

Draft genome sequence of *Francisella tularensis* subsp. *holarctica* BD11-00177

Jordy P. M. Coolen¹, Andreas Sjödin², Boulos Maraha³, Gerard F. Hajer⁴, Mats Forsman², Ellen Verspui⁵, Hendrina M.E. Frenay³, Daan W. Notermans⁶, Maaïke C. de Vries⁶, Frans A.G. Reubsaet⁶, Armand Paauw¹, Guus Roeselers¹.

¹TNO, The Netherlands

²Division for CBRN Defence and Security, FOI - Swedish Defence Research Agency, Umeå, Sweden

³Department of Medical Microbiology, Beatrix Hospital, Gorinchem and Albert Schweitzer Hospital, Dordrecht, The Netherlands.

⁴Department of Surgery, Beatrix Hospital, Gorinchem, The Netherlands

⁵Public Health Service Zuid Holland Zuid, Dordrecht, The Netherlands

⁶Diagnostic Laboratory for Infectious Diseases and Perinatal Screening (LIS), Center for Infectious Disease Control, National Institute of Public Health and the Environment (RIVM), Bilthoven, The Netherlands

Correspondence: Guus Roeselers (guus.roeselers@tno.nl)

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Francisella tularensis is a facultative intracellular bacterium in the class *Gammaproteobacteria*. This strain is of interest because it is the etiologic agent of tularemia and a highly virulent category A biothreat agent. Here we describe the draft genome sequence and annotation of *Francisella tularensis* subsp. *holarctica* BD11-00177, isolated from the first case of indigenous tularemia detected in The Netherlands since 1953. Whole genome DNA sequence analysis assigned this isolate to the genomic group B.FTNF002-00, which previously has been exclusively reported from Spain, France, Italy, Switzerland and Germany. Automatic annotation of the 1,813,372 bp draft genome revealed 2,103 protein-coding and 46 RNA genes.

Abbreviations: CDC- United States Centers for Disease Control and Prevention, TNO- Dutch Organization for Applied Scientific Research, FOI- Swedish Defence Research Agency

Introduction

Francisella tularensis is a Gram negative, non-motile, non-spore forming, facultative intracellular bacterium appearing as short rods or coccoid forms [1]. *F. tularensis* is the etiologic agent of tularemia, a zoonotic infection also known as rabbit fever and deer-fly fever. Transmission to humans has been reported by direct contact with infected animals, arthropod bites, inhalation of contaminated dust or ingestion of contaminated food or water. This pathogen is highly infectious as it can cause infection upon inhalation of as few as 10 cells. This extremely low infectious dose makes transmission via aerosols easy, and previous attempts to weaponize this microorganism have led to its recognition as a category A biothreat agent (CDC classification) [2,3]. *F. tularensis* contains

three subspecies that are infectious to humans; the highly virulent *Francisella tularensis* subsp. *tularensis*, which often causes a lethal multi-systemic disease with a fatality rate of up to 30%, the less virulent *Francisella tularensis* subsp. *holarctica* and *Francisella tularensis* subsp. *mediasiatica*, which both seldom cause infectious in humans. Here we present a summary classification together with the description of the draft genome sequence and annotation of *Francisella tularensis* subsp. *holarctica* BD11-00177, that was isolated from a vesicle on the forehead of a 72-year-old male living in The Netherlands. As the patient had not been abroad for years, this was the first documented case of indigenous tularemia in The Netherlands since 1953.

Classification and features

Francisella is the only genus within the family *Francisellaceae* and is a member of the order *Thiotrichales* and the class *Gammaproteobacteria* [4] [Table 1]. Besides *F. tularensis*, the genus *Francisella* includes the species *Francisella halioticida*, *Francisella hispaniensis*, *Francisella noatunensis*, *Francisella novicida*, *Francisella philomiragia*, *Francisella cantonensis* and the mis-

classified *Wolbachia persica* [4,17, Figure 1]. Only rare human infections with *F. hispaniensis* and *F. novicida*, and *F. philomiragia* are described, often caused after nearly drowning [18,19]. *F. tularensis* is capable of infecting hundreds of different vertebrate and invertebrate hosts [20]. The most widely distributed subspecies is *F. tularensis subsp. holarctica*, which is found throughout much of the Northern Hemisphere and is the only subspecies naturally occurring in Europe [21].

Table 1. Classification and general features

MIGS ID	Property	Term	Evidence code ^a
		Domain <i>Bacteria</i>	TAS [5]
		Phylum <i>Proteobacteria</i>	TAS [6]
		Class <i>Gammaproteobacteria</i>	TAS [7,8]
	Current classification	Order <i>Thiotrichales</i>	TAS [7,9]
		Family <i>Francisellaceae</i>	TAS [7-10]
		Genus <i>Francisella</i>	TAS [11-14]
		Species <i>Francisella tularensis</i>	TAS [11,12]
		Subspecies <i>Francisella tularensis holarctica</i>	TAS [15,16]
		Strain BD11-00177	NAS
	Gram stain	negative	TAS [1].
	Cell shape	short rods or coccoid forms	TAS [1].
	Motility	No	TAS [1].
	Sporulation	No	TAS [1].
	Temperature range	Mesophilic	TAS [1].
	Optimum temperature	37	IDA
	Carbon source	Carbohydrates	TAS [1].
	Energy source	Chemoorganotrophic	TAS [1].
	Terminal electron receptor	Facultative anaerobe	TAS [1].
MIGS-6	Habitat	Host	TAS [1].
MIGS-15	Biotic relationship	Obligate host-dependent	TAS [1].
	Host name	Homo sapiens	TAS [1].
	Host taxon ID	9606	TAS [1].
	Host gender	Male	NAS
	Pathogenicity	Pathogen	
MIGS-14	Biosafety Level	3	TAS [2].
MIGS-4	Geographic location	The Netherlands	IDA
MIGS-5	Sample collection time	October 2011	IDA
MIGS-4.1	Latitude	unknown	
MIGS-4.2	Longitude	unknown	
MIGS-4.3	Depth	unknown	
MIGS-4.4	Altitude	unknown	
	Isolate site Human host		IDA
MIGS-4.5	Isolation source	vesicle on the forehead	IDA

^aEvidence codes - IDA: Inferred from Direct Assay; TAS: Traceable Author Statement [i.e., a direct report exists in the literature]; NAS: Non-traceable Author Statement [i.e., not directly observed for the living, isolated sample, but based on a generally accepted property for the species, or anecdotal evidence]. These evidence codes are from the Gene Ontology project.

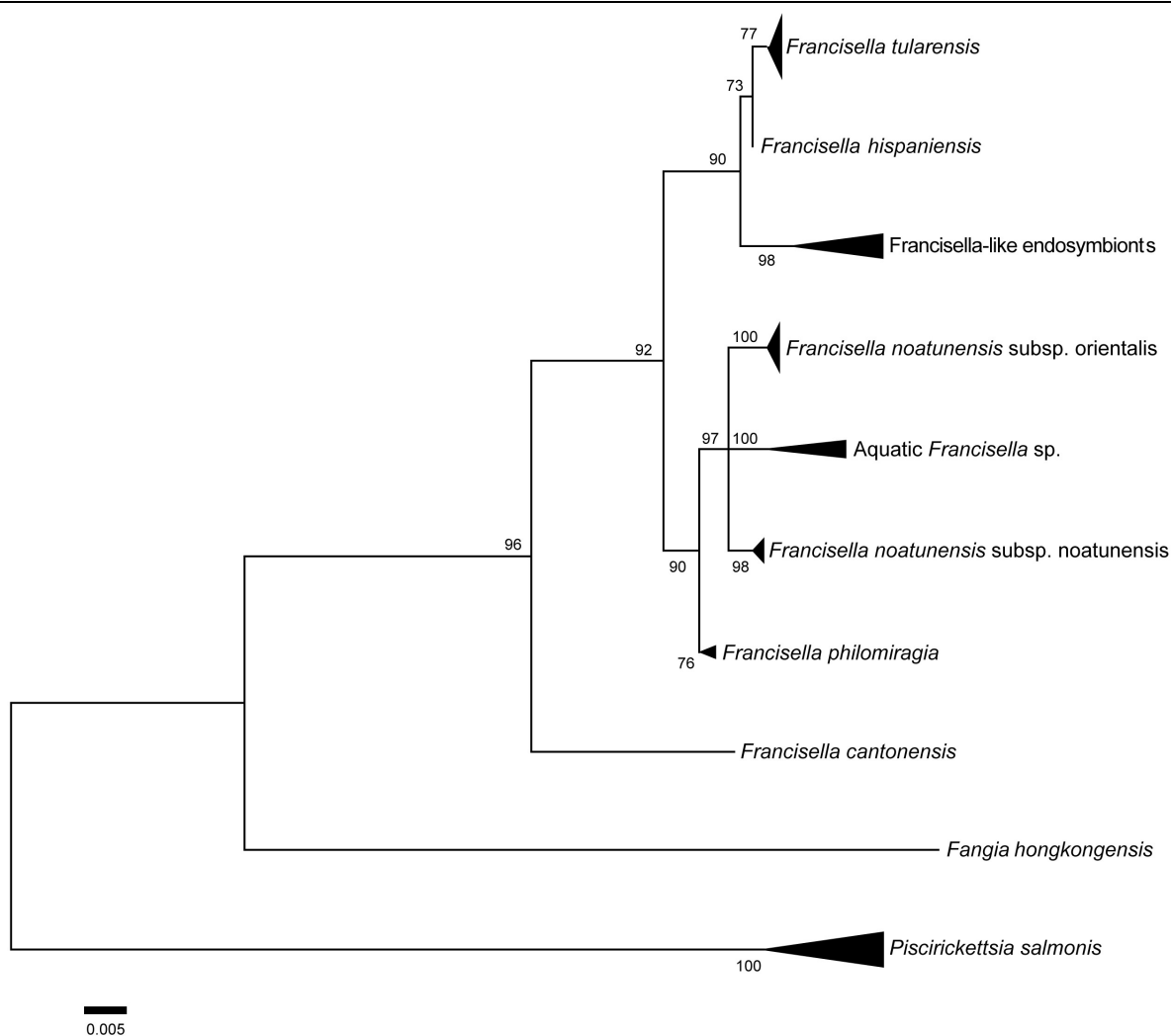


Figure 1. Maximum likelihood tree illustrating the phylogenetic relationships among several members of the genus *Francisella* and members of the order *Thiotrichales* based on full-length 16S rRNA gene sequences.

Genome sequencing information

Genome project history

Strain BD11-00177 was sequenced because of its relevance to biodefense. The draft genome sequence was finished in August 2012. The GenBank accession number for the project is 177784. The genome project is listed in the Genome OnLine Database (GOLD) [22] as project Gi21611. Sequencing was carried out at the Dutch Organization for Applied Scientific Research (TNO) and the Swedish Defense Research Agency (FOI). Initial automatic annotation was performed using the DOE-JGI Microbial Annotation Pipeline (DOE-JGI MAP). Table 2 shows the project information and its association with MIGS 2.0 compliance.

Growth conditions and DNA isolation

For DNA preparation, strain BD11-00177 was grown on 5% sheep blood agar plates for 72 h at

35°C in the presence of 5% CO₂. DNA was extracted using the Qiamp DNA Micro Kit according manufacturers guidelines (Qiagen, Westburg b.v., Leusden, The Netherlands).

Genome sequencing and assembly

Sequencing was performed by the Microbiology and Systems Biology group at TNO and the Division for CBRN Defence and Security at FOI using 454 Roche GS Junior and the Illumina MiSeq platforms. The initial draft assembly yielded 95 large (>1,000 bp) and 86 small (<1,000 bp), non-redundant contigs of 1,813,372 bp by combing 75,245 Roche/454 reads at 23× coverage and 8,289,332 Illumina reads at 690× coverage by hybrid assembly through the Ray Assembler V2.1 [24].

Table 2. Project information

MIGS ID	Property	Term
MIGS-31	Finishing quality	Standard Draft
MIGS-29	Sequencing platforms	Illumina MiSeq, 454 Roche GS Junior
MIGS-31.2	Fold coverage	713×
MIGS-30	Assemblers	Ray Assembler V2.1
MIGS-32	Gene calling method	Prodigal [23]
	GOLD ID	Gi21611
	IMG Taxon ID	1244086
	NCBI PROJECT ID	177784
MIGS-38	Project relevance	Medical, biodefence

Genome annotation

Open Reading Frames (ORFs) were predicted using the Prodigal gene prediction algorithm [23] as part of the DOE-JGI Microbial Annotation Pipeline (DOE-JGI MAP) using default parameters, followed by a round of manual curation. CRISPR elements were predicted using CRT and PILERCR [25]. Predictions from both methods were concatenated. Identification of tRNAs was performed using tRNAscan. Ribosomal RNA genes (5S, 16S, 23S) are predicted using the program RNAmmer [26]. With the exception of tRNA and rRNA, all models from Rfam [27] are used to search the genome sequence. For faster detection, sequences are first compared to a database containing all the ncRNA genes in the Rfam database using BLAST, with a very loose cutoff. Subsequently, sequences that have hits to any genes belonging to an Rfam model are searched using the program INFERNAL [27].

Protein coding genes were compared to protein families (e.g., COGs, Pfam, KEGG) and the proteome of selected “core” genomes, which are publicly available, and the product names were assigned based on the results of these comparisons.

Genome properties

The genome was assembled into 95 large (>1,000 bp) contigs and includes one circular chromosome with a total size of 11,813,372 bp (32.23% GC content). A total of 2,149 genes were predicted, 2,103 of which are protein-coding genes. Of the protein coding genes, 1,592 were assigned to a putative function, with the remaining being annotated as hypothetical proteins. The properties and the statistics of the genome are summarized in Tables 3 and 4.

Table 3. Nucleotide content and gene count levels of the genome

Attribute	Value	% of total ^a
Genome Size (bp)	1,813,372	100.00%
DNA coding region (bp)	1,611,603	88.87%
DNA G+C content (bp)	584,435	32.23%
Total genes ^b	2149	100.00%
RNA genes	46	2.14%
Protein-coding genes	2103	97.86%
Genes in paralog clusters	1262	58.72%
Genes assigned to COGs	1584	73.71%
Protein coding genes connected to KEGG pathways	611	28.43%
not connected to KEGG pathways	1492	69.43%
Genes with signal peptides	111	5.17%
Genes with transmembrane helices	573	26.66%

a) The total is based either on the size of the genome in base pairs or on the total number of protein coding genes in the annotated genome.

Table 4. Number of genes associated with the 25 general COG functional categories

Code	Value	%age ^a	Description
J	152	8.79	Translation
A	1	0.06	RNA processing and modification
K	65	3.76	Transcription
L	198	11.45	Replication, recombination and repair
B	-	-	Chromatin structure and dynamics
D	18	1.04	Cell cycle control, mitosis and meiosis
Y	-	-	Nuclear structure
V	31	1.79	Defense mechanisms
T	24	1.39	Signal transduction mechanisms
M	112	6.47	Cell wall/membrane biogenesis
N	19	1.1	Cell motility
Z	1	0.06	Cytoskeleton
W	-	-	Extracellular structures
U	44	2.54	Intracellular trafficking and secretion
O	66	3.82	Posttranslational modification, protein turnover, chaperones
C	107	6.18	Energy production and conversion
G	118	6.82	Carbohydrate transport and metabolism
E	158	9.13	Amino acid transport and metabolism
F	65	3.76	Nucleotide transport and metabolism
H	96	5.55	Coenzyme transport and metabolism
I	64	3.7	Lipid transport and metabolism
P	71	4.1	Inorganic ion transport and metabolism
Q	37	2.14	Secondary metabolites biosynthesis, transport and catabolism
R	172	9.94	General function prediction only
S	111	6.42	Function unknown
-	565	26.29	Not in COGs

a) The total is based on the total number of protein coding genes in the annotated genome.

Comparisons with other fully sequenced genomes

Comparison of the assembled draft genome sequence of strain BD11-00177 with publicly available *F. tularensis* genome sequences revealed that it clusters in the FTNF002-00 genomic group (B.Br.FTNF002-00 and BIV.FTNF002-00) defined by the FTNF002-00 genome sequence [28-30] within the B.IV clade. The presence of the 1.59 kb RD23 deletion event [31] as well as the 464 bp size of the MLVA marker FtM24 [32], both typical for the FTNF002-00 genomic group, were confirmed *in silico*. Notably, isolates from this genomic group had previously been exclusively reported from Spain, France, Italy, Switzerland and Germany [28,31-35].

A BLAST Ring Image Generator (BRIG) analysis comparing the *F. tularensis* subsp. *holarctica* BD11-00177 genome against the *F. tularensis* subsp. *holarctica* genomes of F92, LVS, and FTNF002-00 revealed that the BD11-00177 draft genome shows

considerable resemblance to FTNF002-00 (Figure 2).

Evolutionary history of *F. tularensis* subspecies *holarctica* strain BD11-00177 was inferred using publicly available whole genome sequences.

The trees in Figure 3 A and B are drawn to scale, with branch lengths in the same units as those of the evolutionary distances used to infer the phylogenetic tree. The evolutionary distances were computed using the *number of differences* method and are in the units of the number of base differences per sequence. The overview of *Francisella* genus involved 52 public genome sequences using *Piscirickettia salmonis* as outgroup (Figure 3A). The detailed analysis involved 14 *F. tularensis* subsp. *holarctica* genome sequences using *F. tularensis* subsp. *tularensis* strain SCHU S4 as outgroup (Figure 3B) [17,30,33,36-41]. All positions containing gaps and missing data were eliminated. There were a total of 1,599,589 positions in the final dataset.

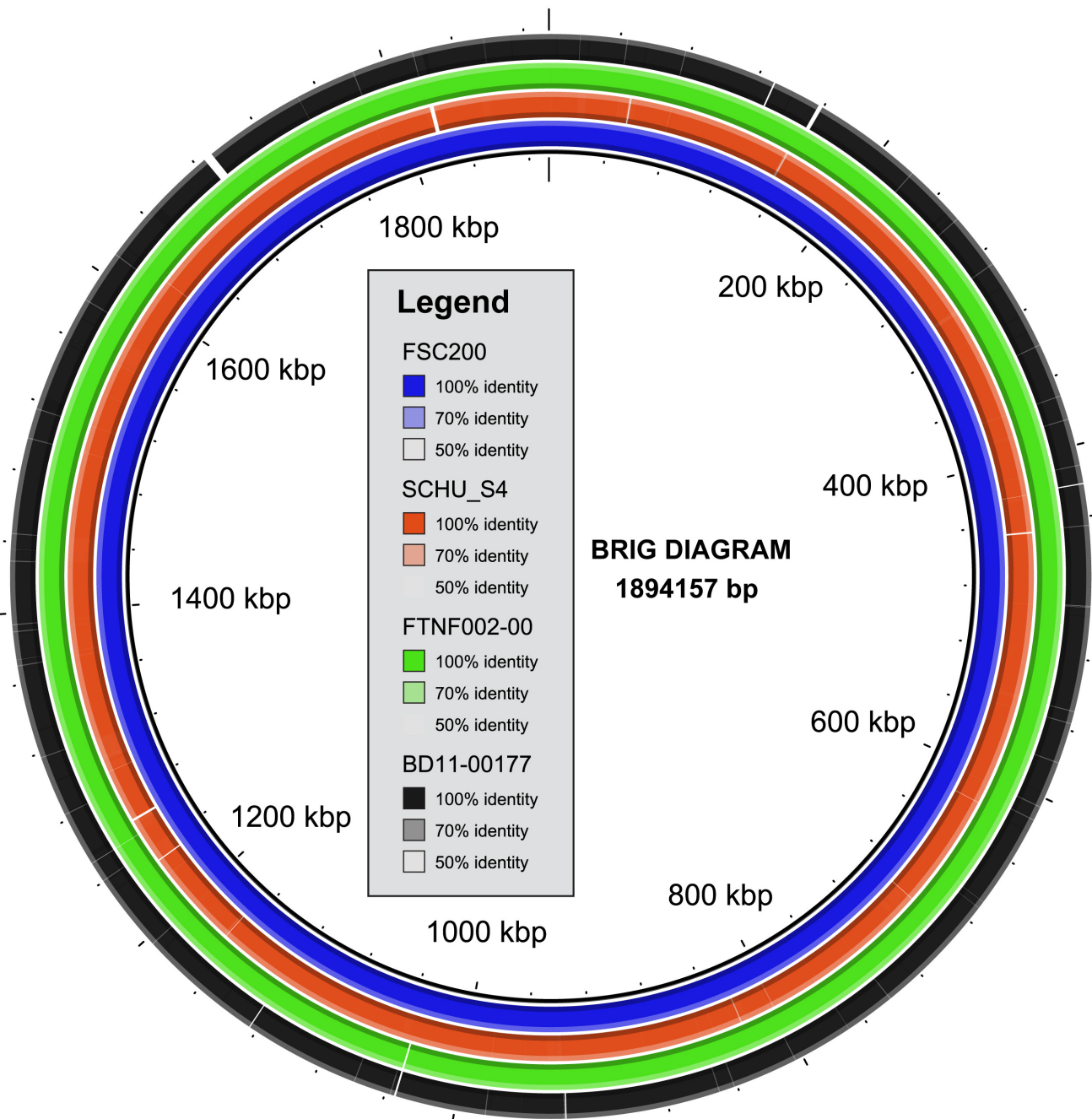


Figure 2. BRIG diagram of the *F. tularensis* subsp. *holarctica* BD11-00177, FTNF002-00 and SCHU S4 genomes using the *F. tularensis* subsp. *holarctica* FSC200 genome as a reference backbone. White regions represent absent genetic regions.

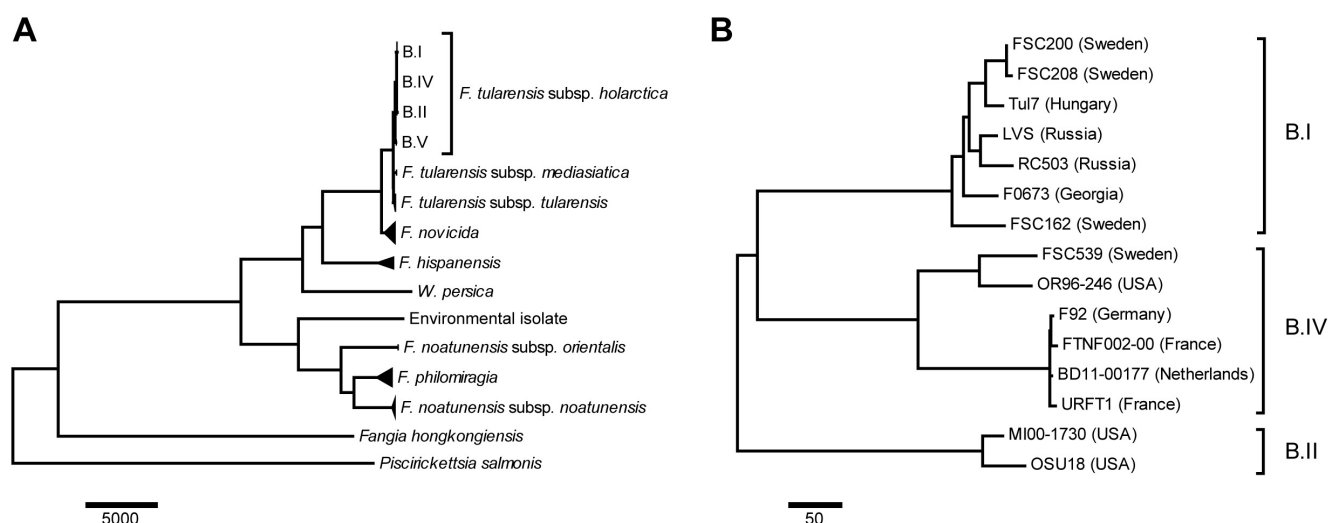


Figure 3. A) Overview of the *Francisella* genus phylogeny based on 52 public whole genome sequences. B) The phylogeny of *F. tularensis subsp. holarctica* strains based on whole genome sequences. The new isolate, BD11-00177 belongs to the FTNF002-00 genomic group inside the B.IV clade.

Conclusion

Here we have presented the draft genome of the first member of FTNF002-00 genomic group of *F. tularensis subspecies holarctica*. As more genetic information of members from this genomic group becomes available, a better understanding of the evolution and biogeography of this pathogen will be gained. This knowledge may help us to understand the epidemiology and potential expansion of the geographical distribution of this genomic group. Despite potential biases associated with discontinuous draft genomes, we would like to

focus on the added value of draft bacterial genome sequencing. Taking advantage of low cost and high-throughput sequencing platforms allows us to probe the vast microbial diversity present in nature and rapidly respond to clinical outbreaks and acute biosecurity hazards. From an evolutionary ecology perspective, increased sequencing efforts allow us to characterize the biogeography of microbial taxa and differentiate between neutral and conserved genome contents.

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References

1. Sjöstedt A. Family XVII. *Francisellaceae*, genus I. *Francisella*. In: Brenner D, Krieg N, Staley J, Garrity G, Boone D, Vos P, et al., editors. *Bergey's Manual of Systematic Bacteriology*. New York: Springer US; 2005. p. 200–210.
2. Rotz LD, Khan AS, Lillibridge SR, Ostroff SM, Hughes JM. Public health assessment of potential biological terrorism agents. *Emerg Infect Dis* 2002; **8**:225-230. [PubMed](http://dx.doi.org/10.3201/eid0802.010164) <http://dx.doi.org/10.3201/eid0802.010164>
3. Forsman M, Sandström G, Sjöstedt A. Analysis of 16S ribosomal DNA sequences of *Francisella* strains and utilization for determination of the phy-
4. Brevik OJ, Ottem KF, Kamaishi T, Watanabe K, Nylund A. *Francisella halioticida* sp. nov., a pathogen of farmed giant abalone (*Haliotis gigantea*) in Japan. *J Appl Microbiol* 2011; **111**:1044-1056. [PubMed](http://dx.doi.org/10.1111/j.1365-2672.2011.05133.x) <http://dx.doi.org/10.1111/j.1365-2672.2011.05133.x>
5. Woese CR, Kandler O, Wheelis ML. Towards a natural system of organisms: proposal for the domains *Archaea*, *Bacteria*, and *Eucarya*. *Proc Natl*

- Acad Sci USA* 1990; **87**:4576-4579. [PubMed](#)
<http://dx.doi.org/10.1073/pnas.87.12.4576>
6. Garrity GM, Bell JA, Lilburn T. Phylum XIV. *Proteobacteria* phyl. nov. In: Garrity GM, Brenner DJ, Krieg NR, Staley JT (eds), *Bergey's Manual of Systematic Bacteriology*, Second Edition, Volume 2, Part B, Springer, New York, 2005, p. 1.
 7. Validation of publication of new names and new combinations previously effectively published outside the IJSEM. List no. 106. *Int J Syst Evol Microbiol* 2005; **55**:2235-2238.
<http://dx.doi.org/10.1099/ijs.0.64108-0>
 8. Garrity GM, Bell JA, Lilburn T. Class III. *Gammaproteobacteria* class. nov. In: Garrity GM, Brenner DJ, Krieg NR, Staley JT (eds), *Bergey's Manual of Systematic Bacteriology*, Second Edition, Volume 2, Part B, Springer, New York, 2005, p. 1.
 9. Garrity GM, Bell JA, Lilburn T. Order V. *Thiotrichales* ord. nov. In: Garrity GM, Brenner DJ, Krieg NR, Staley JT (eds), *Bergey's Manual of Systematic Bacteriology*, Second Edition, Volume 2, Part B, Springer, New York, 2005, p. 131.
 10. Sjöstedt AB. Family III. *Francisellaceae* fam. nov. In: Garrity GM, Brenner DJ, Krieg NR, Staley JT (eds), *Bergey's Manual of Systematic Bacteriology*, Second Edition, Volume 2, Part B, Springer, New York, 2005, p. 199-200.
 11. Skerman VBD, McGowan V, Sneath PHA. Approved Lists of Bacterial Names. *Int J Syst Bacteriol* 1980; **30**:225-420.
<http://dx.doi.org/10.1099/00207713-30-1-225>
 12. Dorofe'ev KA. Classification of the causative agent of tularemia. Symposium Research Works Institute Epidemiology and Microbiology Chita 1947; 1:170-180.
 13. Owen CR. Genus *Francisella* Dorofe'ev 1947, 176. In: Buchanan RE, Gibbons NE (eds), *Bergey's Manual of Determinative Bacteriology*, Eighth Edition, The Williams and Wilkins Co., Baltimore, 1974, p. 283-285.
 14. Huber B, Escudero R, Busse H-J, Seibold E, Scholz HC, Anda P, Kämpfer P, Spletstoesser WD. Description of *Francisella hispaniensis* sp. nov., isolated from human blood, reclassification of *Francisella novicida* (Larson et al. 1955) Olsufiev et al. 1959 as *Francisella tularensis* subsp. *novicida* comb. nov., and emended description of the genus *Francisella*. *Int J Syst Evol Microbiol* 2010; **60**:1887-1896. [PubMed](#)
<http://dx.doi.org/10.1099/ijs.0.015941-0>
 15. Olsufjev NG, Meshcheryakova IS. Subspecific taxonomy of *Francisella tularensis* McCoy and Chapin 1912. *Int J Syst Bacteriol* 1983; **33**:872-874.
<http://dx.doi.org/10.1099/00207713-33-4-872>
 16. Olsufjev NG, Emelyanova OS, Dunaeva TN. Comparative study of strains of *B. tularensis* in the Old and New World and their taxonomy. *J Hyg Epidemiol Microbiol Immunol* 1959; **3**:138-149.
[PubMed](#)
 17. Sjödin A, Svensson K, Ohrman C, Ahlinder J, Lindgren P, Duodu S, Johansson A, Colquhoun DJ, Larsson P, Forsman M. Genome characterisation of the genus *Francisella* reveals insight into similar evolutionary paths in pathogens of mammals and fish. *BMC Genomics* 2012; **13**:268. [PubMed](#)
<http://dx.doi.org/10.1186/1471-2164-13-268>
 18. Wenger JD, Hollis DG, Weaver RE, Baker CN, Brown GR, Brenner DJ, Broome CV. Infection caused by *Francisella philomiragia* (formerly *Yersinia philomiragia*). A newly recognized human pathogen. *Ann Intern Med* 1989; **110**:888-892.
[PubMed](#) <http://dx.doi.org/10.7326/0003-4819-110-11-888>
 19. Larson CL, Wicht W, Jellison WL. A new organism resembling *P. tularensis* isolated from water. *Public Health Rep* 1955; **70**:253-258. [PubMed](#)
<http://dx.doi.org/10.2307/4589039>
 20. Mörner T, Addison E. Tularemia. In: Williams ES, Barker IK, editors. *Infectious diseases of wild animals*. 3rd ed. Ames: Iowa State University; 2001. p. 303-312.
 21. Keim P, Johansson A, Wagner DM. Molecular epidemiology, evolution, and ecology of *Francisella*. *Ann N Y Acad Sci* 2007; **1105**:30-66. [PubMed](#)
<http://dx.doi.org/10.1196/annals.1409.011>
 22. Liolios K, Chen IM, Mavromatis K, Tavernarakis N, Hugenholtz P, Markowitz VM, Kyrpides NC. The Genomes On Line Database (GOLD) in 2009: status of genomic and metagenomic projects and their associated metadata. *Nucleic Acids Res* 2010; **38**(Database issue):D346-D354. [PubMed](#)
<http://dx.doi.org/10.1093/nar/gkp848>
 23. Hyatt D, Chen GL, Locascio PF, Land ML, Larimer FW, Hauser LJ. Prodigal: prokaryotic gene recognition and translation initiation site identification. *BMC Bioinformatics* 2010; **11**:119. [PubMed](#)
<http://dx.doi.org/10.1186/1471-2105-11-119>
 24. Boisvert S, Laviolette F, Corbeil J. Ray: simultaneous assembly of reads from a mix of high-throughput sequencing technologies. *Journal of computational biology: a journal of computational molecular cell biology*. 2010 Nov; **17**(11):1519-1533.

25. Edgar RC. PILER-CR: fast and accurate identification of CRISPR repeats. *BMC Bioinformatics* 2007; **8**:18. [PubMed http://dx.doi.org/10.1186/1471-2105-8-18](http://dx.doi.org/10.1186/1471-2105-8-18)
26. Lagesen K, Hallin P, Rødland EA, Staerfeldt HH, Rognes T, Ussery DW. RNAMmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res* 2007; **35**:3100-3108. [PubMed http://dx.doi.org/10.1093/nar/gkm160](http://dx.doi.org/10.1093/nar/gkm160)
27. Griffiths-Jones S, Moxon S, Marshall M, Khanna A, Eddy SR, Bateman A. Rfam: annotating non-coding RNAs in complete genomes. *Nucleic Acids Res* 2004; **33**(Database issue):D121-D124. [PubMed http://dx.doi.org/10.1093/nar/gki081](http://dx.doi.org/10.1093/nar/gki081)
28. Vogler AJ, Birdsell D, Price LB, Bowers JR, Beckstrom-Sternberg SM, Auerbach RK, Beckstrom-Sternberg JS, Johansson A, Clare A, Buchhagen JL, et al. Phylogeography of *Francisella tularensis*: global expansion of a highly fit clone. *J Bacteriol* 2009; **191**:2474-2484. [PubMed http://dx.doi.org/10.1128/JB.01786-08](http://dx.doi.org/10.1128/JB.01786-08)
29. Svensson K, Granberg M, Karlsson L, Neubauerova V, Forsman M, Johansson A. A real-time PCR array for hierarchical identification of *Francisella* isolates. *PLoS ONE* 2009; **4**:e8360. [PubMed http://dx.doi.org/10.1371/journal.pone.0008360](http://dx.doi.org/10.1371/journal.pone.0008360)
30. Barabote RD, Xie G, Brettin TS, Hinrichs SH, Fey PD, Jay JJ, Engle JL, Godbole SD, Noronha JM, Scheuermann RH. Complete genome sequence of *Francisella tularensis* subspecies holarctica FTNF002-00. *PLoS ONE* 2009; **4**:e7041. [PubMed http://dx.doi.org/10.1371/journal.pone.0007041](http://dx.doi.org/10.1371/journal.pone.0007041)
31. Dempsey MP, Dobson M, Zhang C, Zhang M, Lion C, Gutiérrez-Martín CB. Genomic deletion marking an emerging subclone of *Francisella tularensis* subsp. holarctica in France and the Iberian Peninsula. *Appl Environ Microbiol* 2007; **73**:7465-7470. [PubMed http://dx.doi.org/10.1128/AEM.00646-07](http://dx.doi.org/10.1128/AEM.00646-07)
32. Pilo P, Johansson A, Frey J. Identification of *Francisella tularensis* cluster in central and western Europe. *Emerg Infect Dis* 2009; **15**:2049-2051. [PubMed http://dx.doi.org/10.3201/eid1512.080805](http://dx.doi.org/10.3201/eid1512.080805)
33. Gyuranecz M, Birdsell DN, Splettstoesser W, Seibold E, Beckstrom-Sternberg SM, Makrai L, Fodor L, Fabbri M, Vicari N, Johansson A, et al. Phylogeography of *Francisella tularensis* subsp. holarctica, Europe. *Emerg Infect Dis* 2012; **18**:290-293. [PubMed http://dx.doi.org/10.3201/eid1802.111305](http://dx.doi.org/10.3201/eid1802.111305)
34. Vogler AJ, Birdsell DN, Lee J, Vaissaire J, Doujet CL, Lapalus M, Wagner DM, Keim P. Phylogeography of *Francisella tularensis* ssp. holarctica in France. *Lett Appl Microbiol* 2011; **52**:177-180. [PubMed http://dx.doi.org/10.1111/j.1472-765X.2010.02977.x](http://dx.doi.org/10.1111/j.1472-765X.2010.02977.x)
35. Gehringer H, Schacht E, Maylaender N, Zeman E, Kaysser P, Oehme R, et al. Presence of an emerging subclone of *Francisella tularensis* holarctica in Ixodes ricinus ticks from south-western Germany. *Ticks and tick-borne diseases*. 2013 Feb;4(1-2):93-100.
36. Svensson K, Sjödin A, Byström M, Granberg M, Brittnacher MJ, Rohmer L, Jacobs MA, Sims-Day EH, Levy R, Zhou Y, et al. Genome sequence of *Francisella tularensis* subspecies holarctica strain FSC200, isolated from a child with tularemia. *J Bacteriol* 2012; **194**:6965-6966. [PubMed http://dx.doi.org/10.1128/JB.01040-12](http://dx.doi.org/10.1128/JB.01040-12)
37. Petrosino JF, Xiang Q, Karpathy SE, Jiang H, Yerrapragada S, Liu Y, Gioia J, Hemphill L, Gonzalez A, Raghavan TM, et al. Chromosome rearrangement and diversification of *Francisella tularensis* revealed by the type B (OSU18) genome sequence. *J Bacteriol* 2006; **188**:6977-6985. [PubMed http://dx.doi.org/10.1128/JB.00506-06](http://dx.doi.org/10.1128/JB.00506-06)
38. La Scola B, Elkarkouri K, Li W, Wahab T, Fournous G, Rolain JM, Biswas S, Drancourt M, Robert C, Audic S, et al. Rapid comparative genomic analysis for clinical microbiology: the *Francisella tularensis* paradigm. *Genome Res* 2008; **18**:742-750. [PubMed http://dx.doi.org/10.1101/gr.071266.107](http://dx.doi.org/10.1101/gr.071266.107)
39. Chanturia G, Birdsell DN, Kekelidze M, Zhgenti E, Babuadze G, Tsertsvadze N, Tsanova S, Imnadze P, Beckstrom-Sternberg SM, Beckstrom-Sternberg JS. Phylogeography of *Francisella tularensis* subspecies holarctica from the country of Georgia. *BMC Microbiol* 2011; **11**:139. [PubMed http://dx.doi.org/10.1186/1471-2180-11-139](http://dx.doi.org/10.1186/1471-2180-11-139)
40. Karlsson E, Svensson K, Lindgren P, Byström M, Sjödin A, Forsman M, Johansson A. The phylogeographic pattern of *Francisella tularensis* in Sweden indicates a Scandinavian origin of *EuroSiberian tularaemia*. *Environ Microbiol* 2013; **15**:634-645. [PubMed http://dx.doi.org/10.1111/1462-2920.12052](http://dx.doi.org/10.1111/1462-2920.12052)
41. Antwerpen MH, Schacht E, Kaysser P, Splettstoesser WD. Complete Genome Sequence of a *Francisella tularensis* subsp. holarctica Strain from Germany Causing Lethal Infection in Common Marmosets. *Genome announcements*. 2013 Jan;1(1).