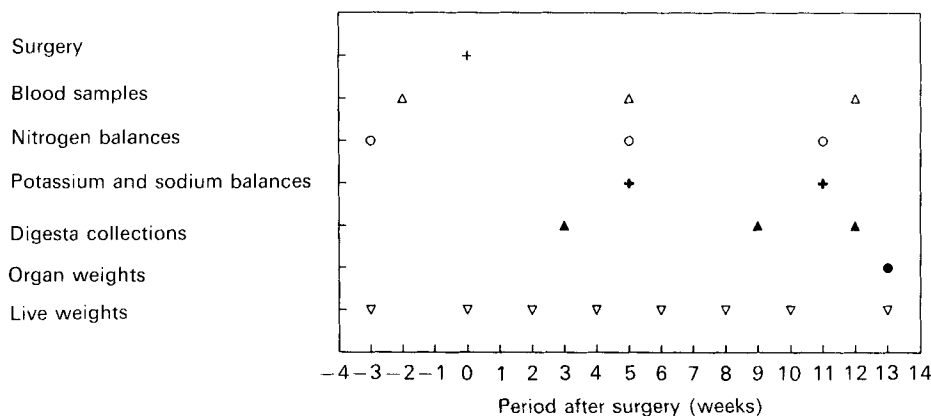


Fig. 1. Experimental design. For details of techniques and procedures, see pp. 306–307.

Table 1. *Composition of the experimental diet (g/kg)*

Ingredients:	
Maize	32.95
Barley	6.00
Wheat	10.50
Soya-bean meal, solvent extracted	22.50
Molasses	4.00
Potato pulp	9.20
Sugar-beet pulp	10.00
Animal fat	1.00
L-lysine hydrochloride	0.18
DL-methionine	0.12
Vitamins and minerals*	1.00
Ca(H <sub>2</sub> PO <sub>4</sub> ) <sub>2</sub> ·H <sub>2</sub> O	1.50
CaCO <sub>3</sub>	0.50
NaCl	0.30
Cr <sub>2</sub> O <sub>3</sub>	0.25
Chemical composition (g/kg dry matter):	
Analysed:	
Crude protein (nitrogen × 6.25)	199.0
Gross energy (MJ)	18.04
Sodium	1.6
Potassium	13.0
Calculated:	
Crude fibre	65.0
Chloride	1.9
Magnesium	1.6
Calcium	2.6

\* Contributed (mg/kg diet): retinol 2.7, cholecalciferol 45  $\mu$ g,  $\alpha$ -tocopherol 40, menadione 3, riboflavin 5, cobalamine 40  $\mu$ g, nicotinic acid 30, D-pantothenic acid 12, choline chloride 150, ascorbic acid 50, KI 500  $\mu$ g, CoSO<sub>4</sub>·7H<sub>2</sub>O 2.5, Na<sub>2</sub>SeO<sub>3</sub> 0.2, FeSO<sub>4</sub>·7H<sub>2</sub>O 0.40 g, CuSO<sub>4</sub>·5H<sub>2</sub>O 0.1 g, MnO<sub>2</sub> 0.07 g, ZnSO<sub>4</sub>·H<sub>2</sub>O 0.2 g. This mixture also supplied 20 mg virginiamycin/kg diet, the mixture was made up to 1 kg with ground maize.

after surgery blood urea levels for IRA-animals and intact pigs were similar; however, PVTC-cannulated animals had higher ( $P < 0.001$ ) levels compared with those for the other two treatments. At 6 and 13 weeks after surgery the blood Na levels of IRA-animals were lower than those of the intact animals ( $P < 0.001$  and  $P < 0.01$  respectively). At 13 weeks after surgery the blood K levels of the IRA-pigs were higher than those of the other animals

Table 2. *Biochemical characteristics in serum of pigs with ileo-rectal anastomosis (IRA), post-valve T-caecum (PVTC) cannula or intact intestinal tract*‡

Treatment group...	Period after surgery (weeks)	IRA	PVTC	Intact	SEM
Creatinine ( $\mu\text{mol/l}$ )	Presurgery	112.22	124.71	111.33	4.20
	6	118.78	145.71**	137.00	5.15
	13	127.11	149.71**	148.33**	4.78
Urea (mmol/l)	Presurgery	3.75	3.66	3.92	0.19
	6	6.77	3.97***	4.53***	0.33
	13	6.02	8.36***	6.37	0.32
Sodium (mmol/l)	Presurgery	146.78	147.57	147.17	0.65
	6	140.11	150.71***	148.17***	1.34
	13	139.33	144.71**	146.33**	1.39
Potassium (mmol/l)	6	5.79	5.50	5.00	0.25
	13	5.61	5.09***	4.75†††	0.09
	6	95.11	99.86	98.50	1.37
Chlorine (mmol/l)	13	92.22	91.86	95.00	1.31
	6	2.60	2.53	2.60	0.03
	13	2.60	2.53	2.60	0.04
Calcium (mmol/l)	6	0.84	0.87	0.87	0.02
	13	0.83	0.90	0.85	0.02
	6	1.19	3.14	5.18***	0.50
Base excess (mmol/l)	13	-0.96	5.24**	6.50**	1.37
	6	7.32	7.31	7.37	0.41
	13	7.26	7.37**	7.38**	0.03
Active bicarbonate (mmol/l)	6	28.63	32.37***	32.30***	0.61
	13	27.89	32.36**	33.88**	1.04

Mean values were significantly different from those for IRA-pigs: \*\*  $P < 0.01$ , \*\*\*  $P < 0.001$ .

Mean values for intact pigs were significantly different from those for PVTC-pigs and IRA-pigs: †††  $P < 0.001$ .

‡ For details of techniques and procedures, see pp. 306–307.

( $P < 0.001$ ). In PVTC-pigs blood K levels were intermediate relative to the other two treatments and significantly higher ( $P < 0.001$ ) than levels for intact pigs. No significant differences between treatments were observed for the blood Cl, Ca and Mg contents. At 6 weeks after surgery the blood base excess of IRA-pigs was ( $P < 0.001$ ) lower than that of intact pigs. The PVTC-animals had intermediate levels between IRA- and intact animals. At 13 weeks after surgery the blood base excess of IRA-pigs was at a very low level and significantly different ( $P < 0.01$ ) from that of the other two treatments. At 13 weeks after surgery serum pH for IRA-animals differed ( $P < 0.01$ ) from that for the other two treatments. At both 6 and 13 weeks post-operatively the amounts of active bicarbonate for IRA-pigs were lower ( $P < 0.001$  and  $P < 0.01$  respectively) than those for PVTC-pigs and intact pigs.

#### *Na balance*

The results of the Na balances are given in Table 3. For the intact and PVTC-cannulated pigs the Na retention (mg/d) decreased with time but remained slightly positive. At 5 weeks after surgery the Na retention (mg/d) of IRA-pigs was in the same range as values obtained for the other treatments. At 11 weeks after surgery the Na balance of the IRA-pigs was lower ( $P < 0.01$ ) than those of the other treatments. At both 5 and 11 weeks the Na retention of PVTC-animals was similar to that of intact pigs. Table 3 also shows that in intact and PVTC-cannulated animals the amount of Na excreted in the urine was higher than that excreted in the faeces. In IRA-animals the renal: intestinal Na ratio was reversed, which finally resulted in a negative ileal digestibility for Na in IRA-pigs.

Table 3. Sodium balances in pigs with ileo-rectal anastomosis (IRA), post-valve T-caecum (PVTC) cannula or intact intestinal tract†‡

Treatment group...	Period after surgery (weeks)	IRA	PVTC	Intact	SEM
Intake (mg)	5	3015	2078***	2160***	36.74
	11	3314	2759***	2924***	46.17
Excretion in faeces (mg)§	5	2319	512***	376***	100.85
	11	3672	1065***	960***	161.33
Excretion in urine (mg)	5	32	867***	1122***	66.79
	11	48	1206***	1575††	87.57
Retention (mg)	5	663	699	662	117.06
	11	-407	488**	390**	181.48
Retention (%)	5	21.9	33.8	30.7	4.43
	11	-12.4	17.9**	13.3**	5.82
Total digestibility (%)	5	23.0	75.5***	82.6***	3.61
	11	-10.9	61.4***	67.2***	4.99

Mean values were significantly different from those for IRA-pigs: \*\*  $P < 0.01$ , \*\*\*  $P < 0.001$ .

Mean values for intact pigs were significantly different from those for PVTC-pigs and IRA-pigs: ††  $P < 0.01$ .

† For details of techniques and procedures, see pp. 306–307.

§ In PVTC-pigs and in intact pigs, excretion in faeces; in IRA-pigs, excretion in digesta.

Table 4. Potassium balances in pigs with ileo-rectal anastomosis (IRA), post-valve T-caecum (PVTC) cannula or intact intestinal tract†‡

Treatment group...	Period after surgery (weeks)	IRA	PVTC	Intact	SEM
Intake (mg)	5	13750	17066***	17776***	301.64
	11	16911	23652***	25069***	395.59
Excretion in faeces (mg)‡	5	9050	2316***	2280***	332.25
	11	11290	3496***	3372***	412.05
Excretion in urine (mg)	5	2711	11752***	11922***	392.86
	11	3400	15837***	16847***	610.38
Retention (mg)	5	1989	2998	3574*	353.06
	11	2221	4319**	4850**	515.74
Retention (%)	5	14.38	17.51	20.00	2.08
	11	12.94	18.21	19.32	2.27
Total digestibility (%)	5	34.02	86.47***	87.14***	2.51
	11	32.99	85.17***	86.55***	2.45

Mean values were significantly different from those for IRA-pigs: \*  $P < 0.05$ , \*\*  $P < 0.01$ , \*\*\*  $P < 0.001$ .

† For details of techniques and procedures, see pp. 306–307.

‡ In PVTC-pigs and in intact pigs, excretion in faeces; in IRA-pigs, excretion in digesta.

#### K balance

The K balances (Table 4) show differences similar to those for Na balances. The faecal apparent digestibility of K in the PVTC-cannulated and intact pigs was about 86%. The intestinal utilization of K in IRA-pigs was 34%. As for Na, the renal:intestinal K loss in IRA-animals did not follow the pattern found for intact and PVTC-animals. The amount of K excreted in urine of intact and PVTC-pigs was higher than that excreted in the faeces ( $P < 0.001$ ). In IRA-pigs more K was excreted in the digesta than in the urine. At 5 weeks after surgery K retention (mg/d) of IRA-pigs was lower ( $P < 0.05$ ) than that of intact

Table 5. *Organ weights (g/kg live weight<sup>0.75</sup>) of pigs with ileo-rectal anastomosis (IRA) or post-valve T-caecum (PVTC) cannula in comparison with intact pigs of the same age 13 weeks after surgery†*

Treatment group...	IRA	PVTC	Intact	SEM
Liver	13.18	12.24	12.68	1.34
Kidneys	3.40	2.80	2.72	0.18
Adrenal glands (mg)	93.00	60.00***	53.33***	0.01
Circumference of rectum (mm)	149.4	99.3***	113.3***	6.8

Mean values were significantly different from those for IRA-pigs: \*\*\*  $P < 0.001$ .

† For details of techniques and procedures, see pp. 306–307.

Table 6. *Creatinine content in blood of pigs*

mg/l	$\mu\text{mol/l}$	Reference
9.0–13.0	80–100	Wrogemann & Holtz (1972)*
22.5–23.7	199–209	Tumbleson <i>et al.</i> (1972)*
15.9–18.0	141–159	Steger (1976)†
1.15	102	Gregor (1979)*
16.0–16.6	142–147	Witkowska (1979)†
	111–150	Present study†

\* Mini pigs.

† Growing pigs.

animals. At 11 weeks after surgery the K retention (mg/d) of PVTC- and intact animals was higher ( $P < 0.01$ ) than that of IRA-pigs. K retention expressed as a percentage of intake shows no differences between the treatments.

#### *Organ weights*

Table 5 shows the relative weights of the liver, the kidneys and the adrenal glands at 13 weeks after surgery. Weights are expressed in g/kg live weight<sup>0.75</sup>. For the IRA-pigs the weights of the organs were higher than for the other animals. For the adrenal glands these differences were significant ( $P < 0.001$ ). In addition to the organ weights the circumference of the rectum was measured. In IRA-animals the circumference of the rectum was larger ( $P < 0.001$ ) than that in PVTC- or intact animals.

#### DISCUSSION

Steger *et al.* (1976) reported that the blood creatinine content of growing pigs is positively correlated with the weight of the animals. In agreement with this statement, Duggal & Eggum (1978) reported that the daily creatine and creatinine excretion of growing pigs was positively related to body-weight and to nitrogen retention. In addition, Murlin *et al.* (1953) concluded from their results that high urinary creatinine excretion indicated high N utilization. It may be assumed from these three studies that high blood creatinine content may be associated with high urinary creatinine excretion; furthermore it may also be related to high N utilization and high muscle mass. Only a limited amount of information is available in the literature on blood creatinine levels of pigs. These findings are summarized in Table 6. The present results, which were in the range 111–149  $\mu\text{mol/l}$  (Table 2), are in agreement with corresponding values from the literature.

Table 7. Urea content in blood of pigs

mg/l	mmol/l	Reference
120-133	2.00-2.22	Tumbleson <i>et al.</i> (1972)*
268-377	4.47-5.62	Steger <i>et al.</i> (1976)
107-387 (Urea-nitrogen)	3.83-13.83	Gregor (1979)*
	4.50-6.20	Bolduan & Schneider (1986)
300-409	5.00-6.82	Mosenthin <i>et al.</i> (1988)
	4.20-6.35	Münchow <i>et al.</i> (1989)
277.7	4.63	Wittenburg <i>et al.</i> (1989)
	3.66-8.43	Present study

\* Mini pigs.

A lower creatinine concentration ( $P < 0.01$ ) and lower daily live weight gain ( $P < 0.01$ ) was found for IRA-pigs than for PVTC- and intact pigs. The IRA-pigs also showed a lower N retention ( $P < 0.001$ ; Köhler *et al.* 1992*b*). Thus, our findings for IRA-pigs were in accordance with values from the literature for pigs with a reduced live weight, daily live weight gain and N utilization.

Berschauer (1977) found a clear negative correlation between the extent of protein utilization and blood urea content. Berschauer (1977) measured blood urea in pigs fed on diets differing in energy:protein ratio and concluded that an energy deficiency results in higher blood urea concentrations.

Bolduan & Schneider (1986) reported that the normal blood urea content of pigs ranges from 4 to 7 mmol/l serum. About 90% of urea is derived from deamination of amino acids after absorption and 10% is derived from ammonia originating from microbial activity and subsequent deamination in the intestine. Blood urea contents reported in the literature are given in different units of measurement. Table 7 gives a brief summary of these results as well as the corresponding values expressed in mmol/l. Our values are in the same range as those reported in the literature. There were significant differences ( $P < 0.001$ ), however, between the treatments in our experiment (Table 2).

The blood urea content also depends on: (a) potential rate of protein gain (breed, sex, age); (b) protein quality (the composition of the amino acids); (c) level of protein intake; (d) energy intake. In the present experiment all these factors were standardized by assessing feed intake on a unit metabolic body-weight basis. The blood urea content of intact pigs and PVTC-cannulated pigs increased with time. This is possibly related to N retention and growth intensity which decreased with time (Köhler *et al.* 1992*b*). In IRA-pigs the blood urea content during the 6th week after surgery was higher ( $P < 0.001$ ) than that in intact or PVTC-cannulated pigs. These findings are in accordance with both the reduced N retention and growth in the IRA-pigs. Rerat (1986) concluded that in intact pigs the amount of urea which is taken up by the intestinal tract and excreted as microbial protein in the faeces was similar to the urinary excretion of urea. This recycled urea is excreted as microbial protein in the faeces. For pigs fitted with an IRA, urea excretion as microbial protein is assumed impossible unless there is compensatory fermentation in the distal ileum or in the rectum, or both. Urea could also be excreted unchanged in ileal digesta, because Mosenthin *et al.* (1988) reported a higher urea concentration in ileal digesta after intravenous infusion of urea. Additional findings (T. Köhler, M. W. A. Verstegen and R. Mosenthin, unpublished results) showed that the N content of digesta from IRA-pigs was higher than that of ileal digesta from PVTC-pigs. Thus, the composition of ileal digesta

from IRA-animals is not similar to that of PVTC-animals. Also, the amounts of volatile fatty acids and diaminopimelic acid (DAPA) in the digesta of IRA-pigs were increased and were higher ( $P < 0.05$  and  $P < 0.001$  respectively) than those in PVTC-animals (Köhler *et al.* 1992*b*). This shows that there is increased microbial activity in the ileum or rectum, or both, in IRA-pigs. As a consequence more N might be excreted because some urea may have been utilized by micro-organisms. This could also explain the higher ( $P < 0.001$ ) concentration of N in the digesta of the IRA-animals compared with the PVTC-pigs. It may indicate a compensatory change of function in the ileum or rectum, or both, in the IRA-animals. A compensatory change in digestive function in IRA-pigs was also suggested by values obtained for the dry matter contents of the digesta. The ileal dry matter content for IRA-animals was higher ( $P < 0.001$ ) than that of ileal digesta for PVTC-pigs (Köhler *et al.* 1992*a*). In addition to these findings, the larger ( $P < 0.001$ ) circumference of the rectum (Table 5) and the irregular digesta outflow (Köhler *et al.* 1992*b*) indicate an increased transit time for digesta in the rectum. This may result in extra water absorption.

Alcantara *et al.* (1980) reported that pigs fed on a low-Na diet had a low blood Na concentration and also had an increased blood K concentration. There are reports that the ileal digestibility of Na is negative and that Na is absorbed in the large intestine (Partridge, 1978; Drochner, 1984; Partridge *et al.* 1986; Den Hartog *et al.* 1988). In IRA-pigs, in the absence of absorption from the large intestine, lower utilization of Na from the feed can be assumed, i.e. the absence of Na absorption in the large intestine may have the same effects as a low-Na diet. This would explain the lower ( $P < 0.001$  and  $P < 0.01$ ) Na concentration in the blood of IRA-pigs at 6 and 13 weeks after surgery respectively (Table 2). In addition, the blood K concentration of IRA-pigs was higher than those of the other treatments. Hyperkalaemia has been reported as a typical manifestation of Na deficiency (Black, 1960). Our findings in IRA-animals show the same tendencies as those reported by Alcantara *et al.* (1980) who fed low-Na diets. Münchow *et al.* (1989) reported serum Cl concentrations in the range 97.0–104.0 mmol/l for pigs fed on different diets. In accordance with these values, our findings (91.9–99.9 mmol/l) were in the same range and they did not differ between treatments. Hennig *et al.* (1986, 1988) reported Ca concentrations in serum of intact and anastomosed pigs in the range 2.59–2.94 mmol/l and Münchow *et al.* (1989) reported 2.52–2.73 mmol Ca/l serum. In the present experiment similar Ca concentrations were found with no differences between treatments.

For Mg there were no differences between the groups and the results were comparable with values reported in the literature. Nuoranne (1983) reported a normal post-absorptive Mg concentration of 0.80–0.96 mmol/l serum; this is in accordance with our values (0.83–0.90 mmol/l).

The IRA-technique had a considerable effect on the base excess and the bicarbonate concentration; both variables were lower ( $P < 0.01$  and  $P < 0.001$  respectively) in pigs provided with an IRA. Patience *et al.* (1987) reported that reducing the dietary electrolyte balance decreased the serum pH and bicarbonate concentration of pigs. In accordance with these findings IRA-animals had a lower ( $P < 0.01$ ) blood pH 13 weeks after surgery. Based on these findings a metabolic acidosis as a result of the reduced electrolyte balance can be expected for IRA-pigs. Findings for PVTC-cannulated pigs were similar to those for intact pigs.

In IRA-animals a significantly lower growth and N retention were observed (Köhler *et al.* 1992*b*). Growth, N retention and protein utilization depend on the level of energy intake. Na is an important element in the active absorption of different nutrients. Therefore, negative absorption of Na may be detrimental to the rate of gain. To compensate for the negative Na balance the IRA-pigs were supplied orally with extra electrolyte solution mixture, as described by Hennig *et al.* (1986). In addition, these authors



Table 8. Sodium contents in urine (mg/l) 5 and 11 weeks after surgery in pigs with ileo-rectal anastomosis (IRA), post-valve T-caecum (PVTC) cannula or intestinal intact tract\*

Treatment group... Period after surgery (weeks)	IRA	PVTC	Intact	SEM
5	15.00 <sup>a</sup>	371.60 <sup>b</sup>	416.00 <sup>b</sup>	17.70
11	12.38 <sup>a</sup>	336.00 <sup>b</sup>	406.83 <sup>c</sup>	17.67

<sup>a,b,c</sup> Mean values in the same row with unlike superscript letters were significantly different  $P < 0.001$ .

\* For details of techniques and procedures, see pp. 306–307.

Table 9. Sodium contents in ileal digesta (g/kg dry matter) 3, 9 and 12 weeks after surgery in pigs with ileo-rectal anastomosis (IRA) or post-valve T-caecum (PVTC) cannula

Treatment group... Period after surgery (weeks)	IRA	PVTC	SEM
3	10.2	30.0*	0.7
9	9.6	28.4*	0.6
12	9.5	31.9*	0.4

Mean values were significantly different from those for IRA-pigs: \*  $P < 0.001$ .

proposed a sugar–starch supplement for IRA-animals to compensate for the absence of energy absorption from the large intestine. They reported similar N utilization and N balance in IRA-pigs and intact pigs (Hennig, 1988). In the present investigation an electrolyte solution mixture, as described by Hennig *et al.* (1986), was added to the diet (200 ml/20 kg live weight per d), but the sugar–starch supplement was not provided. Our findings showed that the addition of the electrolyte solution to the diet of the IRA-pigs was not sufficient to achieve the same level of N retention as intact or PVTC-cannulated animals. In IRA-pigs Na metabolism in the absence of the large intestine was different from that of the other groups (Table 3). Faecal Na digestibility of the intact and PVTC-cannulated pigs ranged from 61.4 to 82.6%. In IRA-pigs total Na digestibility was 23.0 and –10.9%. Clarke *et al.* (1967) reported chronic Na depletion in humans fitted with an ileostomy. Their investigation by means of intravenous saline (9 g sodium chloride/l) infusion revealed an intestinal adaptation to conserve Na. Thus, after Na infusion Na concentration in ileal digesta increased and K concentration decreased. This post-infusion response suggests Na conservation in exchange for K. In our experiment Na loss in the urine was lowest in IRA-pigs. To compensate for the absence of Na absorption in the large intestine of IRA-pigs the urinary Na loss was about thirty times lower than that in intact pigs (Table 8). In humans Clarke *et al.* (1967) concluded that intestinal adaptation is mediated by aldosterone. Increased aldosterone secretion may result in compensatory growth of the adrenal glands. In addition, kidney weights may have increased as the result of increased Na re-absorption. Table 5 shows that in IRA-animals weights of kidneys ( $P > 0.05$ ) and adrenal glands were higher ( $P < 0.001$ ) than those in PVTC- or intact animals. Values for PVTC-animals were similar to those for intact animals. The Na content of digesta from IRA-pigs was lower than that of PVTC-cannulated animals ( $P < 0.001$ ) (Table 9). The low Na content of urine and ileal digesta was accompanied by a low blood

Na concentration in IRA-pigs. The available information indicates that animals fitted with an IRA may have altered Na metabolism. It seems that these animals have a hydro-electrolyte disturbance due to the bypass of the colon.

For K a similar pattern was found (Table 4). The retention and the faecal:urinary K excretion of PVTC-cannulated pigs was comparable to that of intact pigs. There was a considerable change in these variables for anastomosed animals. Thus, IRA-pigs compensate for the absence of K absorption in the large intestine by reducing urinary excretion. The patterns of both Na and K balances demonstrate the important role of the large intestine in mineral absorption and were in agreement with the findings of Clarke *et al.* (1967).

In conclusion, the findings of our experiment suggest that PVTC cannulation has no effect on the mineral balances, organ weights and blood variables measured. Differences between PVTC-pigs and intact pigs with regard to blood urea concentrations suggested an effect of digesta collection on this variable. Further studies are necessary to investigate the effect of digesta collection on urea metabolism in PVTC-pigs. For IRA-pigs the findings suggest that Na supplementation as carried out in the present experiment was insufficient to compensate for the absence of mineral absorption from the large intestine.

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